

The Response of Spring Wheat Cultivars to Arbuscular Mycorrhizal Colonization under Salinity Stresses

Daishu Yi¹, Timothy Schwinghamer¹, Yolande Dalpé², Jaswinder Singh¹ & Shahrokh Khanizadeh²

¹Department of Plant Science, McGill University, 21111 Lakeshore Road, Sainte-Anne-De-Bellevue, Quebec, Canada

²Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, K.W. Neatby Building, 960 Carling Avenue, Ottawa, Ontario, Canada

Correspondence: Shahrokh Khanizadeh, Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, K.W. Neatby Building, 960 Carling Avenue, Ottawa, Ontario, Canada. E-mail: Shahrokh.khanizadeh@agr.gc.ca, <http://khanizadeh.info>

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Abstract

Wheat is an important crop, playing inevitable roles in human life, ranging from major food resource to raw material for biofuel. However, due to the dramatically reduced available arable areas and increasingly severe abiotic and biotic stresses, wheat production nowadays faces extreme challenges. Many approaches have been explored to increase wheat yield including development of new cultivars. One of the most promising approaches is the application of the naturally existent arbuscular mycorrhiza (AM), a mutualistic symbiosis originated over 400 million years ago. AM have long been known to form mutualistic symbiosis with various plants to enhance yield production and to improve stress tolerance, especially drought and salinity. But the benefits vary among AM strains and plant species. Therefore, the objective of the study was to investigate the influence of four AM strains colonized on four selected spring wheat varieties under three salt concentrations (0, 50, 100 mmol/L). The results demonstrated that wheat inoculated with arbuscular mycorrhizal strains *Funneliformis mosseae* and *Rhizoglyphus irregularis* mitigated yield losses caused by increased salinity stresses as well as strengthened root growth in comparison with non-inoculated plant controls. Salinity stress, however, had non-significant negative effects on most variables, except for grain yield, root surface area and root dry weight, in which a significant decrease was observed in root surface area and root dry weight with the increasing of saline concentration.

Keywords: arbuscular mycorrhizae, root morphology, salinity stress, wheat (spring)

1. Introduction

Wheat (*Triticum aestivum* L.) ranks among the top cereal crops and is grown in diverse agricultural ecosystems around the world. In 2013, with a global production of over 0.71 billion t, Wheat occupied the third position of most produced crop over the world, after 0.74 billion t of rice and over 1 billion t of maize (Food and Agriculture Organization of the United Nations, 2013). Wheat is planted on more than 17% of the global crop acreage and it successfully feeds over two-fifths of the population worldwide (Gupta *et al.*, 2008). Today, however, numerous severe stresses, both biotic and abiotic, negatively impede the global wheat production: one of the most severe concerns is soil salinization (Günel *et al.*, 2015). Many approaches have been applied to alleviate yield loss caused by soil salinization, including the exploration of novel stress-tolerant cultivars through modern breeding methods (Mozafar and Goodin, 1986), the optimization of cultivation systems to reduce the negative effects of salinization (Sun *et al.*, 2010), as well as the application of naturally existing beneficial symbioses (Beltrano and Ronco, 2008). One of the most promising approaches is the exploration and use of arbuscular mycorrhizae, a mutualistic symbiosis that could protect plants from salinity damages.

With a history of more than 400 million years, arbuscular mycorrhizae are considered to be one of the world's oldest and most prevalent symbioses formed between plants and fungi (Parniske, 2008; Pirozynski and Malloch, 1975; Remy *et al.*, 1994). Previous studies have reported that more than 85% of the documented plant species are putative hosts of arbuscular mycorrhizal fungi (AMF) and that those species benefit from the symbiosis in various ways, including increased absorption of water and mineral nutrients (especially phosphorus, the crucial mineral for plant growth and development) and improved biotic and abiotic stress tolerance through multiple

complex mechanisms (Daei *et al.*, 2009; Harley and Smith, 1997; Marschner and Dell, 1994; Pozo *et al.*, 2010; Wang and Qiu, 2006). An increase in salinity tolerance was observed in numerous mycorrhizal colonized plants, including crops such as barley and wheat (Rad *et al.*, 2009; Talaat and Shawky, 2014). Nevertheless, such benefits vary depending on the AMF strains selected and the host plant species involved (Gong *et al.*, 2013; Zou and Wu, 2011). Therefore, the objective of this project was to evaluate the response of four selected spring wheat genotypes to inoculation with four arbuscular mycorrhizal strains under three salt concentrations.

2. Materials and Methods

2.1 Experimental Materials

2.1.1 Wheat Cultivars

Four spring wheat cultivars, namely FL62R1, Scotia, Snowbird, and 13NQW1265, were analyzed in the current study (Table 1). In addition to plant biomass and root morphology, the relationship between grain colour and antioxidant components and their responses to wheat mycorrhization were also of great interest. Therefore, the four spring wheat cultivars chosen for our study were mainly because of their respective grain colours (red, white or purple). On one hand, changes in plant biomass and root morphology were measured and analyzed. On the other hand, the harvested grains were further processed for the analysis of grain antioxidant components to determine differences in the antioxidant capacity of coloured wheat with and without arbuscular mycorrhizal inoculation.

Table 1. Wheat cultivars, arbuscular mycorrhizal (AM) strains, and salinity treatments of the experimental design

Spring wheat cultivars		
Wheat cultivar	Grain colour	Seeds per pot
FL62R1	Red	5
Scotia	Red	5
Snowbird	White	5
13NQW1265	Purple	5
AM strains		
AM strain ^{a)}	DAOM ^{b)}	Number of propagules per pot
<i>Funneliformis +mosseae</i>	198274	50
<i>Funneliformis caledonius</i>	242686	50
<i>Rhizoglosum irregulare</i>	240442	50
<i>Myke</i>	197198	50
Salinity treatments		
Saline solution (mmolL ⁻¹)	grams of NaCl (L ⁻¹)	Volume of saline solution per pot (L)
0	0	0.1
50	2.925	0.1
100	5.85	0.1

^{a)} The arbuscular mycorrhizal strains *Rhizoglosum irregulare* (DAOM 240442) and *Myke* (DAOM 197198) originate from the same mycorrhizal species but are different strains.

^{b)} DAOM refers to Canadian National Mycological Herbarium.

--Arbuscular mycorrhizal strains (AMF strains)

Four AMF strains, namely *Funneliformis mosseae* (DAOM 198274), *F. caledonium* (DAOM 242686), and two *Rhizoglosum irregulare* strains (DAOM 240442) and the commercial product *Myke* (DAOM 197198). The AMF strains were obtained from the Glomeromycota in vitro and in vivo Collection, Ottawa Research and Development Centre, Agriculture and Agri-Food Canada.

--Salinity treatments

Saline solutions were made with sodium chloride (NaCl), and three salinity levels (0, 50, or 100 mmolL⁻¹) were used.

2.2 Methods

A greenhouse experiment was conducted under controlled condition with a 16-h photoperiod and a 22 °C/20 °C day/night temperature regime. For each of the four wheat cultivars, five seeds inoculated with one AMF strain at

a rate of 50 spores were planted in each 15.25cm (6-inch) pot, with equal distances between the seeds. Non-inoculated seeds were also planted for each cultivar as controls. Each inoculant was provided in the form of a specific matrix containing relevant AMF spores. The non-mycorrhizal controls were treated with mycorrhizal-free soil instead of mycorrhizal-spore-containing matrix. A total of 360 pots were used, with 60 pots per replicate and 6 replicates, arranged according to a randomized complete block design. The AGRO MIX G10 (AF) soil was used as Substrate matrix to guarantee no extra mycorrhizal fungi in the soil. Constant irrigation was provided throughout the growth period but was stopped on the days when the saline treatments were applied. The plants were fertilized with 100 ml 20-20-20 N-K-P (1 g kg⁻¹) per pot every two weeks.

Salinity treatment with 100 ml of NaCl solution (0, 50, or 100 mmolL⁻¹) per pot was initialled on the fifth week of growth and continued at 3-d intervals for six weeks until harvest. Data on grain yield, root length, root surface area, total root volume, root fresh weight, and root dry weight were collected and measured. Root morphological parameters were collected with *WinRHIZO Pro* Regent Instruments software and root dry weight was then measured after the roots had been dried in an oven at 60 °C for 3 d. The data were analyzed using PROC GLIMMIX (Generalized Linear Mixed Models) of the SAS software package.

3. Results

3.1 Plant Biomass

Grain yield, root fresh weight and root dry weight were measured and analyzed as indicators of plant biomass.

--Grain yield

Grain yield was significantly affected by the interaction between mycorrhizal strains and salinity treatments with a *P*-value of 0.0121 (Table 1). Specifically, salinity levels had a significantly negative effect on yield production in the non-mycorrhizal controls (*P* = 0.0205) and in the wheat cultivars inoculated with *F. caledonius* (*P* = 0.0027), which exhibited a significant yield decrease when treated with the NaCl solution at 100 mmolL⁻¹ in comparison with the non-salt-treated controls. There were, however, no significant changes in grain yield between the salt and non-salt treatments when the wheat had been inoculated with the other AMF strains, indicating that colonization with *F. mosseae*, *R. irregularare*, and *Myke* benefited the host wheat by efficiently mitigating saline-induced yield decreases.

Table 2. Effect of interaction between mycorrhizal strains and salinity levels on grain yield, sliced by mycorrhizal strain ($\alpha = 0.05$)

AM strains	Salinity levels (mmolL ⁻¹)	<i>P</i> -value	Grain yield (estimates) ^{a)}
<i>Funneliformis mosseae</i>	0	<i>P</i> = 0.5024	0.6534a ^{b)}
	50		0.7682a
	100		0.6935a
<i>Funneliformis caledonius</i>	0	<i>P</i> = 0.0027	0.8746a
	50		0.8243a
	100		0.5577b
<i>Rhizoglo mus irregularare</i>	0	<i>P</i> = 0.3737	0.7011a
	50		0.8326a
	100		0.7961a
<i>Myke</i>	0	<i>P</i> = 0.2865	0.7156a
	50		0.5693a
	100		0.6765a
Control	0	<i>P</i> = 0.0205	0.8390a
	50		0.6743ab
	100		0.5667b
Mycorrhizal strains * salinity levels ^{c)}			<i>P</i> = 0.0121

^{a)} Grain yield is presented as estimates, because there was difficulty with the transformation back into means.

^{b)} Estimates followed by the same letters are not significantly different with 95% confidence limits.

^{c)} A two-way-interaction between mycorrhizal strains and salinity levels was found.

--Root fresh weight

Root fresh weight was significantly influenced by the wheat-mycorrhiza interaction, with a *P*-value of 0.0137 (Table 3). Salt treatment, however, had no significant effect on root fresh weight (*P* = 0.0548). Specifically, the

wheat cultivars Snowbird and 13NQW1265 responded positively to inoculation with *R. irregulare*, resulting in significantly heavier root fresh weights in comparison with the non-inoculated controls, as shown in Table 3. Nevertheless, inoculation of either Snowbird or 13NQW1265 with the other AMF, namely, *F. mosseae*, *F. caledonius*, and the commercial strain *Myke*, generally had no significant effects on root fresh weight, presenting as no significant differences in root fresh weight between the inoculated plants and the non-inoculated controls. The only exception was the *F. mosseae*-inoculated Snowbird, which responded positively to that AMF and produced a significantly heavier root fresh weight than non-inoculated Snowbird did. FL62R1 and Scotia did not exhibit any significant change in root fresh weight in response to plant mycorrhization. Furthermore, it was noticed that even though the non-commercial mycorrhizal strain *R. irregulare* and the commercial strain *Myke* belonged to the same AMF species, the performance of *Myke* was apparently worse than that of *R. irregulare*, particularly in the cultivars FL62R1 and Scotia. Root fresh weight was significantly heavier in the *R. irregulare*-inoculated FL62R1 than in the *Myke*-inoculated FL62R1. Similarly, heavier root fresh weight was observed in the Scotia wheat inoculated with *R. irregulare* than in the same cultivar inoculated with *Myke*. In addition, the wheat cultivar FL62R1 (whether inoculated or non-inoculated) generally had higher levels of root fresh weight, whereas Snowbird (whether inoculated or non-inoculated) had the lower root fresh weight. These findings show that the wheat cultivar FL62R1 was the best option among these four experimental spring wheat genotypes in terms of producing stronger root systems.

Table 3. Effect of interaction between mycorrhizal strains and wheat cultivars on root fresh weight, sliced by wheat cultivar ($\alpha = 0.05$)

Wheat cultivars	Mycorrhizal strains	P-value	Root fresh weight (means) ^{a)}
FL62R1	<i>Funneliformis mosseae</i>	$P = 0.0425$	2.4593ab ^{b)}
	<i>Funneliformis caledonius</i>		2.6365ab
	<i>Rhizogloium irregulare</i>		3.0955a
	<i>Myke</i>		2.1972b
	Control		2.6191ab
Scotia	<i>Funneliformis mosseae</i>	$P = 0.0179$	2.1776ab
	<i>Funneliformis caledonius</i>		2.1834ab
	<i>Rhizogloium irregulare</i>		2.2746a
	<i>Myke</i>		1.7020b
	Control		2.1886ab
Snowbird	<i>Funneliformis mosseae</i>	$P = 0.0016$	1.8465a
	<i>Funneliformis caledonius</i>		1.3465ab
	<i>Rhizogloium irregulare</i>		1.8073a
	<i>Myke</i>		1.6429ab
	Control		1.2751b
13NQW1265	<i>Funneliformis mosseae</i>	$P = 0.0108$	2.2588ab
	<i>Funneliformis caledonius</i>		2.0449ab
	<i>Rhizogloium irregulare</i>		2.4982a
	<i>Myke</i>		1.9780ab
	Control		1.9036b
Mycorrhizal strains * wheat cultivars ^{c)}			$P = 0.0137$

^{a)} Root fresh weight is presented as means.

^{b)} Means followed by the same letters are not significantly different with 95% confidence limits.

^{c)} A two-way-interaction between mycorrhizal strains and wheat cultivars was found.

--Root dry weight

Root dry weight was significantly affected by AMF strain ($P < 0.0001$), and salinity treatment ($P = 0.0234$) independently (Tables 4 and 5). A significant difference was observed between cultivars ($P < 0.0001$). In terms of the evaluation of wheat cultivars, FL62R1 had the heaviest root dry weight, whether inoculated with mycorrhizal fungi or not. The cultivar Snowbird had the lowest root dry weight, whereas Scotia and 13NQW1265 had relatively moderate root dry weights, which were lower than that of wheat FL62R1 but higher than that of Snowbird. Multiple comparisons between the wheat plants colonized by the mycorrhizal strains *F. mosseae*, *F. caledonius*, and *R. irregulare* and the non-mycorrhizal controls indicate that mycorrhizal inoculation significantly improved root dry weight. The only exception was inoculation with the commercial AMF strain

Myke, given that *Myke* colonization had no significant effects on the wheat cultivars in comparison with the non-inoculated controls.

Table 4. Back-transformed means^{a)} for root morphology and root dry weight with Bonferroni grouping ($\alpha = 0.05$)

	Root length ^{b)} (mm)	Surface area (mm*mm)	Root volume (mm*mm*mm)	Root dry weight (grams)
Wheat cultivars				
FL62R1	280.07a ^{c)}	84.7314a	2.0491a	0.5401a
Scotia	287.98a	79.0213ab	1.725ab	0.4618b
Snowbird	239.03b	58.6932c	1.2598c	0.3544c
13NQW1265	304.79a	75.2293b	1.438bc	0.437b
<i>P</i> -value	<i>P</i> < 0.0001	<i>P</i> < 0.0001	<i>P</i> < 0.0001	<i>P</i> < 0.0001
Mycorrhizal strains^{d)}				
<i>F. mosseae</i>	301.60a	81.2535ab	1.744ab	0.4732a
<i>F. caledonius</i>	252.54bc	64.8801cd	1.3384c	0.4556a
<i>R. irregularare</i>	320.13a	88.5825a	1.9599a	0.5121a
<i>Myke</i>	237.81c	63.7823d	1.3682bc	0.392b
Control	280.57ab	73.19bc	1.696abc	0.3959b
<i>P</i> -value	<i>P</i> < 0.0001	<i>P</i> < 0.0001	<i>P</i> < 0.0001	<i>P</i> < 0.0001

^{a)} The four variables were analyzed based on their different distributions, and thus the means here are back-transformed means.

^{b)} No two-way interactions were found for these four variables; they were affected by wheat cultivar and mycorrhizal strain independently.

^{c)} Means followed by the same letters are not significantly different with 95% confidence limits.

^{d)} *F. mosseae* = *Funneliformis mosseae*; *F. caledonius* = *Funneliformis caledonius*; *R. irregularare* = *Rhizoglyphus irregularare*.

Table 5. Back-transformed means^{a)} for root surface area and root dry weight under salinity effects with Bonferroni grouping ($\alpha = 0.05$)

Salinity levels (mmolL ⁻¹)	Surface area (mm*mm) ^{b)}	Root dry weight (grams)
0	76.9581a ^{c)}	0.4615a
50	74.4342ab	0.4493ab
100	69.9915b	0.4203b
<i>P</i> -value	<i>P</i> = 0.0200	<i>P</i> = 0.0234

^{a)} The two variables were analyzed based on their different distributions, and thus the means here are back-transformed means.

^{b)} No two-way interactions were found for these variables, and therefore they were affected by salinity level independently.

^{c)} Means followed by the same letters are not significantly different with 95% confidence limits.

3.2 Root Morphology

Total root length, root surface area, and total root volume were measured as representatives of root morphological variables. All the three root architecture parameters depended to the wheat cultivar and the mycorrhizal strain, independently (Table 4). The two-way interaction was not statistically significant at the $P=0.05$ level. Generally, the results indicate that inoculation with *R. irregularare* benefited the host wheat by strengthening root development, which was quantified by larger total root volume, longer root length, and greater root surface area. Root surface area was taken as an example here. A comparison between the *R. irregularare*-inoculated wheat plants and the non-mycorrhizal controls indicates that colonization by *R. irregularare* fungus appeared to positively promote root growth and development, contributing to significantly larger root surface area. From the perspective of wheat cultivar selection, the results demonstrate that the wheat cultivars exhibited different root morphology responses. Generally, cultivar FL62R1 and Scotia had the best performance for all variables, resulting in the longest root length, largest root surface area, and largest root volume, whereas the commonly used cultivar Snowbird had the poorest performance, with the lowest values for all the root morphology variables measured.

3.3 Salinity Effects

The results show that the salinity had a non-significant effect on root length, total root volume, root fresh weight.

In addition to negatively affecting grain yield through an interaction with mycorrhizal strain as discussed above, the salinity treatments also had significantly negative effects on root surface area and root dry weight. Specifically, root surface area and root dry weight decreased significantly under the highest-concentration of salt solution (100 mmolL^{-1}) in comparison with the non-salt control in all wheat-mycorrhizal combinations, and the decrease could be well-described by linear regression. These findings suggest that even the highest level of saline stress applied in this study may be far below the salt sensitivity of wheat cultivars, with the result that the effects of salt were non-significant on most of the variables in this study.

4. Discussion and Conclusion

4.1 Plant Biomass and Root Morphology

Taken together, the above findings demonstrate that the mycorrhizal strains *F. mosseae* and *R. irregulare* improved the performance of the experimental wheat cultivars in terms of both plant biomass (grain yield and root biomass) and root morphology among the four selected AMF strains. Wheat colonized by *F. mosseae* or *R. irregulare* showed significant advantages in terms of both the mitigation of saline-induced yield loss and the promotion of root development, with heavier root biomass and optimized root architecture in comparison with the non-inoculated controls. However, the performance of mycorrhizal strains *F. caledonius* and *Myke* varied among the variables.

With respect to plant biomass, the results illustrate that wheat inoculated with *F. mosseae* and *R. irregulare* experienced lower yield loss under salinity stress and produced heavier root biomass. These effects may be due in part to the integrated effects, which include: increased uptake of phosphorus (which functions as the crucial nutrient and material base for plant development and growth), enhanced water absorption and water-use efficiency (which allow the plant to fight against salinity-induced physiological water shortages), and more efficient photosynthesis (which provides more carbohydrates and improves the transportation rate between the plant and the fungus in the mycorrhizal symbiosis) (Collaet *et al.*, 2008; Sheng *et al.*, 2008; Talaat and Shawky, 2013).

It has been widely reported that mycorrhizae help plants to improve their tolerance against salinity stresses through multiple mechanisms, including changes in root morphology and the enhancement of water and nutrient uptake. The comprehensive effects therefore improve plant vigour and provide hosts with heavier root biomass in comparison with non-mycorrhizal controls under saline conditions (Evelin *et al.*, 2009; Porcel *et al.*, 2012). In consequence, evaluations on root biomass and yield production are insufficient to provide enough information for a full understanding of the influence of plant mycorrhization. As previously reported by Iman *et al.*, (2006), root morphology may change without significant differences in root biomass. Therefore, in addition to root biomass, root morphological parameters must also be evaluated, as was done in this study.

Optimized root architecture was observed in the wheat inoculated with *R. irregulare*, as indicated by larger root surface area, longer root length, and larger total root volume. Particularly, the *R. irregulare* -inoculated wheat had a significantly larger root surface area in comparison with the non-inoculated controls and thus provided larger contact areas with the soil matrix, allowing the plants to absorb mineral nutrients and water more efficiently, an absolute advantage in stress conditions. Similarly, roots with larger total root volume and longer root length were also of great value in terms of nutrient absorption, by exposing roots to a larger area of soil matrix and by drilling deeper into the soil for more available nutrition (Jones *et al.*, 1989). The enhanced uptake of nutrients, especially phosphorus ions, played a crucial role in the promotion of root development and contributed to the increased root fresh weight and root dry weight in this study. Therefore, it is reasonable to conclude that colonization by the mycorrhizal fungus *R. irregulare* significantly optimized root architecture, which was the structural foundation for increased nutrient uptake capacity, and that the increased water and mineral absorption contributed to heavier root biomass. However, the performance of other AMF strains was not as stable and positive as that of *R. irregulare*.

A comparison between the wheat cultivars shows that FL62R1 performed the best in most cases, with significantly heavier root biomass and better optimized root architecture. In contrast, the Snow bird wheat responded the poorest to mycorrhizal colonization. These findings suggest that in terms of better response to mycorrhization and stronger stress tolerance capacity, the wheat cultivar FL62R1 is more suitable than the commercially used Snowbird wheat.

4.2 Salinity Treatment

Salinity had significant negative effects on grain yield, root surface area, and root dry weight at the highest experimental salinity level (100 mmolL^{-1}). Regarding the other experimental variables, the salinity levels used in

this study were too low to induce a detectable stress response on wheat cultivars, the moderately salinity-tolerant crop. Previous studies on the salinity tolerance of wheat used a wide range of salt levels, ranging from 0 mM (control) to 200 mM or even higher (Zair *et al.*, 2003; El-Hendawy *et al.*, 2005), and significant negative effects would generally be observed when the concentration of saline solutions were higher than 150 mmolL⁻¹. It has been reported that the salinity levels applied to a moderately salinity-tolerant wheat variety ranged from 0 mmolL⁻¹ to 320 mmolL⁻¹ and statistically significant changes were found in most variables where salt solutions higher than 120 or 180 mmolL⁻¹. Thus it is reasonable to deduce that the NaCl concentration of 100 mmolL⁻¹ used in our study did not provide sufficient stress for wheat, leading to insignificant effects of salinity on many of the variables. This study still provided useful information about salinity, however. The strongest saline stress used in the study was equal to slight saline condition if we compare the highest salt concentration to the saturation extract (100 mmolL⁻¹, which equals 5.8 gL⁻¹, slightly saline soil conditions). We can therefore deduce that these four wheat cultivars are able to grow in slightly saline soil without negative effects (Brouwer *et al.*, 1985).

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References

- Beltrano, J., Ronco, M. G. (2008). Improved tolerance of wheat plants (*Triticum aestivum* L.) to drought stress and rewatering by the arbuscular mycorrhizal fungus *Glomus claroideum*: Effect on growth and cell membrane stability. *Braz J Plant Physiol*, 20, 29-37. <https://doi.org/10.1590/S1677-04202008000100004>
- Brouwer, C., Goffeau, A., & Heibloem, M. (1985). Irrigation Water Management: Training Manual No.1 - Introduction to Irrigation. Food and Agriculture Organization of the United Nations, Rome, Italy.
- Colla, G., Roupheal, Y., Cardarelli, M., Tullio, M., Rivera, C. M., & Rea, E. (2008). Alleviation of salt stress by arbuscular mycorrhizal in zucchini plants grown at low and high phosphorus concentration. *Biol Fert Soils*, 44, 501-509. <https://doi.org/10.1007/s00374-007-0232-8>
- Daei, G., Ardekani, M. R., Rejali, F., Teimuri, S., & Miransari, M. (2009). Alleviation of salinity stress on wheat yield, yield components, and nutrient uptake using arbuscular mycorrhizal fungi under field conditions. *J Plant Physiol*, 166, 617-625. <https://doi.org/10.1016/j.jplph.2008.09.013>
- El-Hendawy, S. E., Hu, Y., Yakout, G. M., Awad, A. M., Hafiz, S. E., & Schmidhalter, U. (2005). Evaluating salt tolerance of wheat genotypes using multiple parameters. *Eur J Agron*, 22, 243-253. <https://doi.org/10.1016/j.eja.2004.03.002>
- Evelin, H., Kapoor, R., & Giri, B. (2009). Arbuscular mycorrhizal fungi in alleviation of salt stress: A review. *Ann Bot*, 104, 1263-1280. <https://doi.org/10.1093/aob/mcp251>
- Food and Agriculture Organization of the United Nations. (2015). FAOSTAT database. Available online at <http://faostat3.fao.org/home/E> (verified in June 2015).
- Gong, M., Tang, M., Chen, H., Zhang, Q., & Feng, X. (2013). Effects of two *Glomus* species on the growth and physiological performance of *Sophora davidii* seedlings under water stress. *New Forests*, 44, 399-408. <https://doi.org/10.1007/s11056-012-9349-1>
- Gupta, P. K., Mir RR, Mohan, A., & Kumar, J. (2008). Wheat genomics: Present status and future prospects. *Int J Plant Genomics*, 896451. <https://doi.org/10.1155/2008/896451>
- Günel, H., Korucu, T., Birkas, M., Özgöz, E., & Halbac-Cotoara-Zamfir, R. (2015). Threats to Sustainability of Soil Functions in Central and Southeast Europe. *Sustainability*, 7, 2161-2188. <https://doi.org/10.3390/su7022161>
- Harley, J. L., Smith, S. E. (1997). Mycorrhizal symbiosis. *Mycorrhizal Symbiosis*, 3(3), 273-281.
- Iman, A., Wahab, Z., Halim, M. R. A., & Rastan, S. O. S. (2006). The influence of N-P-K fertilizer rates and cropping systems on root biomass and some root morphological variables of sweet corn and vegetable soybean. *J Agron*, 5, 111-117. <https://doi.org/10.3923/ja.2006.111.117>
- Jones, G. P. D., Blair, G. J., & Jessop, R. S. (1989). Phosphorus efficiency in wheat—A useful selection criterion? *Field Crops Res*, 21, 257-264. [https://doi.org/10.1016/0378-4290\(89\)90007-5](https://doi.org/10.1016/0378-4290(89)90007-5)
- Marschner, H., & Dell, B. (1994). Nutrient uptake in mycorrhizal symbiosis. *Plant Soil*, 159, 89-102. <https://doi.org/10.1007/BF00000098>

- Mozafar, A., & Goodin, J. R. (1986). Salt tolerance of two differently drought-tolerant wheat genotypes during germination and early seedling growth. *Plant Soil*, *96*, 303-316. <https://doi.org/10.1007/BF02375135>
- Parniske, M. (2008). Arbuscular mycorrhiza: The mother of plant root endosymbioses. *Nature Rev Microbiol*, *6*, 763-775. <https://doi.org/10.1038/nrmicro1987>
- Pirozynski, K. A., & Malloch, D. W. (1975). The origin of land plants: a matter of mycotrophism. *Biosystems*, *6*(3), 153-64. [https://doi.org/10.1016/0303-2647\(75\)90023-4](https://doi.org/10.1016/0303-2647(75)90023-4)
- Porcel, R., Aroca, R., & Ruiz-Lozano, J. M. (2012). Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. *Agron Sustain Dev*, *32*, 181-200. <https://doi.org/10.1007/s13593-011-0029-x>
- Pozo, M. J., Jung, S. C., López-Rúz, J. A., & Azcón-Aguilar, C. (2010). Impact of arbuscular mycorrhizal symbiosis on plant response to biotic stress: The role of plant defence mechanisms. In Koltai H, Kapulnik Y (eds.) *Arbuscular Mycorrhizas: Physiology and Function* (2nd Ed). Springer, Dordrecht, Netherlands. pp. 193-207.
- Rad, M. Y., Noormohammadi, G., Ardakani, M. R., Hervan, E. M., & Mirhadi, S. J. (2009). Effect of mycorrhiza on morphological characteristics and nutrients content of barley under different salinity levels. *J New Agr Sci*, *5*, 105-114.
- Remy, W., Taylor, T. N., Hass, H., & Kerp, H. (1994). Four hundred-million-year-old vesicular arbuscular mycorrhizae. *P Natl Acad Sci USA*, *91*, 11841-11843. <https://doi.org/10.1073/pnas.91.25.11841>
- Sheng, M., Tang, M., Chen, H., Yang, B., Zhang, F., & Huang, Y. (2008). Influence of arbuscular mycorrhizae on photosynthesis and water status of maize plants under salt stress. *Mycorrhiza*, *18*, 287-296. <https://doi.org/10.1007/s00572-008-0180-7>
- Sun, S-Z., Guo, Y-Z., Guan, H-L., Hong, L-F., & Yang, P-C. (2010). Soil salinization characteristics of carnation cultivation under different management models and induced diseases. *Soils. (in Chinese)*, *42*, 972-977.
- Talaat, N. B., & Shawky, B. T. (2013). Modulation of nutrient acquisition and polyamine pool in salt-stressed wheat (*Triticum aestivum* L.) plants inoculated with arbuscular mycorrhizal fungi. *Acta Physiol Plant*, *35*, 2601-2610. <https://doi.org/10.1007/s11738-013-1295-9>
- Talaat, N. B., & Shawky, B. T. (2014). Protective effects of arbuscular mycorrhizal fungi on wheat (*Triticum aestivum* L.) plants exposed to salinity. *Environ Exp Bot*, *98*, 20-31. <https://doi.org/10.1016/j.envexpbot.2013.10.005>
- Wang, B., & Qiu, Y-L. (2006). Phylogenetic distribution and evolution of mycorrhizas in land plants. *Mycorrhiza*, *16*, 299-363. <https://doi.org/10.1007/s00572-005-0033-6>
- Zair, I., Chlyah, A., Sabounji, K., Tittahsen, M., & Chlyah, H. (2003). Salt tolerance improvement in some wheat cultivars after application of *in vitro* selection pressure. *Plant Cell Tiss Org Culture*, *73*, 237-244. <https://doi.org/10.1023/A:1023014328638>
- Zou, Y-N., & Wu, Q-S. (2011). Efficiencies of five arbuscular mycorrhizal fungi in alleviating salt stress of trifoliate orange. *Int J Agr Biol*, *13*, 991-995.

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