

Production of Xylitol from Oil Palm Empty Friuts Bunch: A Case Study on Bio refinery Concept

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Abstract

The concept of biorefinery offers the utilization of biomass, in particular agricultural waste, to be converted into energy, chemicals, materials, and food. In 2013 Indonesia produced about 27.4 thousand tons of crude palm oil (CPO) which corresponds to approximately 30 thousand tons of oil palm empty fruit bunches (EFB), the biomass waste from palm oil industries. The huge availability of EFB in Indonesia may serve as a good starting point to implement the concept of biorefinery. EFB mainly comprises of cellulose, hemicellulose and lignin. The cellulosic components of EFB have been thoroughly studied, i.e. for the production of bioethanol. The hemicellulosic component of EFB, which is a polymeric substance that comprises mainly of xylose, has been barely explored. This paper reviewed the potential utilization of hemicellulosic component of EFB to be converted to xylitol, the 5-carbon-sugar-alcohol which is low calorie, low Glycemic Index, and anti-cariogenic. The pretreatment and hydrolysis of EFB and the following fermentation of EFB hydrolysate to xylitol will be discussed further.

Keywords: enzymatic hydrolysis, fermentation, hemicellulose and xylitol

1. Introduction

Indonesia produced 17.5 million tons crude palm oil (CPO) in 2008. This tonnage increased by 50% to 26 million tons in 2012 and made Indonesia the biggest CPO producer in the world. Along with the production of CPO, the industry also produces palm kernel and solid wastes, such as empty fruit bunches (EFB), fiber, and shell, as well as liquid wastes, palm oil mill effluent (POME). The processing of 1 ton of oil palm fresh fruit bunch (FFB) would give 0.22 tons of EFB, 0.12 tons of fiber, and 0.05 tons of shell, and up to 2.5 – 3.75 tons of POME (Rupani et al., 2010; Visvanathan et al., 2009). Using this data, besides producing CPO, in 2012, Indonesia produced 28.6 million tons of EFB, 15.6 million tons of fiber, 6.5 million tons of shell, and 325 – 487 million tons of POME. This massive amount of waste could serve as potential raw materials for other industries.

In the concept of biorefinery, biomass feedstock is converted into a spectrum of valuable products. It is analogue to the concept of petroleum refinery where the petroleum is converted into a spectrum of energy and chemicals. This concept has been applied and commercial plants are being developed. Abengoa is developing a commercial plant that applies biochemical processes to convert stover, switch grass, and woody biomass mainly to bioethanol in Kansas, POET is developing a commercial plant in Iowa that applies biochemical processes to convert corn cobs mainly to bioethanol (http://www1.eere.energy.gov/bioenergy/integrated_biorefineries.html). Indeed several processes have been suggested for the utilization of the biomass waste from palm oil industries. The utilization of EFB, for example, was proposed for syngas via gasification (Susanto et al. 1997), for bio-oil (Chang et al., 2014), for compost of biofertilizer (Zainudin et al., 2014), pulp (Singh et al., 2014), polymer composite (Mahjoub et al., 2013), polyhydroxyalkanoate (Hassan et al., 2013), biogas (Embrandiri et al., 2012), or bioethanol (Sudiyani et al., 2013).

Considering that EFB is a lignocellulosic material, EFB can be potentially used as the raw material for the production of xylitol (Figure 1), a 5-carbon atom sugar alcohol that has a similar level of sweetness to sucrose, but lower calorie. It is a natural sweetener that can be found in fruits and vegetable, albeit in small quantity. The glycemic index of xylitol is low and thus can be safely used by diabetic people. In addition, xylitol has a strongly

negative heat of solution thus can give a 'cooling sensation'. Xylitol is widely used in food and pharmaceutical industries. In 2007, the world market of xylitol reached \$340 million at a price of \$ 4-5 per kg (Toyoda et al., 2009). Internet survey shows that China is the main xylitol producer in the world.

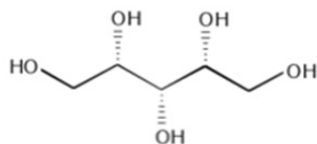


Figure 1. Structural Formula of Xylitol

Steps involved in the production of xylitol from EFB are the conversion of its hemicellulose to xylose via pretreatment and hydrolysis, and reduction of xylose to xylitol. This article highlights the implementation of biorefinery concept for the conversion of oil palm EFB to xylitol. Overall, enzymatic hydrolysis of EFB followed by yeast fermentation of hydrolysate is proposed for the production of xylitol from EFB (Figure 2). The aspects involved in the xylitol production, covering the characterization of EFB, pretreatment, enzymatic hydrolysis, fermentation, downstream processing, as well as the economic aspects will be discussed further.

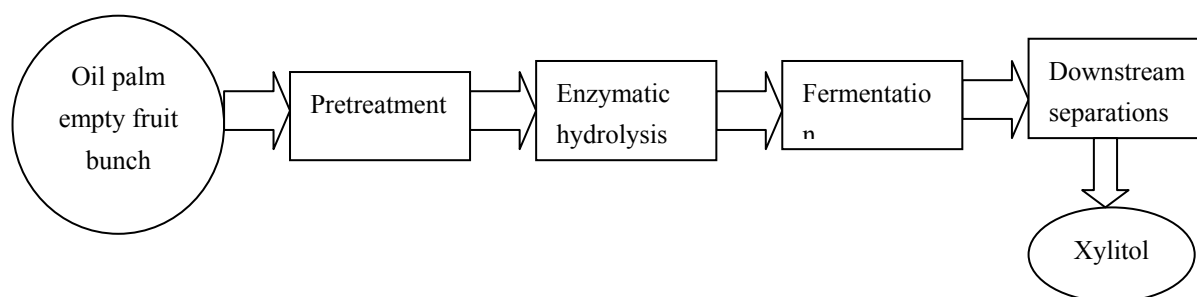


Figure 2. Microbial Production of Xylitol from EFB

2. Oil Palm Empty Fruit Bunch

Oil Palm Empty Fruit Bunch (EFB) is the main biomass waste of the palm oil milling process. According to (Visvanathan et al., 2009), 0.22 tons of EFB is produced from every ton of FFB or 1.1 tons of EFB will be produced for every ton of CPO. EFB is lignocellulosic material, which composition depends on the genetics of the oil palm as well as the environmental condition where the oil palm tree is grown. Despite some variation in the composition, Table 1 shows that cellulose, lignin, and hemicellulose are the main components of EFB. Cellulose is the homopolymer of glucose, linked by β 1 \rightarrow 4 glycosidic bond. The strong hydrogen interaction between glucose monomer makes cellulose has a high tensile strength and strong crystalline structure. Lignin is a complex biopolymer comprises of aromatics compounds. In the lignocellulosic material, lignin links cellulose and hemicellulose, it also provides the structural support for the biomass. Hemicellulose is a heteropolymer comprises mainly of 5-carbon atom sugars, such as xylose and arabinose. Component analysis of EFB showed in (Mardawati et al., 2013; Octavia, 2013) showed that xylose composes 19.2 – 19.6% of EFB, whereas arabinose, mannose and galactose are detected at lower level, successively at 1.9%, 1.4%, and 1.2% (on dry basis). These show that the hemicellulose of EFB is mainly composed of xylose, and further suggests that EFB has the potential to be used as the raw material for xylitol production.

Table 1. Composition of Oil Palm Empty Fruit Bunch (dry basis)

Cellulose	Composition		References
	Hemicellulose	Lignin	
42.85%	11.70%	24.01%	Rahman et al., 2007
33.25%	23.24%	25.83%	Sudiyani et al., 2013
33.25%	23.24%	25.83%	Millati et al., 2011
43-43.47%	22.93-23.67%	21.28-22.10%	Mardawati et al., 2014

3. Pretreatment

Lignocellulosic material is such a complex polymeric material that its physicochemical structure and composition may hinder the enzymatic digestibility of its components. The pretreatment is performed to increase the access of enzyme to the material (Mosier et al., 2005). The pretreatment process can be either physical, chemical, or physicochemical processes. Examples of physical pretreatment process are milling, thermal, or radiation. In the chemical pretreatment process, chemical substances such as dilute alkali or dilute acid are added to degrade the structure of lignocellulosic material. Physicochemical process is a combination of both involving the addition of chemical substances and extreme physical process, such as steam explosion. Different pretreatment process has been evaluated in utilizing EFB for bioethanol production, for example the research studied the dilute alkaline steam explosion (Choi et al., 2013), other research studied the combine use of white rot fungi and phosphoric acid (Isroi et al., 2012), and the use of bisulphite for EFB pretreatment (Tan et al., 2012). It should be noted that the main compound needed for xylitol production, the hemicellulose, is easier to degrade than cellulose or lignin. Therefore the condition for pretreatment should be designed as such that hemicellulose is not over-degraded or lost during the pretreatment. Kim et al. (2012) reported that dilute acid pretreatment of EFB will remove 90% of the hemicellulose and the degradation kinetics of hemicellulose under acid pretreatment. Imman et al. (2013) reported that EFB pretreatment with compressed liquid hot water at 200°C for 5-20 minutes led to high level of hemicellulose solubilising. Our research indicated even autoclaving at 121 °C for 15 minutes, the standard procedure for lab scale sterilization, is an effective pretreatment method for the utilization of EFB for xylitol production (Mardawati et al., 2014).

4. Enzymatic Hydrolysis of EFB

Hydrolysis is the breaking down of a polymeric compound into its monomer with the addition of water molecules. The hydrolysis of EFB can be performed chemically or enzymatically. The chemical hydrolysis uses either acid or alkaline as the catalyst at high temperature and pressure (Sun & Chen, 2002). This process is relatively fast, and thus need to be carefully controlled to prevent the co-production of various degradative products that inhibit the latter processes. Nonetheless, the process condition requires a specific material for the construction of the reactor and the related equipment. Further, subsequent pH neutralization is required. (Rahman et al., 2007), for example, reported the application of acid hydrolysis in the processing of oil palm EFB to xylose using dilute (2-6%) sulfuric acid at moderate temperature, 120°C (Parajo et al., 1998a). The reaction should be executed within a short time such that the possible inhibitory compounds for the following xylitol fermentation, that is furfural and acetic acid, are not co-produced during the acid hydrolysis. The hydrolysis of EFB can also be performed enzymatically using lignocellulolytic enzyme (Parajo et al., 1998b). In particular, the hydrolysis of hemicellulose is catalysed by xylanolytic enzyme or in short, xylanase. Albeit proceeding slowly, the enzymatic hydrolysis works at milder condition and reacts specifically. Thereby the related utility cost for the hydrolysis process is relatively low and cheaper material can be used for the reactor and equipments. This process can be easily integrated with the fermentation (Vazques et al., 2001). The xylanase is actually an enzyme complex that has the activity to hydrolyze hemicellulose. (Shallom et al., 2003) described that this enzyme comprises of many activities, among others are 1,4- β -endoxylanase (EC.3.2.1.8), that breaks the β -1,4 glycosidic bond at the xylan main chain of the hemicellulose, and 1,4- β -exoxylosidase (EC.3.2.1.37), that breaks the β -1,4 glycosidic bond at two or four monomer from the free end of xylan .

4.1 Availability of Xylanase

The xylanases are found in various bacteria and fungi (Table 2). Wheat bran, corn cobs, rice bran, cassava, maize straw, and sugarcane bagasse are mostly used as sources for hemicellulose.

In our research we studied the production of xylanase from EFB in solid state fungal cultivation using *A.niger* ITB L.51, *Trichoderma viride* ITB L.67 and *Penicillium sp* ITB L.96. The best activity was obtained from the *T.viride* cultivated at 32.8 °C for 36 hours and gave the specific xylanase activity of 5,180 U/g dry substrate (Mardawati et al., 2013). The production of xylanase from EFB in solid state fungal cultivation was also reported by (Lakshmi et al., 2011) using *A.terreus* and *A.fumigatus*. Optimization of cultivation condition, such as EFB particle size, moisture content, pH, media composition, inoculum concentration, and additional carbon source may increase the produced enzyme activity up to 40,000 U/g dry substrate. Commercial xylanase for the hydrolysis of lignocellulosic material is available in the market. Among others are produced by Novozyme (Denmark), Genecor (Mardawati et al., 2014) and Meiji Seika (Japan). The application of xylanase can also be found in biobleaching process in the pulp and paper industries, in clarification of fruit juice, quality improvement of bread and feed, waste treatment, and composting.

Table 2. Potential Xylanase Producers

Microorganism	Reference
<i>Streptomyces sp</i>	Meryandini et al., 2008
<i>Bacillus sp</i>	Shallom and Shoham, 2003
<i>Aspergillus sp</i>	Xu et al., 2008; Lakshmi et al., 2011 and Pal et al., 2011
<i>Penicillium</i>	Tran et al., 2004; Knob et al., 2009 and Assamoi et al., 2008
<i>Trichoderma</i>	Kar et al., 2008; Chen et al., 2010 and Zhou et al., 2011

4.2 Operation Condition of Hydrolysis

Depending on the molecular structure and composition, each enzyme has a particular condition (e.g. temperature and pH) whereby its activity is optimum. The optimal conditions of xylanase, for example in hydrolyzing corncob, were reported to be in the range of 45-60°C and pH of 4 – 5 (Vasquez et al., 2001). The optimal condition of commercial xylanase in hydrolyzing oil palm EFB was reported at 60°C and pH 5.0 (Mardawati et al., 2014).

4.3 Kinetics of Enzymatic Hydrolysis

The Michaelis Menten kinetics is mostly used to model enzymatic reactions. In particular, when enzyme, substrates, and products are homogeneously mixed in the aqueous solution. In the Michaelis Menten equation, the rate of enzymatic reactions (v) is described as a saturation curve of substrate concentration (S) as is described in eq. 1.

$$v = v_m [S]/(K_m + [S]) \quad (1)$$

The two kinetic parameters, v_m and K_m , respectively described the maximum rate of the enzymatic reaction at a defined enzyme concentration and the enzyme affinity to the substrate. However, EFB as the substrate of enzymatic hydrolysis is insoluble solid particles and thereby the Michaelis Menten kinetics may not be applicable in describing the EFB hydrolysis reaction kinetics. (Mardawati et al., 2014) showed that at low solid loading, i.e. the ratio of EFB to the buffered enzyme solution, the hydrolysis reaction could be well approached with the Michaelis Menten kinetics (Mardawati et al., 2014). At the optimum condition, temperature of 60 °C and pH 5.0, the kinetics parameters were estimated to be $v_m = 0.34 \text{ g xylan.L}^{-1}.\text{min}^{-1}$ and $K_m = 52.84 \text{ g xylan.L}^{-1}$. Further experiments need to be done to confirm whether this is also valid at high solid loading. Intuitively, the transport of enzyme to the substrate may be limiting. The obtained kinetics parameters can be further used to simulate and design the reaction, such as how long the hydrolysis needs to be undertaken to achieve the targeted yield. Using the above parameters, for example, xylose yield of 90% will be achieved at 6 hours of enzymatic hydrolysis of EFB with 5% solid loading.

5. Fermentation

The reduction of xylose to xylitol can be performed chemically or biologically. The chemical reduction is carried out by contacting liquid xylose with pure hydrogen gas using noble metalbased catalysts such as platinum, palladium, ruthenium, or nickel, at high pressure and temperature (Baudel et al., 2005). The biological reduction of xylose is carried out by fermentation, mainly using yeast (Parajo et al., 1998b). This process is carried out at ambient temperature, pressure, and pH. The study of microbial production of xylitol was dated back to 1969, in search of potential yeast for converting glucose to xylitol (Onishi & Suzuki, 1969). No direct mechanism was found to convert glucose to xylitol. Instead, glucose needs to be converted to arabitol, arabitol to xylulose, and finally xylulose to xylitol. Later studies suggested the use of xylose as the substrate for xylitol fermentation. (Parajó et al., 1998) listed several potential microorganisms for xylitol production from xylose. Among them are *Candida utilis*, *C. tropicalis*, *C. guilliermondi*, *C. mogii*, *Debaromyces hansenii*, *Pachysolen tannophilus*, and *Pichia stipitis*. In using xylose as the substrate, these microorganisms transport xylose from the media to the cell and converting the intracellular xylose into xylitol by xylose reductase (EC.1.1.1.21) using either NADH or NADPH as the co-enzyme. The produced xylitol further oxidized by xylitol dehydrogenase (EC.1.1.1.B19) to xylulose ((Parajo et al., 1998b). The latter can be activated by xylulose kinase into xylulose-5-phosphate that can assimilate with the Pentose Phosphate Pathway of the yeast metabolism. Some part of xylitol is transported back to the media (Ghindea et al., 2010).

How much xylitol is used in the metabolism and how much xylitol is exported as product? This will be affected among others by the amount of xylose in the substrate, the presence of co – substrate, and aeration condition as is summarised in Table 3.

Table 3. Factors Affecting Xylitol Production in Yeast

Factors	Influence of factors	Reference
Substrate concentration	High xylose concentration led to slow xylitol production due to substrate inhibition.	Parajo et al., 1998b
The presence of cosubstrate	Glucose concentration (in small amount) led to higher xylitol productivity due to higher biomass growth.	Ghindea et al., 2010
Aeration condition	High oxygen concentration (aerobic) led to higher biomass growth and low xylitol production	Barbosa et al., 1994
Inoculum condition	Higher initial cell concentration led to higher xylitol production	Kresnowati et al., 2012

In previous study *D.hansenii* ITBCC R85 can be used to produce xylitol from EFB hydrolysate (Kresnowati et al., 2012). Interestingly, the results also show higher xylitol yield, and higher products selectivity in fermentation using EFB hydrolysate compared to using synthetic media. The kinetics of xylitol production is being studied further. Further important factor is the fermentation configuration, such whether the fermentation will be performed in batch, fed batch, or continuous; whether cell will be immobilized and how.

6. Downstream Processing

Not many literatures are found on the topics of downstream processing of xylitol from the fermentation broth. Overall the important considerations in downstream processing of microbial production of xylitol are the characteristics of xylitol as well as other substances found in the fermentation broths. (Kresnowati et al., 2012; Kresnowati et al., 2013) observed that besides xylitol and biomass, ethanol, acetic acid, and glycerol were also produced during the xylitol fermentation. When EFB hydrolysate is used, EFB residue may also be found in the fermentation broth. Main methods used in the downstream processing of xylitol is crystallization, however, considering its composition, direct crystallization of the fermentation broth yields xylitol crystal of low quality (Aliakbarian et al., 2012). Therefore, preceding separation process is necessary. There are applied charcoal adsorption and crystallization as the downstream processing of xylitol fermentation (Sampaio et al., 2006), applied silica gel adsorption and crystallization (Musatto et al., 2006), proposed extraction, precipitation, or vacuum concentration as the preceding steps (Misra et al., 2011), proposed membrane processes that are ultrafiltration and reverse osmosis, as the preceding steps for crystallization (Affleck et al., 2000). Specific downstream processing configuration of xylitol from EFB hydrolysate has not been reported. Some proposed downstream processing configurations are presented in Figure 3.

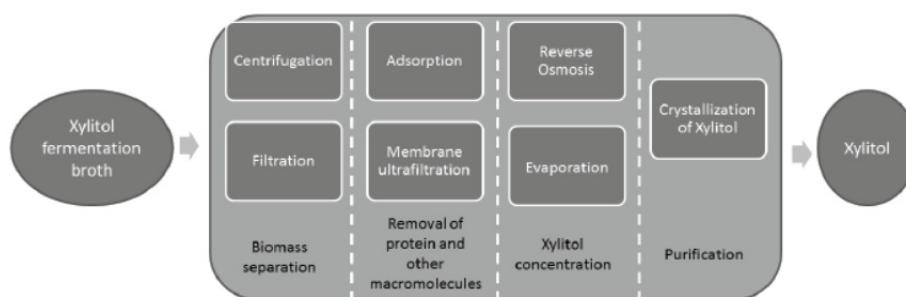


Figure 3. Alternative Process Configurations for the Downstream Processing of Microbial Xylitol Production for EFB

7. Economic Aspect

The economic potential of xylitol production is quite promising. For average xylitol price of 4.5 US\$/kg (Toyoda et al., 2009), average EFB price of 150 IDR/kg, and realistic yields of hydrolysate (80%) and fermentation yield (50%), a gross profit of 4.3 US\$/kg is obtained. Even six folds increase in the EFB price or 20% decrease in the xylitol price still gives a gross profit of 3.5 US\$/kg. This value gives some flexibility in the processing cost. Still, the main process in the downstream processing of xylitol, crystallization, is an energy intensive process. Process improvement and optimization will be an advantage. Further, in considering the industrial scale, the EFB transportation and average scale oil palm industries need to be taken into consideration.

8. Conclusions and Future Recommendation

This paper shows that oil palm EFB has the potential to be converted into xylitol. The process involves pretreatment, hydrolysis, fermentation, and downstream separation. Moderate temperature and pressure thermal process such as autoclave sterilization can effectively be applied as the EFB pretreatment process. Enzymatic hydrolysis followed by microbial fermentation is a more sustainable process alternative. Detail information on these processes, such as suitable microorganism and enzyme, optimal condition, and reaction kinetics, has been investigated. Process improvement and optimization are being studied. More efforts should be spent on the optimization of energy intensive downstream separation to bring this concept closer to the implementation.

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