

# Characterizing the Distribution of Ppm Gluten in Gluten Free Oatmeal Servings Contaminated with a Wheat Kernel

Ronald D. Fritz<sup>1</sup> & Yumin Chen<sup>1</sup>

<sup>1</sup>PepsiCo R&D Measurement Sciences, 617 W. Main Street, Barrington, IL 60010, USA

Correspondence: Ronald D. Fritz, Measurement Sciences, PepsiCo R&D, 205 Beelog Rd., Burnsville NC 28714, USA. Tel: 1-828-678-9342. E-mail: ronald.fritz@pepsico.com

Received: July 21, 2017

Accepted: August 6, 2017

Online Published: August 25, 2017

doi:10.5539/jfr.v6n5p92

URL: <https://doi.org/10.5539/jfr.v6n5p92>

## Abstract

Oats are often contaminated with rogue kernels of gluten-containing grains like wheat, barley and rye. When producing gluten free oatmeal, possessing an understanding of the consequences of this possibility is prudent, as labeling requirements specify a maximum amount of gluten in terms of ‘parts per million’ (ppm) gluten. Variation in contaminant kernels, along with variation due to measurement itself though, can result in a wide range of possible ppm gluten outcomes in contaminated servings. This research pursues characterization of this variability, highlighting contributors to it, doing so by quantifying distributional outcomes of ppm gluten in wheat kernel contaminated servings. This is done via statistical simulation of wheat kernel contaminated servings, done for a collection of wheat types and incorporating various measurement influences. Results indicate substantial variability in ppm gluten per serving for a given wheat type, as well as between them, with this being compounded by the measurement task itself.

**Keywords:** oat, ELISA, gluten, kernel-based gluten contamination, gluten analysis, gluten-free, serving size

## 1. Introduction

Oats are often contaminated with gluten rich grains like wheat and barley (Hernando, Mujico, Mena, Lombardia, & Mendez, 2008; Koerner et al., 2011; Thompson, 2004; Thompson, Lee, & Grace, 2010). This ‘kernel based’ type of contamination complicates attainment and assessment of true gluten free oatmeal (Fritz & Chen, 2017), as these rogue kernels remain largely intact during processing, being transformed into flakes indiscernible from their oat counterparts. This type of contamination produces a binary type compliance circumstance, since if a contaminant kernel exists in a serving that serving will be high in measured gluten and non-compliant, while if no contaminant kernel exists no gluten will be present (Fritz & Chen, 2017).

In previous research, spiking of pure oats with contaminate kernels of wheat was conducted (Fritz, Chen & Contreras, 2017). During this, it was noticed that a surprisingly wide range of ppm gluten outcomes were obtained. This was despite care being taken to contaminate equal serving amounts with wheat kernels of comparable weight and type. These disparate outcomes became the genesis of this research, which seeks to understand and characterize the distribution of ppm gluten outcomes for a wheat kernel contaminated serving of otherwise pure oats.

To accomplish this, kernel attributes which can influence gluten content in wheat were identified, and distributions of these characteristics were obtained/derived from the literature. Measurement influences on the gluten assessment task for oats were also identified and quantified. Where pertinent information could not be found, distributional parameters of certain characteristics were ‘guesstimated’ and sensitivity analysis performed by perturbing these values in the statistical simulation model. This work resulted in a collection of variables which could be used to simulate servings of contaminated oatmeal, and which could then be collectively used to characterize the distribution of gluten (in ppm) in a defined serving size of gluten free oatmeal. This was done for various wheat kernel varieties and gluten measurement circumstances. These circumstances include distributions for ‘actual’ ppm outcomes (i.e., absent of measurement influences), ppm outcomes under various measurement scenarios, and done for various wheat classifications.

## 2. Materials and Methods

### 2.1 Methods – Kernel Variability

It is widely known that gluten resides in the endosperm of wheat, existing in the form of various seed storage proteins (Haraszi, Chassaing, Maquet, & Ulberth, 2011; Huebner, 1970; Rallabhandi, Sharma, Pereira, & Williams, 2015; Shewry, 2009). Based on this, four key variables are believed to directly influence the gluten content of a wheat kernel. They are:

1. kernel weight
2. % of kernel weight due to moisture
3. % of dry kernel weight that is protein
4. % of this kernel protein which is gluten (i.e., gliadins and glutenins) (Mejías et al., 2014; Shewry, 2009).

Found in published literature, estimates of distribution parameters for the above, for both hard and soft wheat classifications, are shown in table 1. Note that the stdev of ‘% protein that’s gluten’ was unable to be found in the literature and has been ‘guesstimated’ here. Sensitivity analysis around this ‘guesstimate’ has been performed and will be reported in the results and discussion section of this paper. Also note that these variables are assumed to be normally distributed. Evidence of this exists for ‘% protein’ (Levi, 1950), and this is believed a reasonable assumption for ‘weight’, ‘moisture’ and ‘% protein that’s gluten’ as well.

Table 1. Approximate distributional parameters for individual wheat kernels based on hardness classification for four key variables affecting individual kernel gluten content

Assumed Distribution Type	Kernel Weight (g)		Kernel Moisture (%)		Kernel % Protein (Gliadins + Glutenins)		% Protein as Gluten	
	Normal		Normal		Normal (Levi, 1950)		Normal	
	~ Avg.	~ Stdev	~ Avg.	~ Stdev	~ Avg.	~ Stdev	~ Avg.	~ Stdev
Soft Wheat	0.041	0.006	12.7%	2.4%	9.6%	2.1%	76.7%	4.0%
Hard Wheat	0.035	0.010	11.1%	1.8%	13.3%	2.4%	80.3%	4.0%
Data Source	(Liu, 2008)		(Liu, 2008)		(Delwiche, 1995)		(Seilmeier, 1991)	Guesstimate

These four variables are also assumed to be independent of each other (i.e., not correlated.) This appears to be the case regarding protein content relative to dry kernel weight (Delwiche, 1995), where no relationship was apparent for hard white wheat in that research. Other relationships between these variables, or lack of them, are not well defined in the literature but are assumed herein not to exist.

### 2.2 Methods – Measurement Variability

Three measurement variables were considered in characterizing the ppm gluten in a wheat contaminated serving of otherwise gluten-free oatmeal. This was to determine gluten content ‘as assessed’ vs. an estimate of ‘actual true gluten’ exclusive of measurement itself. These measurement variables are:

5. % of gluten protein which is gliadin (this is technically a kernel attribute but is included here due to its significant impact on gluten assessment since test kits measure gliadin only)
6. % analytical recovery of gliadin
7. Effects of grinding non-homogeneity (of a wheat kernel in oats).

As mentioned, the variable ‘% of gluten protein which is gliadin’ is included in the measurement category here since gluten test kits rely on measurement of gliadins only and estimate a doubling of this for potential gluten containing glutenin proteins also present in the kernel. Glutenins can contain significant amounts of gluten as well (Huebner, 1970; Selmeier, Belitz & Wieser, 1991; Weiser & Koehler, 2009; Haraszi et al., 2011; Zilic, Barc, Pesic, Dodig & Ignjatovic-Micic, 2011; Diaz-Amigo & Popping, 2013; Khan et al., 2013).

‘% analytical recovery’ has been included since no validation of this for oats is believed to have been performed, as there’s no record of this published by Biopharm at this time. It is also possible that this may vary somewhat

‘test to test’ as well.

The ‘effect of grinding non-homogeneity’ has been included since it has been found that this inflates gluten ppm test outcome variability, producing a lognormal distribution of gluten test results (Fritz et al., 2017).

Estimates of distribution parameters for the above three variables, along with their data sources, are shown in table 2. Note that the stdev of ‘% gluten protein that’s gliadin’ was unable to be found in the literature and has been ‘guesstimated’ here. This is the case with the stdev of ‘analytical recovery’ of gliadin as well. Sensitivity analysis around both of these ‘guesstimates’ has been performed and will be reported in the results and discussion section of this paper.

Also note that ‘% Gluten Protein that’s Gliadin’ and ‘Analytical Recovery (%)’ have been assumed to be normally distributed.

Table 2. Approximate distributional parameters for measurement related variables that affect gluten content assessments of gluten-free oatmeal contaminated with a single wheat kernel

	% Gluten Protein that's Gliadin		Analytical Recovery (%) (Gliadin)		Grinding Non-Homogeneity	
Assumed Distribution Type	Normal		Normal		Log-Normal	
	~ Avg.	~ Stdev	~ Avg.	~ Stdev	~ Avg. of Stdev of Ln values	~ Stdev of Stdev of Ln Values
<b>Wheat</b>	50.0%	7.0%	80.0%	8.5%	0.510	0.082
<b>Data Source</b>	(Huebner, 1970)	Guess-timate	(BioPharm, 2012)	Guess-timate	(Fritz, 2017)	

### 2.3 Methods – Simulation

These seven variables, four being wheat characteristics (i.e., characterized in Table 1) and three measurement variables (i.e., characterized in Table 2), have been used to simulate a serving of gluten-free oatmeal (of 40g size) which is contaminated by a single wheat kernel. A simulated serving then, is 40g of pure oats, contaminated by a randomly selected wheat kernel of a certain weight, moisture, protein content, and ‘% of that protein that’s gluten’ (i.e., gliadin plus glutenin). These values are based on random selection from the distributions for these variables defined in table 1. These kernel characteristics are considered independent of each other, so a kernel of a certain weight is randomly chosen from the ‘weight’ distribution, then a moisture content is randomly assigned to it from the ‘moisture’ distribution, then ‘% protein’ and ‘% protein that’s gluten’ values are assigned, again with these coming from their respective distributions of possibilities. So, for a ‘true’ ppm gluten assessment (exclusive of any measurement error), the following defines a single simulated wheat kernel’s contribution in ppm gluten in a 40g serving of pure oats:

$$PPM\ gluten_{true} = [(weight\ in\ \mu g) \times ((100 - \% moisture)/100) \times (\% Protein/100) \times (\% Protein\ that's\ Gluten/100)]/40g \tag{1}$$

To this, measurement variables are then assigned. So, the ‘% gluten that’s gliadin’ is randomly chosen from the distribution defined for it, along with an analytical recovery value from its distribution. Both of these are assumed normally distributed as well. This gives us the following for a ‘pre-grind’ ppm gluten for a serving:

$$PPM\ gluten_{pre-grind} = (PPM\ gluten_{true}) \times 2 \times ((\% Gluten\ that's\ Gliadin)/100) \times (\% Analytical\ Recovery/100) \tag{2}$$

By ‘pre-grind’ is meant prior to incorporating the variability due to the inability to achieve an homogenous grind of gluten in a serving of oats (Fritz et al., 2017). Also, the ‘2’ in this equation is the recommended test kit multiplier which presumes a Gliadin/Glutenin ratio of 1:1, and therefore a doubling of the gliadin to obtain an estimate of total gluten.

For the final ‘post-grind’ ppm value for a serving, the ‘PPM gluten<sub>pre-grind</sub>’ outcome for a kernel in 40g of oats is used as the gluten ppm average of a log-normally distributed collection of 0.25g test results in a 40g serving



Also shown in Table 3, are outcomes including measurement variability for the same hard wheat kernel category. Here we see that sample grind non-homogeneity alters the distribution of ppm gluten/serving from a normal-like distribution to a log-normal-like one (Fritz et al, 2017.) And, that this is responsible for further inflating the variability of possible outcomes (relative to the true variability due just to ‘kernel to kernel’ variability.) Solutions to this grinding non-homogeneity issue could substantially improve ppm gluten outcomes ‘as measured’, providing more accuracy and precision in assessment of serving gluten content.

Along with this, the other two measurement variables play into inflated variability of ppm gluten/serving outcomes as well. For instance, the ‘% gluten that’s gliadin’ plays a big role as test kits measure only gliadin and assume a 1:1 ratio of gliadin to glutenin (Huebner, 1970). Doing a survey of research regarding gliadin/glutenin ratios though, shows this ratio can vary substantially (Huebner, 1970; Selmeier et al., 1991; Weiser & Koehler, 2009; Haraszi et al., 2011; Zilic et al., 2011; Diaz-Amigo & Popping, 2013; Khan et al., 2013), as illustrated in Table 4. Varying gliadin as a percent of total gluten, from 33% to 70%, produces average ‘ppm/serving gluten’ from 50 ppm to 110 ppm, and with stdevs from 37 ppm to 83 ppm respectively, as shown in Table 3. This illustrates that improved test kit design, which better accounts for the glutenin portion of overall gluten, would provide improved accuracy and precision in the assessment of gluten content caused by wheat kernel contamination in oats.

Table 4. % Gluten due to Gliadin and Glutenin cited in various research studies

Study	% of Total Gluten	
	Gliadin	Glutenin
Huebner, 1970	50%	50%
Selmeier, 1991	38%	62%
Weiser, 2009	60% - 76%	24%-40%
Haraszi, 2011	65%	35%
Zilic, 2011	33% - 50%	50% - 67%
Diaz-Amigo, 2013	57% - 62%	38% - 43%
Khan, 2013	62% - 69%	31% - 38%

Finally, analytical recovery of gluten in oats is not believed to have been validated as of yet. (At least there’s no record of this published by Biopharm at this time.) The 80% figure often cited is for corn (R-Biopharm, 2012). To account for recovery of gluten in oats possibly being under or over stated due to this, we performed sensitivity analysis by perturbing analytical recovery from 70% to 90%. We found that this range affects average ppm gluten/serving by about 20 ppm, and increased the stdev of ppm gluten/serving from 48 to 61 ppm as shown in the bottom two rows of Table 3.

Note also that in table 3, there are comments regarding sensitivity analysis to the ‘guesstimates’ made for the stdevs of:

- ‘% Protein that’s Gluten
- ‘% Gluten that’s Gliadin’
- ‘Analytical Recovery.’

In all cases, altering these as noted produced only modest influences on resultant ppm gluten/serving results.

In summary, kernel to kernel variability can produce widely varying outcomes in terms of ppm gluten/serving when a wheat kernel exists in a serving. Also, measurement further compounds this by inflating variability due to varying gliadin/glutenin ratios, non-homogenous grinding and potentially a test method not yet validated for oats in terms of analytical recovery of gluten.

### 3.2 Comparing PPM Distributions of Gluten/Serving in Oats Contaminated with a Wheat Kernel, Using Different Wheat Types

There are substantial differences in kernel size, moisture, protein, etc. between different varieties of wheat (<http://www.uswheat.org/reports>). Table 5 shows approximate values of weight, moisture content and protein for some major sub-classifications of wheat. Table 5 also shows simulated outcomes of the distribution of ppm gluten/serving as measured (employing stdevs from the larger categories of hard and soft wheat used earlier, and also with assumptions of 50/50 gliadin to glutenin ratio and 80% recovery). The differences in average contamination obtained between wheat types, varies from a low of 50 ppm up to 110 ppm. This illustrates the differences that exist in contamination potency between differing types of wheat.

Table 5. Simulated outcomes of ppm gluten in gluten-free oatmeal servings (40g) contaminated with a single wheat kernel for both Hard and Soft Wheat. (n = 25,000 simulated servings.)

Simulation	Theoretical PPM Calc. (12% Moisture)	Kernel Variable Settings								Measurement Variable Settings								Dist. Shape	Avg. PPM Gluten	Stdev of PPM Gluten	Plot
		Kernel Weight (g)		Kernel Moisture (%)		Kernel % Protein (Gliadins + Glutenins)		% Protein as Gluten		% Gluten that's Gliadin		Analytical Recovery (% Gliadin)		Grinding Non-Homogeneity							
		Avg. *	Stdev **	Avg. *	Stdev **	Avg. *	Stdev **	Avg. ***	Stdev	Avg.	Stdev	Avg.	Stdev	Avg. of Stdev of 'In' values	Stdev of Stdev of 'In' Values						
Hard Red Winter	65	0.0291	0.01	11.1%	1.8%	12.7%	2.4%	80.3%	4% ****	50%	7.0%	80%	8.5%	0.51	0.082	Log-Normal Like	61	46			
Hard Red Spring	76	0.0304	0.01	12.2%	1.8%	14.1%	2.4%									Log-Normal Like	70	52			
Soft Red Winter	55	0.0326	0.006	13.0%	2.4%	10.0%	2.1%	76.7%	4% ****	50%	7.0%	80%	8.5%	0.51	0.082	Log-Normal Like	50	35			
Soft White	60	0.0353	0.006	9.4%	2.4%	10.0%	2.1%									Log-Normal Like	56	38			
Northern Durum	93	0.0392	0.01	11.6%	1.8%	13.5%	2.4%	80.3%	4% ****	50%	7.0%	80%	8.5%	0.51	0.082	Log-Normal Like	86	60			
Desert Durum	113	0.048	0.01	6.4%	1.8%	13.4%	2.4%									Log-Normal Like	110	74			

\* - Five year average values reported in 2015 Crop Quality Report by US Wheat Associates, <http://www.uswheat.org/cropQuality>

\*\* - Liu (2008)

\*\*\* - Seilmeier (1991)

\*\*\*\* - Guesstimate (variation in this shown to be a minimal influencer on ppm gluten/serving)

#### 4. Conclusion

Oats are easily contaminated with gluten rich kernels of wheat. As shown here, the extent of actual contamination per serving in terms of ppm gluten can vary widely when this occurs. This is due to kernel to kernel variability within varieties, as well as between them. The measurement of gluten itself compounds this variability as well. Primary components of measurement influence are the use of gliadin as a 50% marker for glutenins when gliadin/glutenin ratios can actually vary substantially, the inability to homogeneously grind a wheat kernel within a serving of otherwise gluten-free oats and finally the potential for variability in analytical recovery sample to sample. All of this adds up to produce a substantial range of possible outcomes when a gluten kernel exists in a serving of otherwise gluten-free oats.

#### Acknowledgements

The authors declare that they have no competing interests. All the authors are salaried employees of PepsiCo Inc. or Quaker Foods and Snacks (QFS), a subsidiary of PepsiCo Inc., which funded this research. QFS has a commercial interest in gluten-free foods. The views expressed in this manuscript are those of the authors and do not necessarily reflect the position or policy of PepsiCo Inc.

#### References

Delwiche, Stephen, R. (1995). Single Wheat Kernel Analysis by Near-Infrared Transmittance: Protein Content. *Cereal Chemistry*, 72(1), 11-16.

Diaz-Amigo, C., & Popping, B., (2013). Accuracy of ELISA Detection Methods for Gluten and Reference Materials: A Realistic Assessment. *J. Agric. Food Chem.*, 61, 5681-5688. <https://doi.org/10.1021/jf3046736>

Fritz, R. D., & Chen, Y. (2017). Kernel-based gluten contamination of gluten-free oatmeal complicates gluten assessment as it causes binary-like test outcomes. *International Journal of Food Science & Technology*, 52(2), 359-365. <https://doi.org/10.1111/ijfs.13288>

Fritz, R. D., Chen, Y., & Contreras, V. (2017). Gluten-containing grains skew gluten assessment in oats due to sample grind non-homogeneity. *Food Chemistry*, 216, 170-175. <https://doi.org/10.1016/j.foodchem.2016.08.031>

Haraszi, R., Chassaing, H., Maquet, A., & Ulberth, F. (2011). Analytical Methods for Detection of Gluten in

- Food - Method Developments in Support of Food Labeling Legislation. *Journal of AOAC International*, 94(4), 1006-1025
- Hernando, A., Mujico, J. R., Mena, M. C., Lombardia, M., & Mendez, E. (2008). Measurement of wheat gluten and barley hordeins in contaminated oats from Europe, the United States and Canada by Sandwich R5 ELISA. *Eur J Gastroenterol Hepatol*, 20(6), 545-554. <https://doi.org/10.1097/MEG.0b013e3282f46597>
- Huebner, F. R. (1970). Comparative Studies on Glutenins From Different Classes of Wheat. *Agricultural and Food Chemistry*, 18, 256-259. <https://doi.org/10.1021/jf60168a006>
- Huebner, F. R., Kaczkowski, J., & Bietz, J. A. (1990). Quantitative Variation of Wheat Proteins from Grain at Different Stages of Maturity and from Different Spike Locations. *Cereal Chemistry*, 67(5), 464-470.
- Khan, S., Ghanghro, A.B., Memon, A. N., Tahir, I., Shah, A. M., Sahito, M. A., Talpur, F. N., & Qureshi, S. (2013). Quantitative Analysis of Wheat Proteins in Different Varieties Grown In Sindh, Pakistan. *Intl J Agri Crop Sci.*, 5(16), 1836-1839.
- Koerner, T. B., Cleroux, C., Poirier, C., Cantin, I., Alimkulov, A., & Elamparo, H. (2011). Gluten contamination in the Canadian commercial oat supply. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 28(6), 705-710. <https://doi.org/10.1080/19440049.2011.579626>
- Levi, I., & Anderson, J. A. (1950). Variations in Protein Contents of Plants, Heads, Spikelets, and Individual Kernels of Wheat. *Canadian Journal of Research, F*, 28, 71-81. <https://doi.org/10.1139/cjr50f-006>
- Liu, K. (2008). Measurement of Wheat Hardness by Seed Scarifier and Barley Pearler and Comparison with Single-Kernel Characterization System. *Cereal Chemistry*, 85(2), 165-173. <https://doi.org/10.1094/CCHEM-85-2-0165>
- Mejias, J., Lu, X., Osorio, C., Ullman, J. L., von Wettstein, D., & Rustgi, S. (2014). Analysis of Wheat Prolamins, the Causative Agents of Celiac Sprue, Using Reversed Phase High Performance Liquid Chromatography (RP-HPLC) and Matrix-Assisted Laser Desorption Ionization Time of Flight Mass Spectrometry (MALDI-TOF-MS). *Nutrients*, 6, 1578-1597. <https://doi.org/10.3390/nu6041578>
- R-Biopharm, AG. (2012), Ridascreen Gliadin Art. No. R7001 Validation Report.
- Rallabhandi, P., Sharma, G. M., Pereira, M., & Williams, K. M. (2015). Immunological Characterization of the Gluten Fractions and Their Hydrolysates from Wheat, Rye and Barley. *J. Agric. Food Chem.*, 63, 1825-1832. <https://doi.org/10.1021/jf505716p>
- Seilmeier W, Belitz, H-D, & Wieser, H. (1991). Separation and quantitative determination of high-molecular-weight subunits of glutenin from different wheat varieties and genetic variants of the variety Siccio. *Zeitschrift fu r Lebensmittel-Untersuchung und Forschung*, 192, 124-129. <https://doi.org/10.1007/BF01202625>
- Shewry, P. R. (2009). Wheat. *Journal of Experimental Botany*, 60, 1537-1553. <https://doi.org/10.1093/jxb/erp058>
- Thompson, T. (2004). Gluten Contamination of Commercial Oat Products in the United States. *New England Journal of Medicine*, 351(19), 2021-2022. <https://doi.org/10.1056/NEJM200411043511924>
- Thompson, T., Lee, A. R., & Grace, T. (2010). Gluten contamination of grains, seeds, and flours in the United States: a pilot study. *J Am Diet Assoc*, 110(6), 937-940. <https://doi.org/10.1016/j.jada.2010.03.014>
- Weiser, H., Koehler, P. (2009). Is the calculation of the gluten content by multiplying the prolamins content by a factor of 2 valid?, *Euro. Food Res. & Tech.*, 229(1), 9-13. <https://doi.org/10.1007/s00217-009-1020-5>
- Zilic, S., Barac, M., Pesic, M., Dodig, D., & Ignjatovic-Micic, D. (2011). Characterization of Proteins from Grain of Different Bread and Durum Wheat Genotypes. *Int. J. Mol. Sci.*, 12, 5878-5894. <https://doi.org/10.3390/ijms12095878>

## Copyrights

Copyright for this article is retained by the author(s), with first publication rights granted to the journal.

This is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).