

Factors Influencing Aflatoxin Contamination in Before and After Harvest Peanuts: A Review

Enjie Diao¹, Haizhou Dong¹, Hanxue Hou¹, Zheng Zhang¹, Ning Ji¹ & Wenwen Ma¹

¹ College of Food Science & Engineering, Shandong Agricultural University, Taian 271018, PR China

Correspondence: Haizhou Dong, College of Food Science & Engineering, Shandong Agricultural University, Taian 271018, PR China. Tel: 86-538-824-2850. E-mail: haizhoudong@126.com

Received: October 29, 2014 Accepted: November 18, 2014 Online Published: December 29, 2014

doi:10.5539/jfr.v4n1p148

URL: <http://dx.doi.org/10.5539/jfr.v4n1p148>

Abstract

Aflatoxin contamination of peanuts has been of worldwide concern since the 1960s. Aflatoxins are fungal secondary metabolites that have been associated with severe toxicological effects to human and animals, which can cause enormous economic loss to producers, handlers, processors and marketers of contaminated peanuts. Aflatoxin contamination of peanuts is unavoidable due to the varied factors in pre-harvest, harvesting, and post-harvest stages of peanuts. The review summarizes the factors influencing aflatoxin contamination in the whole supply chain of peanuts.

Keywords: aflatoxins, influencing factors, peanut, pre-harvest, post-harvest

1. Introduction

Peanut (*Arachis hypogaea* L.) is an important crop in China and other countries in the world due to its nutritional and economic values. About 39.81 million metric tons per year of peanuts are produced in the world, and China and Argentina are the world's largest producer and exporters, respectively (USDA, 2013). In China, peanuts are consumed mainly as whole seeds and pressed oil, which contains 25 to 32% protein and 42 to 52% oil (Woodroof, 1983).

However, peanut is one of the most susceptible host crops to *Aspergillus flavus* resulting in aflatoxin contamination. Aflatoxins have attracted worldwide attention due to their toxicological effects to human and animals. They can cause development defects and immune system suppression, cancer, and even death in severe acute exposure (Shenasi et al., 2002; Proctor et al., 2004; Nakai et al., 2008; Bhat et al., 2010; Michael et al., 2010). *Aspergillus flavus* and *Aspergillus parasiticus* are the major producers of aflatoxins (Richard & Payne, 2003; Klich, 2007; Varga et al., 2011). Aflatoxin contamination in peanuts can occur in the field before harvest, during harvesting, or after harvest, and be affected by many factors. The preharvest factors include peanut cultivars, soil type, species of fungi, climate, weather conditions, agricultural practices, phytoalexin production, water activity and maturity of peanuts; the optimum harvest time and timely drying of peanuts are the main factors during harvesting; the storage and transportation conditions are the main factors resulting in the aflatoxin contamination after harvesting peanuts (Dorner et al., 1989; Cotty & Jaime-Garcia, 2007).

2. Pre-Harvest Factors Influencing Aflatoxin Contamination of Peanuts

2.1 Peanut Cultivars

In the 1980s, many researchers had attempted to find peanut cultivars resistant to *Aspergillus flavus* contamination and aflatoxin formation but failed due to presenting the intricate factors influencing the growth and distribution of the fungus and aflatoxin formation (Blankenship et al., 1985; Azaizeh et al., 1989). In recent years, transgenic technology has been widely used for cultivar improvement. Transgenic peanuts containing the Bt (*Bacillus thuringiensis*) gene had significantly lower levels of aflatoxin than non-Bt peanuts in preliminary analysis of log-transformed data (Ozias-Akins et al., 2002). Guo et al. (2008) identified the resistance-related genes (*iso ara h3* and *LEA 4*) in peanut against *Aspergillus parasiticus* infection and subsequent aflatoxin contamination, and then developed a peanut microarray to identify candidate genes that confer resistance to *Aspergillus flavus* infection (Guo et al., 2011). Additionally, cultivar improvement in increasing the resistance of peanut to diseases, can also significantly reduce the incidence of fungal contamination compared to the unimproved varieties (Mutegei et al., 2012).

2.2 Soil Type

It is well known that peanuts can grow in different soil types, such as light sandy soil and heavier soils. Light sandy soil benefits for the rapid proliferation of *Aspergillus flavus*, particularly under dry conditions in the later growth period. On the contrary, heavier soil can reduce the level of aflatoxin contamination in peanut grown because of having a higher water-holding capacity (CAC, 2004; Torres et al., 2014).

2.3 Species of Fungi in Soil

Soil is a reservoir of varied microorganisms including fungi, and peanuts are in direct contact with soil populations of aflatoxigenic fungi (Horn & Pitt, 1997). Common fungal contaminants of peanuts include *Aspergillus*, *Penicillium*, *Rhizopus* and *Fusarium* species (Gachomo et al., 2004; Youssef et al., 2008). Many literatures reported that *Aspergillus flavus* and *Aspergillus parasiticus* are the two closely related species of fungi that invade peanuts and subsequently lead to their contamination with aflatoxins B₁, B₂, G₁ and G₂ (Dorner, 2002; Vaamonde et al., 2003; Mutegi et al., 2012). Presence of other fungi such as *Penicillium* and *Fusarium* species, reduces the aflatoxin production due to competitive inhibition (Horn & Dorner, 1998). Additionally, different morphological types for the same species also affect the aflatoxin contamination of peanuts, such as *Aspergillus flavus*, which has two types, the S- and L-strains. The incidence of *Aspergillus flavus* S-strain has a strong relationship with the peanut contamination with aflatoxin, while L-strain was not positively correlated with the levels of total aflatoxins in peanuts (Mutegi et al., 2012).

2.4 Climate

Marsh and Taylor (1958) reported that aflatoxin contamination was occurred in most growing areas but the highest incidence of aflatoxin was in the warmer, more humid growing regions and followed the same geographical pattern. *Aspergillus flavus* can be isolated from soil in all climatic zones, and it is isolated relatively more frequently in warm temperature zones (latitudes 26-35°) than in tropical or cooler temperature zones. It is quite rare in latitudes above 45° (Manabe & Tsuruta, 1978; Klich, 2002; CAST, 2003). Therefore, the aflatoxin contamination of peanuts is often found in latitudes below 35° (Logrieco & Visconti, 2004).

2.5 Weather Conditions

Sanders and coworkers (1985) found that aflatoxin contamination is not always directly correlated with the incidence of invasion by *Aspergillus flavus*. Cole and coworkers (1985) suggested that after the invasion of aflatoxigenic fungi occurred, growth of the fungi and aflatoxin production could not occur until a natural resistance mechanism broke down as a result of environmental stress (drought and high temperature). Drought and temperature stress are common factors for aflatoxin contamination of peanuts (Cole et al., 1984; Sanders et al., 1993; Craufud et al., 2006). Cole and coworkers (1984) reported that drought stress and soil temperature of 29 °C for 85-100 days yielded the greatest number of colonized edible grade peanuts and high aflatoxin levels. End of season drought stress and elevated soil temperature are more benefit for promoting aflatoxin contamination (Rachaputi et al., 2002; Bankole et al., 2006). The reason is that drought stress induces a great increase in proline in plants, which can enhance aflatoxin production (Barnett & Naylor, 1966; Payne & Hagler, 1983). Therefore, adequate rainfall can control or reduce aflatoxin production of peanuts. In addition, contamination has been found to be widespread where peanuts are grown under rain-fed conditions compared to those grown under irrigation (Reddy et al., 2003).

2.6 Agricultural Practices

Improper agricultural practices, including crop rotation, tillage, planting date, and irrigation and fertilization, can also increase the incidence of *Aspergillus Flavi* and aflatoxin contamination in peanuts (Torres et al., 2014).

The continued cultivation of peanuts on the same land may result in the high infection of fungi and aflatoxin contamination (Ortiz et al., 2011). Crop rotation may lower the rate of between-season survival of different species/strains, especially if it involves crops that are non-host to *Aspergillus* species (Middleton et al., 1994; Mutegi et al., 2012). However, the effects of crop rotation on aflatoxin contamination depend on the planting environment, for example, in a semi - arid environment, populations of *Aspergillus* may be very high, and crop rotation may have little influence on the fungal activity (CAC, 2004).

In noninoculated, non-insecticide-sprayed areas, dense population of plants or reduced fertilization, appear to have a positive influence on the incidence of contamination by aflatoxin (Anderson et al., 1975).

Insects might play an important role in the aflatoxin contamination of crops since nearly all aflatoxins were found in areas damaged by insects. The insect damage peanut tissue, thereby creating entry portals for the fungus, and then lead to the high aflatoxin production in peanuts (Anderson et al., 1975; Waliyar et al., 2008). Many

literatures reported that some insecticides and fungicides can inhibit aflatoxigenic fungi growth and subsequent aflatoxin biosynthesis in field (Dorner et al., 1992; Bowen & Mack, 1993; D'Mello et al., 1998; Lee et al., 2001; Dorner et al., 2003). Bowen and Mack (1993) used the insecticide to treat peanuts during cultivation, and reduced the levels of *Aspergillus flavus* infection and aflatoxin contamination.

In addition, early-sowing in optimum time, rational planting, scientific fertilizing, strong field management were needed in reducing the aflatoxin contamination of peanuts.

2.7 Phytoalexin Production

Phytoalexins are antimicrobial substances synthesized by plants that accumulate rapidly at areas of pathogen infection. Although the chemical nature of the phytoalexins was not determined, it was shown that peanuts produced phytoalexins when challenged by several species of fungi, including *A. flavus*. (Keen, 1975; Ingham et al., 1976; Keen et al., 1976; Aguamah et al., 1981; Wotton & Strange, 1985). It was verified as early as 1972 that the resistance of immature peanut pods to fungi was due to phytoalexins produced in high quantities in response to fungal infection (Vidhyasekaran et al., 1972). As long as peanuts had the capacity for phytoalexin production, they were not contaminated with aflatoxins (Dorner et al., 1989). In immature peanuts, aflatoxin did not form until phytoalexin production ceased in drought stressed plants (Dorner et al., 1989).

Water activity (a_w) of peanut kernels appeared to be the most important factor controlling the capacity of kernels to produce phytoalexins. Peanuts produce sufficient phytoalexins at high a_w (>0.97) to inhibit growth of *Aspergillus flavus* and subsequent aflatoxin contamination. As kernel a_w decreases, as a result of prolonged drought, the capacity of those kernels to produce phytoalexins also decreases and eventually is lost ($a_w < 0.95$) (Dorner et al., 1989).

2.8 Maturity

Preharvest contamination appears first, and is more concentrated in smaller, immature kernels. Several studies have shown consistently that levels of both *Aspergillus flavus* invasion and aflatoxin contamination are higher in smaller, more immature kernels than in mature kernels (Sanders et al., 1981; Cole et al., 1982; Hill et al., 1983; Cole et al., 1985; Sanders et al., 1985). Immature peanuts lose the capacity to produce phytoalexins faster than that of mature ones, leading to earlier contamination. In addition, the mature peanuts have sustained resistance to aflatoxin contamination well, even after the disappearance of phytoalexin production (Dorner et al., 1989).

3. Factors During Harvesting

Aflatoxin contamination can occur if peanuts are not timely harvested, or mechanically harvested (Heathcote & Hibbert, 1978; Cole et al., 1995; Torres et al., 2014). The harvest time depends on the maturity of peanuts and weather conditions during harvesting. Overmature or immature pods at harvest can all lead to the high contamination of aflatoxins in the final product. Harvesting in the dry weather, aflatoxin contamination does not occur but often present in very humid weather (Kabak et al., 2006). Mechanical damage to kernels during digging and threshing peanuts makes them much more vulnerable to the invasion of molds and subsequent aflatoxin contamination (Heathcote & Hibbert, 1978).

4. Post-Harvest Factors Influencing Aflatoxin Contamination of Peanuts

Generally, kernel moisture contents of 10% or higher post-harvest peanuts are prone to producing aflatoxins. Timely drying and maintaining at safe moisture level can effectively control aflatoxin contamination of peanuts after harvest (Torres et al., 2014). Diener and Davis (1970) found that aflatoxin production can be prevented by rapidly drying to or below a a_w of 0.83 for post-harvest peanuts. Before-storage segregating to remove contaminated peanuts is the most effective way to reduce aflatoxin production (Cole et al., 1995; Dorner, 2008).

To prevent an increase in aflatoxin contamination occurring during storage and transportation, it is important to control the moisture content, the temperature in the environment, and the hygienic conditions (Dickens, 1977; CAC, 2004). Inappropriate kernel moisture during storage can proceed from leaky roofs, condensation because of improper ventilation in the warehouse, high-moisture foreign material associated with stored peanuts, and high-moisture peanuts initially going into storage (Davidson et al., 1982). Therefore, the storage and transportation conditions are the most important factors in controlling aflatoxin contamination of peanuts.

5. Conclusion

Aflatoxin contamination of peanuts is a worldwide problem, which poses a risk to human health and has been identified as a major constraint to trade in the world. It is not avoided due to the effects of interactions among many factors. These factors present in pre-harvest, during harvesting, and post-harvest processes, especially pre-harvest management and storage conditions of post-harvest are the main factors. These factors can vary

considerably from one location to another, and between seasons in the same location. Some environments may be particularly favorable to fungal infection and subsequent aflatoxin contamination of peanuts, and in these circumstances it would be necessary to consider whether or not the peanuts should be grown in such areas.

According to the factors influencing aflatoxin contamination of peanuts before and after harvest stages, the corresponding strategies to prevent the growth of aflatoxigenic fungi as well as to inhibit aflatoxin biosynthesis have been studied. For these strategies, only one strategy can not completely prevent and control aflatoxin contamination of peanuts, it is necessary to use two or more methods in the whole supply chain of peanuts based on the guide of total quality management.

Since aflatoxin contamination of peanuts can not be prevented completely, it is necessary to develop new methods for detoxification in process. The research in detoxification of aflatoxins in peanuts is till a hotspot, and physical, chemical, and biological methods have been used to remove or degrade aflatoxins in foods. Presently, there is no one method to effectively degrade aflatoxins and maintain the quality of treated products. In addition, aflatoxin risk forecasting based on the influencing factors is also a hotspot in the future study, which can assist in preventing and minimizing pre-harvest contamination and especially in optimizing harvest post-harvest management.

Conflict of Interest

The authors declare that there are no conflicts of interest.

Acknowledgements

The authors wish to thank to the big data plan of agriculture in Shandong Agricultural University, and the Agricultural Technology Innovation Project in Shandong Province for giving the opportunity to prepare this review article.

References

- Aguamah, G. E., Langcake, P., Leworthy, D. P., Page, J. A., Pryce, R. J., & Strange, R. N. (1981). Two novel stilbene phytoalexins from *Araehis hypogaea*. *Phytochemistry*, *20*, 1381-1383. [http://dx.doi.org/10.1016/0031-9422\(81\)80044-1](http://dx.doi.org/10.1016/0031-9422(81)80044-1)
- Anderson, W. F., Holbrook, C. C., Wilson, D. M., & Matheron, M. E. (1995). Evaluation of preharvest aflatoxin contamination in several potentially resistant peanut genotypes. *Peanut Science*, *22*, 29-32. <http://dx.doi.org/10.3146/pnut.22.1.0007>
- Azaizeh, H. A., Pettit, R. E., Smith, O. D., & Taber, R. A. (1989). Reaction of peanut genotypes under drought stress to *Aspergillus flavus* and *A. parasiticus*. *Peanut Science*, *16*, 109-113. <http://dx.doi.org/10.3146/i0095-3679-16-2-12>
- Bankole, S., Schollenberger, M., & Drochner, W. (2006). Mycotoxins in food systems in Sub Saharan Africa: a review. *Mycotoxin Research*, *22*, 163-169. <http://dx.doi.org/10.1007/BF02959270>
- Barnett, N. M., & Naylor, A. W. (1966). Amino acid and protein metabolism in Bermuda grass during water stress. *Plant Physiology*, *41*, 1222-1230. <http://dx.doi.org/10.1104/pp.41.7.1222>
- Bhat, R., Rai, R. V., & Karim, A. A. (2010). Mycotoxins in food and feed: present status and future concerns. *Comprehensive Reviews in Food Science and Food Safety*, *9*, 57-81. <http://dx.doi.org/10.1111/j.1541-4337.2009.00094.x>
- Blankenship, P. D., Cole, R. J., & Sanders, T. H. (1985). Comparative susceptibility of four experimental peanut lines and the cultivar Florunner to preharvest aflatoxin contamination. *Peanut Science*, *12*, 70-72. <http://dx.doi.org/10.3146/pnut.12.2.0006>
- Bowen, K. L., & Mack, T. P. (1993). Relationship of damage from the lesser cornstalk borer to *Aspergillus flavus* contamination in peanuts. *Journal of Entomological Science*, *28*, 29-42.
- CAST. (2003). Mycotoxins: Risks in Plant, Animal, and Human Systems. Report 139. Ames, IA: Council for Agricultural Science and Technology.
- Codex Alimentarius Commission–CAC. (2004). Code of Practice for the prevention and reduction of aflatoxin contamination in peanuts. CAC/RCP 55.
- Cole, R. J., Blankenship, P. D., Hill, R. A., & Sanders, T. H. (1984). Effect of geocarposphere temperature on preharvest colonisation of drought stressed peanuts by *Aspergillus flavus* and subsequent aflatoxin contamination. In H. Kurata & Y. Ueno (Eds), *Toxigenic Fungi–Their Toxins and Health Hazard* (pp. 44-52). Tokyo: Kodansha Ltd.

- Cole, R. J., Hill, R. A., Blankenship, P. D., Sanders, T. H., & Garren, K. H. (1982). Influence of irrigation and drought stress on invasion of *Aspergillus flavus* of corn kernels and peanut pods. *Developments in Industrial Microbiology*, *23*, 229-236.
- Cole, R. J., Dorner, J. W., & Holbrook, C. C. (1995). Advances in mycotoxin elimination and resistance. In H. E. Pattee & H. T. Stalker (Eds.), *Advances in peanut science* (pp. 456-474). Stillwater, OK, USA: American Peanut Research Educational Society.
- Cole, R. J., Sanders, T. H., Hill, R. A., & Blankenship, P. D. (1985). Mean geocarposphere temperatures that induce preharvest aflatoxin contamination of peanuts under drought stress. *Mycopathologia*, *91*, 41-46. <http://dx.doi.org/10.1007/BF00437286>
- Cotty, P. J., & Jaime-Garcia, R. (2007). Influences of climate on aflatoxin producing fungi and aflatoxin contamination. *International Journal of Food Microbiology*, *119*, 109-115. <http://dx.doi.org/10.1016/j.ijfoodmicro.2007.07.060>
- Craufurd, P. Q., Prasad, P. V. V., Waliyar, F., & Taheri, A. (2006). Drought, pod yield, preharvest *Aspergillus* infection and aflatoxin contamination on peanut in Niger. *Field Crops Research*, *98*, 20-29. <http://dx.doi.org/10.1016/j.fcr.2005.12.001>
- Davidson, J. I., Hill, R. A., Cole, R. A., Mixoon, A. C., & Henning, R. U. (1982). Field performance of two peanut cultivars relative to resistance to invasion by *A. flavus* and subsequent aflatoxin contamination. *Proceedings of the American Peanut Research and Education Society*, *14*, 74-78.
- D'Mello, J. P. F., Macdonald, A. M. C., Postel, D., Dijksma, W. T. P., DuJardin, A., & Placinta, C. M. (1998). Pesticide use and mycotoxin production in *Fusarium* and *Aspergillus* phytopathogens. *European Journal of Plant Pathology*, *104*, 741-751. <http://dx.doi.org/10.1023/A:1008621505708>
- Dickens, J. W. (1977). Aflatoxin control program for peanuts. *Journal of American Oil Chemical Society*, *54*, 225-228. <http://dx.doi.org/10.1007/BF02894413>
- Diener, U. L., & Davis, N. D. (1970). Limiting temperature and relative humidity for aflatoxin production by *Aspergillus flavus* in stored peanuts. *Journal of American Oil Chemical Society*, *47*, 347-351. <http://dx.doi.org/10.1007/BF02639000>
- Dorner, J. W. (2008). Management and prevention of mycotoxins in peanuts. *Food Additives and Contaminants*, *25*, 203-208. <http://dx.doi.org/10.1080/02652030701658357>
- Dorner, J. W. (2002). Simultaneous quantitation of *Aspergillus flavus* / *A. parasiticus* and aflatoxin in peanuts. *Journal of AOAC International*, *85*, 911-916.
- Dorner, J. W., Cole, R. J., & Blankenship, P. D. (1992). Use of a bio-competitive agent to control preharvest aflatoxin in drought stressed peanut. *Journal of Food Protection*, *55*, 888-892.
- Dorner, J. W., Cole, R. J., Connick, W. J., Daigle, D. J., Mcguire, M. R., & Shasha, B. S. (2003). Evaluation of biological control formulations to reduce aflatoxin contamination in peanuts. *Biological Control*, *26*, 318-324. [http://dx.doi.org/10.1016/S1049-9644\(02\)00139-1](http://dx.doi.org/10.1016/S1049-9644(02)00139-1)
- Dorner, J. W., Cole, R. J., Sanders, T. H., & Blankenship, P. D. (1989). Interrelationship of kernel water activity, soil temperature, maturity, and phytoalexin production in preharvest aflatoxin contamination of drought-stressed peanuts. *Mycopathologia*, *105*, 117-128. <http://dx.doi.org/10.1007/BF00444034>
- Gachomo, E. W., Mutitu, E. W., & Kotchoni, O. S. (2004). Diversity of fungal species associated with peanuts in storage and the levels of aflatoxins in infected samples. *International Journal of Agriculture and Biology*, *6*, 955-959.
- Guo, B. Z., Chen, X. P., Dang, P., Scully, B. T., Liang, X. Q., Holbrook, C. C., & Yu, J. J. (2008). Peanut gene expression profiling in developing seeds at different reproduction stages during *Aspergillus parasiticus* infection. *BMC Developmental Biology*, *4*, 8-12.
- Guo, B. Z., Fedorova, N. D., Chen, X. P., Wan, C. H., Wang, W., Nierman, W. C., ... Yu, J. J. (2011). Gene expression profiling and identification of resistance genes to *Aspergillus flavus* infection in peanut through EST and microarray strategies. *Toxins*, *3*, 737-753. <http://dx.doi.org/10.3390/toxins3070737>
- Heathcote, J. G., & Hibbert, J. R. (1978). *Aflatoxin chemical and biological aspects*. Elsevier Scientific Publishing Company, Amsterdam.
- Hill, R. A., Blankenship, P. D., Cole, R. J., & Sanders, T. H. (1983). Effect of soil moisture and temperature on preharvest invasion of peanuts by the *Aspergillus flavus* group and subsequent aflatoxin development. *Applied and Environmental Microbiology*, *45*, 628-633.

- Horn, B. W., & Dorner, J. W. (1998). Soil populations of *Aspergillus* species from section *Flavi* along a transect through peanut growing regions of the United States. *Mycologia*, *90*, 767-776. <http://dx.doi.org/10.2307/3761317>
- Horn, B. W., & Pitt, J. I. (1997). Yellow mold and aflatoxin. In N. Kokalis-Burelle D. M., Porter, R., Rodriguez-Kabana D. H. Smith & Subrahmanyam (Eds.), *Compendium of peanut diseases* (pp. 44-49). St. Paul, MN, USA: Am. Phytopathol. Soc.
- Ingham, J. L. (1976). 3,5,4'-trihydroxystilbene as a phytoalexin from groundnuts (*Arachis hypogaea*). *Phytochemistry*, *15*, 1791-1793. [http://dx.doi.org/10.1016/S0031-9422\(00\)97494-6](http://dx.doi.org/10.1016/S0031-9422(00)97494-6)
- Kabak, B., Dobson, A. D. W., & Var, I. (2006). Strategies to prevent mycotoxin contamination of food and animal feed: A review. *Critical Reviews in Food Science and Nutrition*, *46*, 593-619. <http://dx.doi.org/10.1080/10408390500436185>
- Keen, N. T. (1975). The isolation of phytoalexins from germinating seeds of *Cicer arietinum*, *Vigna sinensis*, *Arachis hypogaea*, and other plants. *Phytopathology*, *65*, 91-92. <http://dx.doi.org/10.1094/Phyto-65-91>
- Keen, N. T., & Ingham, J. L. (1976). New stilbene phytoalexins from American cultivars of *Arachis hypogaea*. *Phytochemistry*, *15*, 1794-1795. [http://dx.doi.org/10.1016/S0031-9422\(00\)97495-8](http://dx.doi.org/10.1016/S0031-9422(00)97495-8)
- Klich, M. A. (2007). *Aspergillus flavus*: the major producer of aflatoxin. *Molecular Plant Pathology*, *8*, 713-722. <http://dx.doi.org/10.1111/j.1364-3703.2007.00436.x>
- Klich, M. A. (2002). Biogeography of *Aspergillus* species in soil and litter. *Mycologia*, *94*, 21-27. <http://dx.doi.org/10.2307/3761842>
- Lee, S. E., Campbell, B. C., Molyneux, R. J., Hasegawa, S., & Lee, H. S. (2001). Inhibitory effects of naturally occurring compounds on aflatoxin B₁ biotransformation. *Journal of Agricultural and Food Chemistry*, *49*, 5171-5177. <http://dx.doi.org/10.1021/jf010454v>
- Logrieco, A., & Visconti, A. (2004). *An Overview on Toxigenic Fungi and Mycotoxins in Europe*. Dordrecht Netherlands: Kluwer Academic Publishers. <http://dx.doi.org/10.1007/978-1-4020-2646-1>
- Manabe, M., & Tsuruta, O. (1978). Geographical distribution of aflatoxin-producing fungi inhabiting in southeast Asia. *Japan Agricultural Research Quarterly*, *12*, 224-227.
- Marsh, P. B., & Taylor, E. E. (1958). The geographical distribution of fiber containing fluorescent spots associated with *Aspergillus flavus* in the United States cotton crop of 1957. *Plant Disease Reporter*, *42*, 1368-1371.
- Michael, J. T., John, O. D., Melissa, M. C., Ahmedin, J., & Elizabeth, M. W. (2011). The global burden of cancer: priorities for prevention. *Carcinogenesis*, *31*, 100-110.
- Middleton, K. J., Pande, S. S., Sharma, S. B., & Smith, D. H. (1994). Diseases. In J. S. Smart (Ed.), *Groundnut Crop: A Scientific Basis for Improvement* (pp. 336-378). London: Chapman and Hall. http://dx.doi.org/10.1007/978-94-011-0733-4_10
- Mutegi, C. K., Ngugi, H. K., Hendriks, S. L., & Jones, R. B. (2012). Factors associated with the incidence of *Aspergillus* section *Flavi* and aflatoxin contamination of peanuts in the Busia and Homa bay districts of western Kenya. *Plant Pathology*, *61*, 1143-1153. <http://dx.doi.org/10.1111/j.1365-3059.2012.02597.x>
- Nakai, V., de Oliveira Rocha, L., Goncalvez, E., Fonseca, H., Ortega, E., & Corrêa, B. (2008). Distribution of fungi and aflatoxins in a stored peanut variety. *Food Chemistry*, *106*, 285-290. <http://dx.doi.org/10.1016/j.foodchem.2007.05.087>
- Ortiz, M. P., Barros, G. G., Reynoso, M. M., Torres, A. M., Chulze, S. N., & Ramirez, M. L. (2011). Soil populations of *Aspergillus* section *Flavi* from the main and new peanut growing areas in Argentina. ISM Conference 2011 "Strategies to reduce the impact of mycotoxins in Latin America in a global context". Abstract Book.
- Ozias-Akins, P., Yang, H., Perry, E., Akasaka, Y., Niu, C., Holbrook, C., & Lynch, R. E. (2002). Transgenic peanut for preharvest aflatoxin reduction. *Mycopathologia*, *155*, 98.
- Payne, G. A., & Hagler, W. M. (1983). Effect of specific amino acids on growth and aflatoxin production by *Aspergillus parasiticus* and *Aspergillus flavus* in defined media. *Applied and Environmental Microbiology*, *46*, 805-812.
- Proctor, A. D., Ahmedna, M., Kumar, J. V., & Goktepe, I. (2004). Degradation of aflatoxins in peanut kernels/flour by gaseous ozonation and mild heat treatment. *Food Additives and Contaminants*, *21*, 786-793. <http://dx.doi.org/10.1080/02652030410001713898>

- Rachaputi, N. R., Wright, G. C., & Krosch, S. (2002). Management practices to minimize pre-harvest aflatoxin contamination in Australian groundnuts. *Australian Journal of Experimental Agriculture*, 42, 595-605. <http://dx.doi.org/10.1071/EA01139>
- Reddy, T. Y., Sulochanamma, B. N., Subramanyam, A., & Balaguravaiah, D. (2003). Influence of weather, dry spells and management practices on aflatoxin contamination in groundnut. *Indian Phytopathology*, 56, 262-265.
- Richard, J. L., & Payne, G. A. (2003). Mycotoxins: Risks in plant, animal, and human systems. Task Force Report No. 139. Ames, IA: Council for Agricultural Science and Technol. CAST.
- Sanders, T. H., Cole, R. J., Blankenship, P. D., & Dorner, J. W. (1993). Aflatoxin contamination of peanuts from plants drought stressed in pod or root zones. *Peanut Science*, 20, 5-8. <http://dx.doi.org/10.3146/i0095-3679-20-1-2>
- Sanders, T. H., Cole, R. J., Blankenship, P. D., & Hill, R. A. (1985). Relation of environmental stress duration to *Aspergillus flavus* invasion and aflatoxin production in preharvest peanuts. *Peanut Science*, 12, 90-93. <http://dx.doi.org/10.3146/pnut.12.2.0011>
- Sanders, T. H., Hill, R. A., Cole, R. J., & Blankenship, P. D. (1981). Effect of drought on occurrence of *Aspergillus flavus* in maturing peanuts. *Journal of American Oil Chemical Society*, 58, 966-970. <http://dx.doi.org/10.1007/BF02679302>
- Shenasi, M., Aidoo, K. E., & Candlish, A. A. G. (2002). Microflora of date fruits and production of aflatoxins at various stages of maturation. *International Journal of Food Microbiology*, 79, 113-119. [http://dx.doi.org/10.1016/S0168-1605\(02\)00185-X](http://dx.doi.org/10.1016/S0168-1605(02)00185-X)
- Torres, A. M., Barros, G. G., Palacios, S. A., Chulze, S. N., & Battilani, P. (2014). Review on pre- and post-harvest management of peanuts to minimize aflatoxin contamination. *Food Research International*, 62, 11-19. <http://dx.doi.org/10.1016/j.foodres.2014.02.023>
- USDA. (2013). Foreign agricultural service: Table 13 peanut area, yield, and production.
- Vaamonde, G., Patriarca, A., Pinto, V. F., Comerio, R., & Degrossi, C. (2003). Variability of aflatoxin and cyclopiazonic acid production by *Aspergillus* section *Flavi* from different substrates in Argentina. *International Journal of Food Microbiology*, 88, 79-84. [http://dx.doi.org/10.1016/S0168-1605\(03\)00101-6](http://dx.doi.org/10.1016/S0168-1605(03)00101-6)
- Varga, J., Frisvad, J., & Samson, R. (2011). Two new aflatoxin producing species and an overview of *Aspergillus* section *Flavi*. *Studies in Mycology*, 69, 57-80. <http://dx.doi.org/10.3114/sim.2011.69.05>
- Vidhyasekaran, P., Lalithakumari, D., & Govindaswamy, C. V. (1972). Production of a phytoalexin in groundnut due to storage fungi. *Indian Phytopathology*, 25, 240-245.
- Waliyar, F., Kumar, P. L., Traore, A., Ntare, B. R., Diara, B., & Kodio, O. (2008). Pre and post harvest management of aflatoxin contamination in peanuts. In J. F. Leslie, R. Bandyopadhyay & A. Visconti (Eds.) *Mycotoxins: Detection Methods, Management, Public Health and Agricultural Trade* (pp. 209-218). Trowbridge: Cromwell Press.
- Woodroof, J. G. (1983). *Peanut: Production, processing, products*. Westport, CT: AVI.
- Wotton, H. R., & Strange, R. N. (1985). Circumstantial evidence for phytoalexin involvement in the resistance of peanuts to *Aspergillus flavus*. *Journal of General Microbiology*, 131, 487-494.
- Youssef, M. S., El-Maghraby, O. M. O., & Ibrahim, Y. M. (2008). Mycobiodata and mycotoxins of Egyptian peanut (*Arachis hypogaea* L.) seeds. *International Journal of Botany*, 4, 349-360. <http://dx.doi.org/10.3923/ijb.2008.349.360>

Copyrights

Copyright for this article is retained by the author(s), with first publication rights granted to the journal.

This is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).