Quality Parameters and Bioactive Compounds of Red Tomatoes (Solanum lycopersicum L.) cv Roma VF at Different Postharvest Conditions

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Received: April 18, 2014 Accepted: June 14, 2014 Online Published: June 16, 2014

Abstract

Tomato is one of the most important crops at worldwide; however, much of the production is lost during the postharvest due to the inadequate storage conditions. The aim of this study was to evaluate the effects of different postharvest conditions on some quality and bioactive parameters of tomatoes. Tomatoes Roma VF variety were stored at three temperature (7, 22, and 37 °C) to evaluate some physicochemical characteristics (pH, total soluble solids, titratable acidity, color, and firmness) and bioactive compounds (vitamin C, lycopene, carotenoids, and antioxidant activity) during five days; time in which tomato change from physiological to edible maturity. The *a* color parameter increased significantly (p < 0.05) at the storage temperature of 22 and 37 °C. The firmness was higher in tomato stored at 7 °C after 5 days, while at 37 °C the firmness decrease in 8%. Tomatoes stored at 22 °C showed a decrease of chlorophyll *a* and *b*, reaching the maximum lycopene (31.7 ± 1.5 mg/kg fresh weight) and carotenoids contents (118.7 ± 0.1 μ g/100 g fresh weight), while at 7 and 37 °C the chlorophyll *a* did not change during the storage. However, the vitamin C was higher in tomatoes stored at 7 and 37 °C. The antioxidant activity remains constant during the time of storage, regardless the temperature. The storage temperature of 22 °C not affected significantly (p < 0.05) the color and firmness of tomatoes, at the same temperature, the bioactive compounds (carotenoid, vitamin C, and antioxidant activity) presented highly correlated with the developments of the red color.

Keywords: antioxidant activity, bioactive compounds, temperature, tomatoes

1. Introduction

Tomato (*Solanum lycopersicum* L.) is one of the most important crop vegetable and also one of the most commercialized and industrialized (Ogwulumba, Ugwuoke, & Omemaramadu, 2011). In 2012, the SIAP informed, that tomato production in Mexico reached 2, 838, 369.87 tons, with a national yield of 51.38 ton/ha. Puebla, Mexico had a production around 52.85 tons; however, the yield was 63.58 ton/ha. The SAGARPA in 2009 pointed out, that in the northwest of Puebla the tomato crop under greenhouse was increased during the past decade. This suggests that the greenhouse tomatoes are preferred by the consumers because of its good quality, either if it's sold fresh or processed (Kubota, Thomson, & Javanmardi, 2006).

Tomatoes has been considered an important source of bioactive compounds, such as, vitamins (principally vitamin C and vitamin E), carotenoids (lycopene), and phenolic compounds like flavonoids (Ketsa & Wongveerakhan, 1987; Giovanelli, Lavelli, Peri, & Nobili, 1999; Raffo, Leonardi, Fogliano, Ambrosino, Salucci, & Gennaro, 2002), which provide phytonutrients to the diet that are associated with the prevention of chronic degenerative diseases, like cardiovascular and carcinogenic diseases (Willcox, Catignani, & Lazarus, 2003). According to the FAO (1979) one tomato provides 40% of vitamin C (ascorbic acid) and 20% of vitamin A, of the recommended daily intake; also provides significant amount of potassium, fiber, calcium, iron, and small

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amounts of magnesium, thiamine, riboflavin, and niacin.

Color and firmness are important quality factors of tomato; the red color comes from pigments like carotenoids, which develop during ripening and storage; these compounds are synthesized from chlorophyll degradation (Satyan & Patwardhan, 1983). Moreover, firmness is affected by the storage conditions, postharvest handling, and the enzymatic activity (Kader, Stevens, Albright-Holton, Morris, & Algazi, 1977). Other quality attributes of tomatoes are associated with the physicochemical characteristics, such as, total soluble solids, pH, weight, and organic acids (Kader et al., 1977). During the ripening and storage of tomatoes occur several desirable changes, such as, increasing the flavor and aroma compounds, increasing the ratio between citric to malic acid; and also, the ascorbic acid, phenolic compounds, and flavonoids are increased (Leonardi, Ambrosino, Esposito, & Fogliano, 2000; Toor & Savage, 2006). Nevertheless, if the storage conditions are inadequate the tomatoes could present undesirable characteristics and nutrient composition (Javanmardi & Kubota, 2006).

To extend the shelf life of tomato it is important to reduce the oxidative metabolism by storing at low temperature, in combination with modified atmosphere (especially at high concentration of carbon dioxide). However, storage of tomatoes at critical low temperatures predisposes them to chilling injury (Toor & Savage, 2006). Therefore, the aim of this research was to evaluate the effects of the postharvest conditions on the quality parameters and bioactive compounds of tomato Roma VF variety.

2. Materials and Methods

2.1 Tomato Samples

Tomatoes (*Solanum lycopersicum* L.) cv Roma VF were grown in fertiirrigation greenhouse located in Sierra Norte of Puebla, Mexico, during the fall-winter season, 2012. Tomatoes were sorted free from physical and microbiological damages; the physiological maturity was 5 (light red color) according to color chart of USDA (1997). Tomatoes were recollected (one day before the beginning of the storage), washed with distilled water, and the excess of water was eliminated with absorbent paper.

2.2 Storage Conditions

Tomatoes were randomly sorted and divided in 3 batches. The tomatoes were stored at 7 °C (90% RH) in a refrigerator (TAPPAN EUR251p7w6, Ohio, USA); 22 (40% RH) and 37 °C (60% RH) in a controlled temperature camera (Electro tech systems, inc, model 5518, PA, USA). The quality characteristics and bioactive compounds were evaluated after 1, 3, and 5 days of storage, during this time the tomatoes change from physiological maturity to edible maturity.

2.3 Physicochemical Characteristics

The pH, total soluble solids (°Brix), and titratable acidity (% citric acid) were evaluated according to the 981.12, 932.12, and 942.15 AOAC (2000) methods, respectively.

2.4 Color

Three tomatoes of each storage condition were used to evaluate the L (luminosity, white-black), a (red-green), and b (yellow-blue) color parameters, in the Hunter scale, using a Colorflex M 6405 (Virginia, USA) colorimeter in the reflectance mode. The evaluation was made in the equatorial zone of each tomato (Arias, Lee, Specca, & Janes, 2000).

2.5 Firmness

The firmness was evaluated in the equatorial zone of each tomato using a penetrometer TE model ST32 (FO, Italia). The tomato was compressed 2 mm in depth, using a conical plate at a speed of 5 mm/s (Arias et al., 2000).

2.6 Ascorbic Acid

The ascorbic acid (vitamin C) was evaluated with the ascorbate oxidase activity, according to the Foyer, Rowell and Walker (1983) methodology with modifications. 0.1 g of tomato was mixed with percloric acid (1M) (JT Baker, Avantor Perfomance Materials, PA, USA) during 1 min. The solution was centrifuged at 5000 rpm (4 °C) during 20 min. The supernatant was taken and the pH was adjusting to 4.5 with a buffer solution of sodium phosphate (pH 7.6), the solution was centrifuged again. 200 μ L of the stock solution was mixed with 140 μ L of buffer solution and 20 μ L of DDT (ditioriol 0.077 g/mL) (Sigma-Aldrich, Toluca, Mexico). The mix was stored at room temperature in darkness during 30 min. Moreover, 400 μ L of the stock solution was taken and mixed with 1600 μ L of EDTA solution (0.380 g/100 mL of sodium phosphate buffer) (Fermont, Monterrey, Mexico), and 40 μ L of L-ascorbate oxidase enzyme (Sigma-Aldrich, Toluca, Mexico). The absorbance was measured after

30 min at 265 nm using a UV-Visible Jenway spectrophotometer model 6405 (Staffordshire, UK). The ascorbic acid was calculated using the next equation:

$$AA = [(A_i - A_f) * EC] * MW * 100$$
 (1)

Where AA is the ascorbic acid content (mg of ascorbic acid/100 g of sample), A_i is the initial absorbance, A_f is the final absorbance, EC is the extinct coefficient (12.8), MW is the molecular weight of ascorbic acid (176.1 g/gmol).

2.7 Lycopene

The lycopene concentration was evaluated according to the Sadler and Dezman (1990) methodology. One g of tomato was homogenized with 1 mL of distilled water; then, an aliquot (0.4-0.6 g) of the homogenized was mixed with a solution of hexane/acetone/ethanol (2:1:1) (JT Baker, PA, USA) and BHT 0.05% (w/v) (Sigma-Aldrich, MO, USA), the mixed was stored at room temperature in a dark environment. The samples were agitated to 180 rpm during 30 min, and then 10 mL of distilled water were added. After the total phase's separation (15 min), the absorbance of nonpolar phase was evaluated at 472 nm, using a hexane as blank. The lycopene content was calculated using the next equations:

$$TS = \frac{(ITS*HTS)}{(ITS+1)}$$
 (2)

$$LC = \frac{(Abs*EC)}{TS}$$
 (3)

where TS is the tissue sample (g), ITS is the initial tomato sample (g), HTS is the homogenized tomato sample (g), LC is the lycopene content (mg of lycopene/kg of sample), EC is the extinct coefficient (31.2).

2.8 Chlorophylls and Carotenoids

The carotenoids content was evaluated according to the Lichtenthaler and Wellbum (1983) methodology. One g of tomato pulp was macerated with 5 mL of acetone solution (80% v/v), 2 g of calcium carbonate (Meyer, Edo. de México, México) and 2 g of sea sand; the mixed was centrifuged at 3500 rpm at room temperature during 10 min. The absorbance was evaluated at 470, 645, and 662 nm to evaluate the chlorophyll a and b, and total carotenoid. Acetone was used as a blank and the calculated of carotenoids was using the next equation:

$$C_a = 11.75 \text{Abs}_{667} - 2.350 \text{Abs}_{645}$$
 (4)

$$C_b = 18.61 \text{Abs}_{645} - 3.96 \text{Abs}_{662} \tag{5}$$

$$C_{c} = \frac{1000 \text{Abs}_{470} - 2.27C_{a} - (81.4C_{b})}{227}$$
 (6)

where Abs is the absorbance, Ca is the chlorophyll a, Cb is the chlorophyll b, and Cc is the carotenoid content (µg of carotenoids/100 g of sample).

2.9 Antioxidant Activity

Antioxidant activity was evaluated by the DPPH radical (1,1-diphenyl-2-picrylhydrazyl) according to the Mongkolsilp, Pongbupakit, Sae-Lee, and Sitthithaworn (2004) method. To obtain the tomato extract; six g of tomato pulp (fresh weight) was mixed with 30 mL of methanol: water solution (80:20% v/v). The mixed was agitated at 125 rpm during 12 hours at 40 °C. Two mL of DPPH radical (0.1 mM) (Sigma-Aldrich, MO, USA) was mixed with 500 μ L of extract and was stored for 30 min at darkness. The absorbance was measured at 517 nm using methanol as blank. The inhibition percentages and antioxidant activity [Trolox Equivalent Antioxidant Activity (TEAC)] were calculated using the next equations:

$$I = \frac{(Abs_b - Abs_s)}{Abs_s} *100 \tag{7}$$

$$AA = \left(\frac{I - b}{m}\right) * 100\tag{8}$$

where I is the inhibition (%), Abs_b is the absorbance of blank, and Abs_s is the absorbance of the methanol extract, AA is the amount of Trolox (μ M TEAC/100 g), m is the slope (4084.2 g/ μ M TEAC), and b is the intercept

(1246.6%) of the standard curve $(R^2 = 0.98)$.

2.10 Statistical Analysis

The Microsoft Excel Program (Microsoft Inc. Redmond, WA) was used to calculate averages and Pearson correlations. The analysis of variance (ANOVA) was performed using the Minitab 15 program (Minitab Inc. PA, USA, 2008). Differences between treatments were analyzed by Tukey's pairwise mean comparisons ($\alpha = 0.05$).

3. Results and Discussions

3.1 Physicochemical Characteristics

Table 1 presents the physicochemical characteristics of tomatoes during the storage at different conditions. The total soluble solids were constant during the five days of storage. Wold, Rosenfeld, Holte, Baugerød, Blomhoff, and Haffner (2004) reported similar values of total soluble solids (4.9 to 5.7%). During 8 days of storage at 15.4 to 16.2 °C; Tigist, Workneh, and Woldetsadik (2013) pointed out that total soluble solids of tomatoes Roma VF variety were constant (4.1 to 4.3). Arias et al. (2000) reported similar values of pH (4.0) to those obtained in this study. The pH of tomatoes stored at 7 and 22 °C did not significantly changed (p > 0.05); while at 37 °C, the pH was increased (p < 0.05). Gómez and Camelo (2002) reported that during the storage of tomatoes in controlled atmosphere (12 °C), the pH slightly increased. In general, the titratable acidity was $0.3 \pm 0.0\%$ (citric acid), and was under the reported by other researchers, like Wold et al. (2004); they informed a titratable acidity of 0.44-0.77%. During the storage, titratable acidity of tomatoes stored at 7 °C did not change significantly (p > 0.05). However, in tomatoes stored at 22 and 37 °C, the titratable acidity significantly decreased (p < 0.05) after 5 and 3 days, respectively. Both, pH and titratable acidity are based on organic acids presents in tomatoes; generally the organic acids decrease during the storage, because they are used as substrate in the respiration process, which increases with increasing the temperature of storage (Wills, Lee, Graham, McGlasson, & Halls, 1981).

Table 1. Physicochemical characteristics of tomato during the storage^a

Time (d)	Total	soluble solid	ls (%)		pН		Titratable acidity ^b			
	7°C	22°C	37°C	7°C	22°C	37°C	7°C	22°C	37°C	
0	3.9±1 _a	3.9±1 _a	3.9±1 _a	4.1±0 _a	4.1±0 _a	4.1±0 _a	0.3±0 _a	0.3±0 _a	0.3±0 _a	
1	$3.3\pm0_a$	$3.5\pm0_a$	$3.7\pm0_a$	$4.0{\pm}0_a$	$4.0\pm0_a$	$4.2\pm0_{ab}$	$0.3\pm0_a$	$0.3\pm0_a$	$0.3{\pm}0_a$	
3	$3.9\pm1_a$	$3.8 \pm 1_a$	$3.7\pm1_a$	$4.0\pm0_a$	$4.1\pm0_a$	$4.3\pm0_{ab}$	$0.3\pm0_a$	$0.2\pm0_a$	$0.2\pm0_b$	
5	$3.8\pm0_a$	$3.6 \pm 0_a$	$4.4\pm1_a$	$4.2\pm0_a$	$4.1 \pm 0_a$	$4.4\pm0_{b}$	$0.3\pm0_a$	$0.2\pm0_b$	$0.2\pm0_b$	

^aAverage (n = 3) \pm Standard deviation. Different letters within the same column indicate significant difference (p < 0.05).

3.2 Color

Figure 1 presents the L, a, and b color parameters of tomatoes during the storage at different conditions. The L, a, and b color parameters at the beginning of the storage were 37 ± 2 , 21.7 ± 2.5 , and 24 ± 1 , respectively. Similar values were reported by Arias et al. (2000) from tomato cv. Laura. They informed that the L, a, and b values were 41 ± 0.4 , 22 ± 0.5 , and 24 ± 0.4 , respectively. De Souza, Scalon, Chitarra, and Chitarra (1999) pointed out that in tomatoes the L and a color parameters were related to lycopene concentration; however, the correlation of lycopene with b value was poorly. The L color parameter did not significantly changed (p > 0.05) during the storage at different temperatures. However, increasing the storage temperature reduces the luminosity (L), which indicating a darkening of tomatoes. The most important color parameter in tomato is the a (red-green) color, which significantly increased (p < 0.05) at 22 °C, this could be due to the ripeness and senescence process of the fruit and the increased synthesis of lycopene; at storage temperatures of 7 and 37 °C did not exist changes (p > 0.05) of the a color. Žnidarčič, Dan, Oplanić, and Karić (2010) pointed out that during the storage of tomato (cv. Belle), red color showed higher increase with increasing the storage temperature. The b color parameter significantly (p < 0.05) increased at 7 °C, which indicate an increase of yellow color; this could be probably due to the chilling injury (De Castro, Vigneault, Charles, & Cortez, 2005) or due to the synthesis of flavonoids pigments, like quercentin (Crozier, Lean, McDonald, & Black, 1997). The total color change was not

b% citric acid.

significantly (p > 0.05) affected by the storage temperature; at the end of the storage, the total color change in tomatoes were 5.8 ± 1.3 , 6.1 ± 1.9 , and 5.6 ± 2.0 to the storage temperature of 7, 22, and 37 °C, respectively.

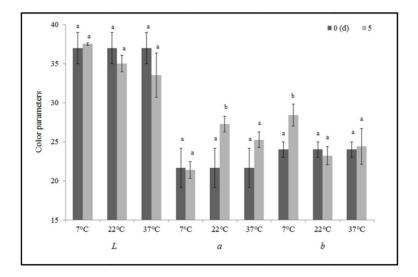


Figure 1. Change of color parameters in tomato during the storage at different conditions

3.3 Firmness

The firmness and crispness in vegetables are critical quality factors (Ochoa-Velasco & Guerrero-Beltrán, 2014); during the postharvest is very important that the storage conditions are adequate to maintain the firmness of tomatoes. During the ripening and storage, the loss of firmness in tomatoes is because the actions of different enzymes like cellulase, pectinesterase, and polygalacturonase on cell wall, media lamella, and plasmatic membrane (Tucker, Robertson, & Grierson, 1980; García & Barrett, 2002). The polygalacturonase is the principal responsible of the softening in tomatoes due to this enzyme is synthesized of novo and increases during the ripening. Figure 2 presents the firmness of tomatoes during the storage at different conditions. It was observed that increasing the temperature of storage decrease the firmness of tomato. This is probably because increasing the storage temperature increases the metabolism of tomatoes, accelerating senescence process such as the loss of firmness (Peet & Bartholemew, 1996). Moreover, the synthesis of polygalacturonase could be diminished at low temperature (Aguayo, Escalona, & Artés, 2004). De Castro et al. (2005) reported that during the storage (8 days) of tomato cv Santa Clara at 14 and 24 °C, the firmness were 7.4 and 5.0 N, respectively.

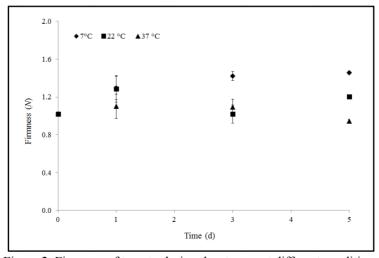


Figure 2. Firmness of tomato during the storage at different conditions

3.4 Ascorbic Acid

Table 2 presents the vitamin C (ascorbic acid) contents of tomatoes during the storage at different conditions. At the beginning of the storage, the vitamin C content was 35.3 ± 1.3 mg/100 g of fresh weight (FW). During the storage it was observed a significantly increase (p < 0.05) of the vitamin C in tomatoes stored at 7 and 37 °C, while in tomatoes stored at 22 °C the vitamin C decreased significantly (p < 0.05) at day 5 of storage. Palop, Özdikicierler, Köstekli, Escriva, Esteve, and Frígola (2010) pointed out that during the refrigeration storage of tomatoes (Pera variety), the vitamin C was increased (20%) for 6 days of storage. Zapata, Gerard, Davies, & Schvab (2007) reported that the synthesis of vitamin C in tomatoes were higher in those grown in the months of highest average temperature. Moneruzzaman, Hossain, Sani, and Saifuddin (2008) reported that the ascorbic acid decreased with the ripening of tomato fruits; also during the storage at 30 ± 1 °C the ascorbic acid decreased in 20% after 6 days of storage. It is known that ascorbic acid decreases with increase in temperature (Emese & Nagymate, 2008); however, Oyetade, Oyeleke, Adegoke, and Akintunde (2012) performed an experiment of the stability of ascorbic acid from different sources (laboratory grade, pharmaceutical tablets, and grape juice) storage at different temperature (4-5 °C, room temperature, and 35 °C). They reported that the reduction of ascorbic acid was higher at room temperature in grape juice.

3.5 Lycopene

Table 2 shows the lycopene content of tomatoes stored at different condition. At the beginning of the storage, the lycopene content was 24.5 ± 2.8 mg/ kg of fresh weight (FW). Similar results $(2.8 \pm 0.12 \text{ mg/100 g})$ were obtained by Toor and Savage (2005) for the pulp of tomato; however, they found that the highest content of lycopene was in tomato skin $(8.7 \pm 1.1 \text{ mg/100 g})$. According to Shi and Le Maguer (2000) the lycopene constitutes the 80-90% of the total pigments present in tomatoes. During the storage it was observed a significantly (p < 0.05) increase of lycopene content in tomatoes stored at 22 °C (31.7 ± 1.5). While at 7 and 37 °C the lycopene content increased at the beginning of the storage (1 day) but decreased at the end. The synthesis of lycopene can be affected by the temperature; at temperature below to 12 °C and above of 32 °C the lycopene precursors are inhibited (Abushita, Hebshi, Daood, & Biacs, 1997). Fraser, Truesdale, Bird, Schuch, and Bramley (1994) pointed out that during the ripening of tomato the pigments change from xanthophylls in green fruits to lycopene, phytoene, fitofueno, β , and γ carotenes in mature fruits. However, during the ripening, the highest concentration of pigments are found in the outermost tissues and decreased in the internal tissue (López-Casado, Matas, Cuartero, Heredia, & Romero-Aranda, 2003).

3.6 Chlorophylls and Carotenoids

The carotenoids have a lot of biological function in human body, such as provitamin A activity, protect low density lipoproteins (LDL) against oxidation, scavengers activity against free radicals, tumor-suppressive activity, carcinogenesis, and protection of DNA against peroxidation (Voutilainen, Nurmi, Mursu, & Rissnaen, 2006). However, only provitamin A activity has been demonstrated in the physiology of humans. Table 2 presents the chlorophylls a and b, and the total carotenoids content of tomatoes during the storage at different conditions. It was observed that the chlorophyll a and b decreased during the storage, being significantly (p < 0.05) at 22 and 37 °C. During the storage, the total carotenoids increased with a concomitant decrease in chlorophylls. However, at the storage temperature of 7 °C, the synthesis of carotenoids from chlorophylls was limited (1 day) by the temperature. At the end of the storage, the total carotenoids content was significantly higher (p < 0.05) in tomatoes stored at 22 °C. During the ripening, the total carotenoids increased with a decrease of chlorophyll, reached the highest content of carotenes when the fruit was overripe (Fraser et al., 1994). It is important to note that the tomatoes storage at 22 °C presents the higher transformation of chlorophyll a and a in carotenoids, which indicate that the carotenoids production in tomatoes is affected by the temperature of storage (Arias et al., 2000).

Table 2. Chlorophyll a and b, lycopene, vitamin C, and total carotenoids contents in tomato during the storage^a

	Chlorophyll a		Chlorophyll b			Lycopene			Vitamin C			Total carotenoids			
Time (d)	$(\mu g/100g FW)$		$(\mu g/100g FW)$			(mg/kg FW)			(mg/100g FW)			$(\mu g/100g FW)$			
	7°C	22°C	37°C	7°C	22°C	37°C	7°C	22°C	37°C	7°C	22°C	37°C	7°C	22°C	37°C
0	63.2 _b	63.2 _a	63.2 _a	112.1 _b	112.1 _a	112.1 _a	24.5 _{ab}	24.5 _c	24.5 _c	35.3 _b	35.3 _a	35.3 _b	39.2 _b	39.2 _d	39.2 _c
1	91.0_a	33.8_{c}	44.2 _b	176.4 _a	47.5_{b}	77.3_{b}	28.2_a	21.0_{c}	50.3_{a}	32.3_b	32.7_{a}	28.7_{b}	33.4_b	61.1_{c}	58.8_{b}
3	83.2 _a	48.8_{b}	44.2 _b	110.7_{b}	57.1 _b	71.9_{b}	22.4_{ab}	48.7_{a}	41.2 _b	36.6_b	36.7_{a}	42.8_{a}	99.0_a	81.6_{b}	86.9 _a
5	53.7 _b	23.8_{c}	44.0_{b}	$51.0_{\rm c}$	$22.2_{\rm c}$	$47.3_{\rm c}$	19.9_{b}	31.7_{b}	$22.3_{\rm c}$	44.2 _a	18.0_{b}	47.9_{a}	92.8_{a}	118.7 _a	74.2_a

 $^{^{}a}$ Average (n = 3). Different letters within the same column indicate significant difference (p < 0.05).

3.7 Antioxidant Activity

Figure 3 presents the antioxidant activity of tomatoes storage at different condition. In tomatoes, the principal antioxidants compounds are carotenoids (lycopene), phenolic compounds, flavonoids, and vitamins like C and E (Beutner et al., 2001; Sahlin, Savage, & Lister, 2004; Toor, Lister, & Savage, 2005). In this study, the antioxidant activity was $126.9 \pm 0.2~\mu M$ TEAC/100 g of tomato pulp. Toor and Savage (2005) informed values of 63 and 94 μM TEAC/100 g of tomato pulp for Tradiro and Excell cultivar, respectively. The antioxidant activity presents in pulp of tomato was lower to those obtained in skin and seed. Moreover, the hydrophilic extract presents significantly higher antioxidant activity to the lipophilic extract (Toor & Savage, 2005). During the storage, the antioxidant activity did not change significantly (p > 0.05), regardless the storage temperature. Zapata et al. (2007) reported that antioxidant compounds change during the marketing of tomato; increasing the content of lycopene and phenols; but decreasing the content of ascorbic acid.

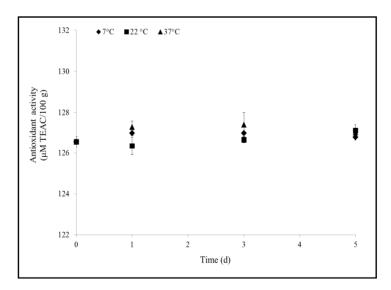


Figure 3. Antioxidant activity of tomato during the storage at different conditions

3.8 Relationship Between Quality Factors

Table 3 presents the Pearson correlation between a and b color parameters and some quality parameters of tomatoes. In tomatoes stored at 22 °C there was a high correlation (> 0.86) between the contents of carotenoids, vitamin C, and antioxidant activity with the increase of red color. Arias et al. (2000) informed that in tomatoes ripened on plant (room temperature) presented a high concentration of antioxidant compounds and high development of red color. The lycopene did not have a good fitting between the a and b color parameters, regardless the temperature of storage. At 37 °C, there is a good fitting between the antioxidant components (except lycopene) and firmness with the a color parameters. Moreover, in tomatoes stored at 7 °C it was observed that only vitamin C presents a high correlation with the a color parameter and the a/b index. Hart &

Scott (1995) informed that the antioxidant content of tomato depends of multiple factors, and some of them are the ripening stage and the storage temperature (Madhavi & Salunkhe, 1998). However, Javanmardi and Kubota (2006) pointed out that low temperature reduce the lycopene contents, but increases the antioxidant compounds.

Table 3. Pearson correlation ($\alpha = 0.05$) between color parameters and quality parameters of tomatoes during the storage

Compounds -		7 °C			22 °C			37 °C	
Compounds	а	a/b	b	а	a/b	b	а	a/b	b
Lycopene	-0.04	0.18	-0.38	0.67	0.6	-0.2	0.46	0.8	-0.81
Carotenoids	0.41	0.01	0.62	0.96	0.97	-0.86	0.8	0.64	0.33
Vitamin C	0.97	0.92	0.04	0.97	0.97	0.85	0.84	0.65	0.35
Antioxidants	0.65	0.03	0.69	0.86	0.9	0.97	0.87	0.89	-0.3
Firmness	-0.39	-0.79	0.58	-0.44	-0.54	0.89	0.93	0.8	0.25

4. Conclusions

During the postharvest of tomatoes occurs different change in the physicochemical characteristics and bioactive compounds that are affected by the storage temperature. The a color parameter significantly increased at higher temperature of storage (22 and 37 °C); moreover, the firmness increases as the storage temperature decrease. Color and firmness are critical factor for the acceptance of the consumers. At the end of the storage the higher values of lycopene (31.7 mg/kg FW) and total carotenoids (118.7 μ g/100 g FW) were observed in tomato stored at 22 °C. However, the vitamin C was higher in tomato stored at 7 and 37 °C, while the antioxidant activity did not change during the storage time. The bioactive compounds (carotenoids, vitamin C, and antioxidant activity) present a high correlation with the development of the red color at the temperature of 22 °C.

Acknowledgements

The authors thank the Program for the Improvement of Teaching, PROMEP (Project: 103.5/12/4310) for financial support of this research.

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