

Nutritional Analysis and Quality Evaluation in Muscle of Crucifix Crab *Charybdis feriatus* From Three Wild Populations

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Received: December 16, 2013 Accepted: January 8, 2014 Online Published: January 13, 2014

doi:10.5539/jfr.v3n2p27

URL: <http://dx.doi.org/10.5539/jfr.v3n2p27>

Abstract

Three wild populations of crucifix crab *Charybdis feriatus* were sampled and muscle was analyzed for proximate composition, amino acid and fatty acid composition, aimed to quantify and compare the nutritional quality from three different locations in China. Results showed that crude protein content in muscle of female crucifix crab *C. feriatus* from Zhoushan (ZS) and Xiapu (XP) (84.84%-88.35%) were significantly higher than that of crucifix crab *C. feriatus* from Qionghai (QH) (74.33%), while there was no significant difference in terms of crude fat content (3.82%-4.07%). The highest content of ash was found in muscle of crucifix crab *C. feriatus* from QH (5.36%). The muscle of crucifix crab *C. feriatus* from ZS and XP had significantly higher contents of total amino acids, essential amino acids, non-essential amino acids and delicious amino acids than those of QH group ($P < 0.05$). The total saturated and unsaturated fatty acid content in the muscle of crucifix crab *C. feriatus* from three wild populations showed significant difference ($P < 0.05$). The ZS group had highest content of poly-unsaturated fatty acids (33.64%) and total content of EPA and DHA (22.85%) as well, followed by XP and QH group ($P < 0.05$). Overall, the differences in chemical composition in muscle of *C. feriatus* from different locations could be attributed to environmental variables and maturation stage.

Keywords: *Charybdis feriatus*, wild population, proximate composition, amino acid, fatty acid

1. Introduction

Crustaceans are beneficial and highly appreciated due to their favorable taste and nutritional quality. In the last decades, the chemical composition and nutritional value of crustaceans have been investigated intensively (Skonberg & Perkins, 2002; Çelik et al., 2004; Naczek et al., 2004; Küçükgülmez et al., 2006; Chen et al., 2007; Barrento et al., 2009a; Barrento et al., 2009b; Barrento et al., 2010; Marques et al., 2010; Tsape et al., 2010; Wu et al., 2010; Maulvault et al., 2012; Risso & Carelli, 2012). The taste and nutritional quality of crab meat is to a large extent related to the presence of amino acids and fatty acids (Chen et al., 2007). And a balanced amino acid and fatty acid composition is essential for promoting good health as well (Wu et al., 2010).

The crucifix crab, *Charybdis feriatus*, is one of the most consumed crustaceans in countries of East and Southeast Asia (Josileen, 2011). The meat of this crab is particularly tasty and desirable, and is available throughout the whole year. This crab has a striking feature of distinctive red, black and white exoskeleton with a cross on the median surface of the carapace (Padayatti, 1990). The juvenile crucifix crab selects its habitat in the sandy shore, and the adult inhabits muddy offshore areas (Baylon & Suzuki, 2007). This crab is abundantly distributed in the eastern and southeastern coasts of mainland China and is mostly captured and consumed in coastal areas of Zhejiang, Fujian and Hainan provinces. Considering that the chemical composition of a particular species may vary according to habitat (Souchet & Laplante, 2007; Barrento et al., 2010), the aim of this study was to determine nutritional composition and quality of the muscle of crucifix crab *Charybdis feriatus* captured from these three geographical locations in China. In particular, proximate chemical composition, amino acid content and fatty acid composition among the populations were investigated and compared. To the best of our knowledge, this is the first study to report the chemical composition of crucifix crab, and the results would give additional nutritional information to consumers.

2. Materials and Methods

2.1 Sampling

A total of 45 female crucifix crabs of species *Charybdis feriatus* were captured from 3 different locations along the southeastern coastal waters of China in March, 2013, including Zhoushan (ZS), Zhejiang province (305.09 ± 23.66 g, n=15), Xiapu (XP), Fujian province (306.71 ± 37.24 g, n=15) and Qionghai (QH), Hainan province (307.99 ± 32.52 g, n=15). These locations are listed in Table 1. Wild adult specimens collected from these locations were transported to laboratory immediately, weighed and prepared for sampling the muscle. Each set of 15 crabs was divided into three groups and muscle (including claws, walking legs and body) from each group (5 specimens) were pooled together. Each sample was subsequently homogenized with a grinder, packed in small plastic bags and stored at -40 °C until analysis.

Table 1. Characteristics of three locations of *Charybdis feriatus*

Location	Code	Capacity	Latitude (N)	Longitude (E)
Zhoushan	ZS	15	29°16"	122°08"
Xiapu	XP	15	26°46"	120°42"
Qionghai	QH	15	19°36"	110°13"

2.2 Proximate Composition Analysis

The crab muscle samples were analyzed for moisture, protein, lipid and ash by using AOAC 2005 (Association of Official Analytical Chemists) standard methods 950.46, 928.08, 991.36 and 920.153, respectively. In brief, moisture was determined by drying the samples at 105 °C to constant weight. Crude protein was measured by determining nitrogen content ($\times 6.25$) using automated Kjeldahl analysis (Foss Tecator Kjeltac Auto 2200 analyzer, Warrington, UK). Lipid was determined by petroleum ether extraction in a soxlet apparatus (Foss Tecator 148 Soxtec system 2043 Auto Extraction apparatus, Warrington, UK). Ash was determined by combustion to a constant weight in a muffle furnace at 550 °C (Lindberg/Blue M, Thermo Fisher Scientific Inc., Waltham, USA).

2.3 Amino Acid Composition Analysis

For amino acid analysis, the samples were hydrolyzed with $6 \text{ mol}\cdot\text{L}^{-1}$ HCL and determined by amino acids analyzer (Biochrom Ltd., Cambridge, UK) with a C18 column ($25 \text{ cm} \times 4.6 \text{ mm}$), according to the method of Chinese Standard GB/T 5009.124-2003. For measuring tryptophan content, each sample was hydrolyzed with $5 \text{ mol}\cdot\text{L}^{-1}$ NaOH before analysis. The amino acid content was calculated by comparison with retention time and the peak areas of standard amino acids.

2.4 Fatty Acid Composition Analysis

For fatty acid analysis, total lipids of samples were extracted and fatty acid methyl esters were prepared in accordance with Chinese Standard GB/T 22223-2008. Briefly, total lipids were extracted with chloroform-methanol solution (2:1, v/v). Fatty acid methyl esters (FAMES) were prepared using a 15% (w/v) BF₃-methanol reagent. FAMES were measured by using HP-6890 GC series gas chromatograph and a column ($60 \text{ m} \times 0.25 \text{ mm} \times 0.25 \mu\text{m}$). Nitrogen was the gas carrier and the column temperature was set to increase from 130 °C to 230 °C, at a rate of 4 °C/min. The fatty acids composition was determined by comparing the areas of the various fatty acids analyzed to the areas of a fixed concentration of the individual and mixed FAME standard.

2.5 Calculations and Statistical Analysis

Amino acid score (AAS) was calculated according to FAO/WHO reference amino acid standard for adults, and the chemical score (CS) was calculated with respect to the amino acid composition of egg protein. The AAS and CS were calculated using the following formula (Biel et al., 2009; Gong et al., 2013):

Amino acid score= Sample amino acid (mg/g N) / FAO/WHO reference amino acid (mg/g N)

Chemical score= Sample amino acid (mg/g N) / amino acid of egg protein (mg/g N).

The propensity of crab muscle to the incidence of coronary heart disease, atherogenic index (AI) and thrombogenic index (TI) were calculated according to the following formula (Barrento et al., 2010):

AI, atherogenic index= $(\text{C12:0}+4\times\text{C14:0}+\text{C16:0}) / [(\text{MUFA} + \text{PUFA} (\text{n-6}) \text{ and } (\text{n-3}))]$

TI, thrombogenic index= $(C14:0+C16:0+C18:0) / [(0.5 \times PUFA + 0.5 \times PUFA (n-6) + 3 \times PUFA (n-3) + (n-3) / (n-6)]$.

Analyses were carried out in triplicate and the results were presented as mean and standard deviation (means \pm SD). The data were analyzed by one-way analysis of variance (ANOVA) through SAS 9.0 statistical software (SAS Institute Inc., Cary, NC, USA). Significant ($P < 0.05$) differences among the samples were ranked by Duncan's Multiple Range Test.

3. Results and Discussion

3.1 Proximate Chemical Composition

Proximate chemical composition of crab muscle samples from three wild populations is shown in Table 2. The crab muscle had an average moisture content of 78.93%, in which QH group showed highest contribution (80.26%), followed by ZS group (78.70%) and XP group (77.83%). Crude protein content of muscle averaged between 74.33%-88.35% and together with lower content of crude lipid (3.82%-4.07%) in dry weight basis. These results were similar to those of other reported crab species, which were determined to be 80.60%-88.97%, 2.00%-4.25% of crude protein and crude lipid in dry weight, respectively (Skonberg & Perkins, 2002; Küçükgülmez et al., 2006; Chen et al., 2007; Maulvault et al., 2012). Differences in proximate composition were observed among the sampled populations from three fishing locations. QH population had the lowest protein content (74.33%), compared with those from the other two populations ($P < 0.05$). No significant differences in lipid content were observed among three populations ($P < 0.05$). The highest ash content was found in QH population (5.36%), followed by XP (4.41%) and ZS population (4.23%). The similar proximate composition found in ZS and XP group might be much closer geographic habitats with similar living environments.

Table 2. Proximate chemical composition of three wild populations of *Charybdis feriatus* (% , dry weight) n=3, means \pm SD

Nutritional components	ZS	XP	QH
Moisture	78.70 \pm 0.97	77.83 \pm 0.60	80.26 \pm 1.21
In dry weight, %			
Crude protein	84.83 \pm 0.65 ^a	88.35 \pm 1.23 ^a	74.33 \pm 1.55 ^b
Crude fat	3.94 \pm 0.01 ^a	3.82 \pm 0.02 ^a	4.07 \pm 0.02 ^a
Ash	4.23 \pm 0.02 ^c	4.41 \pm 0.02 ^b	5.36 \pm 0.30 ^a

ZS, crabs from Zhoushan; XP, crabs from Xiapu; QH, crabs from Qionghai

Within the same row, values with different superscripts are significantly different ($P < 0.05$).

3.2 Amino Acid Composition

The detailed amino acid composition of muscle sampled from three wild populations is given in Table 3. The most abundant amino acid in the samples was glutamic acid (11.05%-12.42%), followed by aspartic acid (6.86%-7.61%), arginine (6.58%-7.46%), lysine (5.71%-6.41%), glycine (5.51%-5.86%), leucine (5.27%-5.79%), and alanine (3.45%-4.41%) in decreasing amount. The lowest content of amino acid was tryptophan among the three wild populations. High levels of taurine were found in muscle samples which are between 0.80% and 1.23%. Taurine is not incorporated in protein synthesis, but it plays a vital role in physiological functions during development and aging (Ripps & Shen, 2012). Since humans have limitation in biosynthesizing of taurine, the dietary supplementation seems to be necessary. No significant difference in any single amino acid content was found among the samples in the present study. The total amino acid (TAA) content ranged from 70.22% to 78.55%. The essential amino acids (EAA) content of all samples (23.56%-26.57%) was found to be lower than non-essential amino acids (NEAA) content (46.67%-51.98%), and the ratio of EAA/NEAA ranged from 57.37% to 58.16%. The amino acid pattern of higher amounts of non-essential amino acids was found in other reported crustaceans as well (Barrento et al., 2010; Wu et al., 2010). The EAA and delicious amino acids (DAA) accounted for 33.54%-33.82%, and 38.12%-38.42% of total amino acid content (TAA) of the muscle samples, respectively. Variations among different locations were also observed in amino acid content. Among three wild populations, significant higher content of EAA, NEAA, DAA and TAA was found in ZS and XP group than QH group ($P < 0.05$) as found in protein content. In general, muscle is the main protein storage location in crabs.

Environmental factors (water temperature, salinity, dissolved oxygen, photoperiod, and diet type and availability) and maturation stage would have effect on protein metabolism in crab muscles, resulting in different amino acid contents of crabs at different locations (Antunes et al., 2010).

Table 3. Amino acid content of three wild populations of *Charybdis feriatus* (% dry weight) n=3, means \pm SD

Amino acids	ZS	XP	QH
Taurine	1.23 \pm 0.01	1.01 \pm 0.01	0.80 \pm 0.01
Aspartic acid	7.61 \pm 0.09	7.29 \pm 0.12	6.86 \pm 0.06
Threonine	3.31 \pm 0.05	3.27 \pm 0.03	3.09 \pm 0.03
Serine	3.07 \pm 0.04	3.00 \pm 0.05	2.88 \pm 0.01
Glutamic acid	12.42 \pm 0.14	12.40 \pm 0.17	11.05 \pm 0.10
Glycine	5.51 \pm 0.06	5.86 \pm 0.08	5.54 \pm 0.05
Alanine	4.41 \pm 0.03	4.15 \pm 0.12	3.45 \pm .74
Valine	3.31 \pm 0.01	3.21 \pm 0.05	2.98 \pm 0.03
Methionine	1.50 \pm 0.03	1.58 \pm 0.03	0.93 \pm 0.01
Isoleucine	3.09 \pm 0.04	3.06 \pm 0.03	2.78 \pm 0.03
Leucine	5.79 \pm 0.10	5.71 \pm 0.08	5.27 \pm 0.05
Tyrosine	3.05 \pm 0.02	2.91 \pm 0.04	2.64 \pm 0.02
Phenylalanine	3.15 \pm 0.03	3.06 \pm 0.06	2.80 \pm 0.03
Histidine	2.06 \pm 0.01	1.87 \pm 0.05	1.82 \pm 0.02
Lysine	6.41 \pm 0.07	6.23 \pm 0.07	5.71 \pm 0.05
Arginine	7.31 \pm 0.08	7.46 \pm 0.11	6.58 \pm 0.06
Proline	2.92 \pm 0.03	2.82 \pm 0.07	2.69 \pm 0.03
Cystine	3.02 \pm 0.07	2.79 \pm 0.07	2.64 \pm 0.02
Tryptophan	0.62 \pm 0.00	0.63 \pm 0.00	0.52 \pm 0.00
TAA	78.55 \pm 0.91 ^a	77.29 \pm 1.25 ^a	70.22 \pm 1.34 ^b
EAA	26.57 \pm 0.33 ^a	26.12 \pm 0.35 ^a	23.56 \pm 0.22 ^b
NEAA	51.98 \pm 0.58 ^a	51.17 \pm 0.90 ^a	46.67 \pm 1.13 ^b
DAA	29.94 \pm 0.31 ^a	29.70 \pm 0.49 ^a	26.90 \pm 0.96 ^b
EAA/TAA	33.82	33.79	33.54
EAA/NEAA	58.05	58.16	57.37
DAA/TAA	38.12	38.42	38.30

ZS, crabs from Zhoushan; XP, crabs from Xiapu; QH, crabs from Qionghai.

Within the same row, values with different superscripts are significantly different ($P < 0.05$).

As shown in Table 4, the amino acid score (AAS) in three sampled populations was with no difference, and the scores were similar or higher than reference FAO/WHO pattern except for valine and tryptophan (AAS < 0.8), while the chemical score (CS) of amino acid was lower than the reference egg protein data except for lysine (CS > 1.0). The high lysine content in crab muscle samples could be a supplementation of plant-protein based diets. Tryptophan was found to be the EAA with the lowest content, and it would be the limiting amino acid in muscle of crucifix crab *Charybdis feriatus*.

Table 4. AAS and CS among three wild populations of *Charybdis feriatus*

	Amino acids	FAO/WHO pattern	Egg protein	ZS	XP	QH
AAS	Ile	250		0.91	0.87	0.94
	Leu	440		0.97	0.92	1.01
	Lys	340		1.39	1.30	1.41
	Met+Cys	220		1.51	1.40	1.36
	Phe+Tyr	380		1.20	1.11	1.20
	Thr	250		0.98	0.93	1.04
	Val	310		0.79	0.74	0.81
	Trp	60		0.76	0.73	0.73
CS	Ile		331	0.69	0.65	0.71
	Leu		534	0.80	0.76	0.83
	Lys		441	1.07	1.01	1.09
	Met+Cys		386	0.86	0.80	0.78
	Phe+Tyr		565	0.81	0.75	0.81
	Thr		292	0.84	0.79	0.89
	Val		411	0.59	0.55	0.61
	Trp		106	0.43	0.42	0.41

AAS, amino acid score; CS, chemical score.

3.3 Fatty Acid Composition

The fatty acid composition of muscle sampled from three wild populations is given in Table 5. The saturated fatty acids (SFA) (37.73%-42.55% of total lipid) were found to be the main group of fatty acids in three samples followed by polyunsaturated fatty acids (PUFA) (25.87%-33.64% of total lipid) and monounsaturated fatty acids (MUFA) (28.49%-32.84% of total lipid). Variations among wild populations were also found in crab's FA profile. For instance, QH population had the highest SFA compared with that of the other two populations ($P < 0.05$). Content of PUFA in muscle samples were in descending order: ZS (33.64%) > XP (29.44%) > QH (25.87%). The most abundant saturated fatty acid in samples was palmitic acid (C16:0), similar to the data obtained from reports of other crab species (Barrento et al., 2010; Maulvault et al., 2012). For monounsaturated fatty acids, the oleic acid (C18:1n-9) was the most abundant fatty acid. Docosahexaenoic acid (DHA, 22:6n-3) and eicopentaenoic acid (EPA, 20:5n-3) were the main PUFA in muscle samples. Similarly high amount of DHA and EPA is also reported in other crab species in literatures (Barrento et al., 2010; Marques et al., 2010; Maulvault et al., 2012). These two fatty acids are related to lower risk of cardiovascular disease and inflammation (Breslow, 2006; Calder, 2006). The total amount of EPA and DHA were considerably higher in ZS group than that of the other two wild groups ($P < 0.05$). The n-3 and n-6 PUFA of the samples ranged 24.72%-32.04% and 1.15%-1.71% of the total lipid, respectively. And the higher percentage of total n-3 PUFA ($P < 0.05$) in ZS group were mainly due to higher amount of DHA and EPA. The ratio of n-3 to n-6 PUFA in the present study ranged from 16.27 to 21.46, which is higher than those reported marine crabs (2.55-5.24) (Barrento et al., 2010; Wu et al., 2010; Maulvault et al., 2012). The AI and TI in the samples was 0.38-0.43 and 0.28-0.35, respectively. The values in this crab muscle were lower when compared to those of land animals (0.50-1.00) (Rosa & Nunes, 2004), indicating lower incidence of coronary heart disease. The differences in crab's fatty acid profile is considered to be environmental variables (e.g. water temperature, salinity, diet type and availability) and different stage of maturation of crabs captured from different locations as well (Vinagre et al., 2007; Antunes et al., 2010).

Table 5. Fatty acid composition among three wild populations of *Charybdis feriatus* (% of total lipid) n=3, means \pm SD

Fatty acids	ZS	XP	QH
C15:0	0.88 \pm 0.06	0.67 \pm 0.33	1.29 \pm 0.14
C16:0	24.73 \pm 0.01	23.48 \pm 0.37	24.49 \pm 0.51
C17:0	0.45 \pm 0.07	0.63 \pm 0.22	2.15 \pm 0.71
C18:0	11.80 \pm 0.27	12.94 \pm 0.11	14.62 \pm 0.87
C16:1	6.09 \pm 0.07	7.75 \pm 0.01	3.08 \pm 0.76
C18:1n9	22.11 \pm 0.18	24.72 \pm 0.06	28.30 \pm 0.67
C20:1n9	0.29 \pm 0.08	0.37 \pm 0.14	-
C18:2n6	1.24 \pm 0.14	1.39 \pm 0.26	1.15 \pm 0.54
C20:2n6	0.36 \pm 0.51	0.32 \pm 0.05	-
C20:3n3	9.19 \pm 0.06	7.52 \pm 0.18	9.22 \pm 0.28
C20:5n3(EPA)	12.69 \pm 0.32	11.96 \pm 0.33	9.59 \pm 0.38
C22:5n3	-	0.85 \pm 0.17	-
C22:6n3(DHA)	10.16 \pm 0.34	7.40 \pm 0.03	5.92 \pm 0.07
SFA	37.87 \pm 0.25 ^b	37.73 \pm 0.15 ^b	42.55 \pm 1.22 ^a
MUFA	28.49 \pm 0.02 ^b	32.84 \pm 0.08 ^a	31.58 \pm 0.09 ^a
PUFA	33.64 \pm 0.23 ^a	29.44 \pm 0.23 ^b	25.87 \pm 1.12 ^c
n-6	1.60 \pm 0.38	1.71 \pm 0.19	1.15 \pm 0.54
n-3	32.04 \pm 0.60 ^a	27.73 \pm 0.05 ^b	24.72 \pm 0.58 ^c
EPA+DHA	22.85 \pm 0.66 ^a	19.36 \pm 0.30 ^b	15.50 \pm 0.30 ^c
n-3/n-6	20.01	16.27	21.46
AI	0.40	0.38	0.43
TI	0.28	0.31	0.35

ZS, crabs from Zhoushan; XP, crabs from Xiapu; QH, crabs from Qionghai; AI, atherogenic index; TI, thrombogenic index.

Within the same row, values with different superscripts are significantly different ($P < 0.05$).

4. Conclusions

The present study was carried out to quantify the chemical composition and nutritional value of crucifix crab *Charybdis feriatus* captured from three geographical locations in China. This is, to the best of our knowledge, the first study documenting the amino acid content, fatty acid composition of crucifix crab *C. feriatus*. Crabs are exposed to environmental variables in different habitats, such as water temperature, salinity, dissolved oxygen, photoperiod, and diet type and availability or maturation stage-all of which may produce differences in chemical composition.

Acknowledgements

This work was supported by a special research fund (Project NO. 2012M04) for the national non-profit institutes (East China Sea Fisheries Research Institute).

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