Wine Oligosaccharides: Underutilized or Irrelevant? A Study into the Effects of Oligosaccharides on Wine Taste and Mouthfeel

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Received: October 2, 2021 Accepted: October 20, 2021 Online Published: October 25, 2021
doi:10.5539/jfr.v10n5p60 URL: https://doi.org/10.5539/jfr.v10n5p60

Abstract

The taste and mouthfeel of a wine are two of the most important aspects of wine tasting. However, while much is known about phenolic compounds and other macromolecules direct effects on wine taste and mouthfeel, little is known about other wine compounds such as oligosaccharides. This experiment uses Fructo-oligosaccharide (FOS) and Galacto-oligosaccharide (GOS) at two different concentrations, 450 mg/L and 900 mg/L within a simple model wine matrix. A model matrix was used to control for any unknown interactions between oligosaccharides and the multitude of wine components. Oligosaccharides were added individually to the model wine matrix at each concentration to create four treatments. Triangle tests were performed on all treatments against the control base model wine and between the high and low concentrations of each oligosaccharide treatment. Following the triangle tests, each treatment and the control underwent descriptive analysis (DA) using line intensity scales for sweetness, bitterness, astringency, acidity, and viscosity. Triangle test results revealed a significant difference only between the FOS450 and FOS900 samples. The wine matrix was made more complicated by adding polyphenols and still, none of the four oligosaccharide treatment groups were found to be significantly different. DA found no significant differences for the five attributes but did show clear trends in increased sweetness and acidity, decreased bitterness, as well as changes to astringency and viscosity. This suggests there may be more complex interactions happening within the mouth. However, given the lack of significant results in the simple wine model and the more complex wine model wine, any complex interactions between oligosaccharides and other wine compounds are likely to be minimal.

Keywords: galacto-oligosaccharides, Fructo-oligosaccharides, triangle tests, polyphenols, descriptive analysis

1. Introduction

Wine is an inherently complex system consisting of hundreds of different components that potentially impact organoleptic perception. Oligosaccharides are just one class of compounds that are present in wine and are generally defined as carbohydrates with low degrees of polymerization, specifically a mean degree of polymerization (mDP) of 2-20 units (BeMiller, 2019). Oligosaccharides consist of monomeric saccharides covalently linked via glycosidic bonds. Finished wines of differing varietals have different oligosaccharide compositions (Apolinar-Valiente et al., 2015). While the composition and concentration of oligosaccharides has been found to vary (Bordiga et al., 2012; Boulet et al., 2016), the potential impact on wine sensory perception has yet to be investigated thoroughly.

Current understanding shows that oligosaccharide concentration can vary between 50-550 mg/L and is dependent on several factors (Bordiga et al., 2012; Boulet et al., 2016). They originate from both grape and yeast cell walls (Chong et al., 2019) and differences in composition and concentration are due to grape cultivar, grape ripeness, terroir, enzymatic activity, winemaking, and vineyard management practices (Apolinar-Valiente et al., 2013, 2014, 2015; Bordiga et al., 2012; Ducasse et al., 2010, 2011; Vicens et al., 2009; Zietsman et al., 2015). The sheer number of factors known to affect the composition and concentration of oligosaccharides presents challenges in understanding how to alter oligosaccharide content in wine and how oligosaccharide modification might impact wine quality.

Current understanding of oligosaccharides on wine sensory perceptions is quite limited to date. Research has shown an impact on astringency perception and its relation to the presence of galactose and mannose within the oligosaccharide fraction of wine (Quijada-Morín et al., 2014). Astringency is defined as an overall drying,
puckering or rough sensation within the mouth (Huang & Xu, 2021). While astringency is an important aspect within wine sensory, it is only one attribute within an enormous array of sensory perceptions. In addition to wine taste and aroma, mouthfeel is generally difficult to understand. It is the result of multiple tactile oral stimulations and is understood by consumers as “a holistic multisensory perception of flavour” (Laguna et al., 2017). While astringency is the most heavily researched characteristic, there are many other descriptors used for wine mouthfeel such as drying, puckering, gritty, and hot (Gawel et al., 2000; Pickering & Demiglio, 2008).

The sensory impact of oligosaccharides on wine is mostly unknown, but oligosaccharides of various types have been linked to other organoleptic responses in other food systems such as low-fat yogurt. Oligosaccharides in water demonstrated that the compounds can alter sweetness, although the impact is dependent on concentration, degree of polymerization, type of oligosaccharide, and chemical linkage (Lapis et al., 2014, 2016; Low et al., 2017; Pullicin et al., 2017, 2019; Ruiz-Aceituno et al., 2018). Additionally, some connections have been found for oligosaccharides effects on other mouthfeel characteristics but are typically dependent on the food system in which they are evaluated. For example, inulin when added to low-fat yogurt was described as “thick”, “airy” and “sticky”, depending on mDP and concentration (Kip et al., 2006).

Previous studies investigating oligosaccharide mouthfeel perception in wine analyzed wine samples for oligosaccharide make up and correlated the analyses with sensory results (Apolinar-Valiente et al., 2015; Bordiga et al., 2012). While this approach helps gain an initial understanding of the mechanisms at work, wine is simply too complex to gain a completely accurate understanding from this method alone. The present study was separated into three distinct tests. The first investigated if adding different oligosaccharide types and concentrations to a simple model wine solution resulted in perceivable taste and/or mouthfeel differences. The second test aimed to quantify any differences found in the initial triangle tests via descriptive analysis and to determine how the oligosaccharides influence perception. The third experiment explored potential taste and mouthfeel differences due to oligosaccharides when combined with compounds known to impact mouthfeel perception. Polyphenols influence mouthfeel, specifically perceived astringency (Ferrer-Gallego et al., 2014). This test used polyphenols in addition to oligosaccharides of different types and concentrations to investigate the impact on taste and mouthfeel perception.

Overall, these three experiments investigate oligosaccharide influence on taste and mouthfeel within a simple and more complex system, quantify those differences and relate them with specific attributes. Knowledge of compositional aspects that cause specific sensory attributes are key when trying to achieve or maintain specific qualities in wine, particularly when each year the starting material, grapes, may be different. The causes of specific mouthfeel attributes are largely unknown, and this work aims to provide more information in this area.

2. Method

2.1 Chemicals

Fructo-oligosaccharide (FOS) and Galacto-oligosaccharide (GOS) were used as mouthfeel stimuli. The FOS used is a commercially available dry powder containing at least 95% Fructo-oligosaccharides (NUTRAFLORA® P-95, soluble prebiotic fiber, Ingredion, Westchester, Illinois, USA). The FOS powder consisted of oligosaccharide chains of three different degrees of polymerization, with the remaining percentage consisting of fructose, glucose, and sucrose. The sample provided for the study contained fructo-oligosaccharides at the following concentrations: DP3 36.1%, DP4 50.5%, and DP5 10.6%, with average molecular weight of 490.90 g/mol. The GOS sample used is a commercially available dry powder containing at least 68% galacto-oligosaccharides (Manufacturer requests omission from publication). The sample provided had an exact concentration of GOS of 71.6%, with the remainder of the syrup being comprised of lactose at 24%, and glucose + galactose at 4%. The degree of polymerization of the oligosaccharide fraction of the GOS sample was DP2 31%, DP3 38%, DP4 18%, DP5 8%, and DP6 5% with an average molecular weight of 533.63 g/mol. Both samples underwent preliminary tasing at concentrations higher than experimental values to ensure none of the residual shorter chain sugar molecules created noticeable sweetness (data not shown).

2.2 Wine Matrix

The base model wine consisted of 12% (v/v) ethanol (Everclear, Luxco, St. Louis, MO, USA) and 4 g/L tartaric acid (Modernist Pantry, Eliot, MO, USA) in deionized water and was adjusted to pH 3.5 using 0.5 M sodium hydroxide. The base model wine was created in five-gallon glass carboys three days prior to testing and stored at 4°C until use.

2.3 Sensory Analysis

Approval for work was granted by Institutional Review Board at Oregon State University (IRB-2020-0610).
Inclusion criteria for panelists was individuals who consumed 1 glass of wine a week on average. Individuals suffering from taste deficits and other oral disorders, oral lesions, canker sores, or wine allergies were excluded from participation. Smokers, pregnant persons, and individuals with tongue, lip, or cheek piercings were also excluded.

90 eligible panelists comprised of 24 male, 65 female, and 1 non-binary person ages 21 to 60+ attended the initial triangle test panel. Panelists were told they would be evaluating wine and/or model wine solutions that contain different additions. From the 90 participants of the triangle tests, 22 panelists were invited back for the descriptive analysis. Of those 22 participants, 4 were male, 18 were female, and ages ranged from 21 to 60+.

The second triangle test consisted of 75 panelists comprised of 14 males, 61 females, ages 21 to 60+. All panels were conducted in the Arbuthnot Dairy Center on the OSU Campus (Corvallis, OR). Compusense® Cloud Software (Version 21.0.7773.192939) was used to administer each test. The testing room was held at 20-22°C and two Winix Plasmawave air purifiers (Winix, Vernon Hills, IL, USA) were used to ensure air quality. Custom built white plastic tabletop tri-folds (61cm x 71cm center, 61cm x 65cm sides) were used to create individual booths for panelists. The room was a mix of artificial and natural light. All flights were served in black INAO wine glasses (Lehmann glass, Kiyasa Group, New York, NY, USA). Panelists were instructed to wear a foam-padded nose clip (Biotronics, Davie, FL, USA) during each triangle test. Panelists were provided with spit cups for expectorating samples and a 1 g/L pectin (Modernist Pantry, Eliot, MO, USA) rinse to use between samples. A high-speed immersion blender (Mueller Austria Ultra-Stock, City of Industry, CA) was used to suspend the pectin in deionized water.

2.4 Stimuli

The stimuli treatments consisted of two different oligosaccharide groups, FOS and GOS, at two different concentration levels, 450 mg/L and 900 mg/L. FOS and GOS were selected as they are both commercially available as food grade products. GOS is naturally occurring within wine (Osborne et al., 2019), while FOS can be produced by yeast, dependent upon yeast strain and fermentative conditions (Deffert et al., 2017). 450 mg/L was selected because previous studies have shown this is the average oligosaccharide concentration in Pinot noir (Osborne et al., 2019). 900 mg/L was selected to see if greatly increasing the concentration of oligosaccharides influenced taste and mouthfeel. To prepare samples for sensory analysis, the model wine matrix was taken from the cooler and dispensed into 750 mL bottles (Tricor, St. Louis, MO, USA) and closed with screwcaps (Amcor, Zürich, Switzerland) no more than 24 hours before the panel. For the control treatment no other compounds were added to the base model wine matrix. Both FOS and GOS were added to 500mL of model wine via gentle stirring (no solubility difficulties due to low pH noticed), resulting in stock solutions with the concentration of 16.875 g/L for each oligosaccharide. The stock solutions of each oligosaccharide were then added to bottles to create the four treatments: FOS450, FOS900, GOS450, and GOS900. The solutions were added to the base model wines in bottle no more than 24 hours prior to the day of the panel.

2.5 Initial Triangle Test

Triangle tests were conducted in accordance with the ISO 4120:2004(E) method (ISO 4120, 2004). Each panelist received six different triangle tests, four tests compared each of the treatment groups against the control, one compared FOS450 to FOS900, and another compared GOS450 to GOS900. Flights were presented in a randomized order and treatments presented within the flights were presented in a balanced order. There was a 60-second break between each triangle test, during which panelists were instructed to rinse their palate with the pectin rinse. Panelist wore nose clips while tasting and were only allowed to remove the clips during breaks between tests.

2.6 Descriptive Analysis

Descriptive analysis was performed using 100 mm line intensity scales for five different taste/mouthfeel characteristics. Training for descriptive analysis took place over two, one-hour training sessions. Panelists were trained on sweetness, bitterness, viscosity, astringency, and acidity standards. Initial concentrations of training standards were selected to encompass a wide range of responses for each attribute because of the limited knowledge about the effects of oligosaccharides and the desire to encompass all possible responses. Each training standard underwent preliminary testing via Check-All-That-Apply (CATA) to ensure consumers associated the standard with the given attribute (data not shown). Definitions of each attribute were provided during training and during the descriptive analysis (Table 1). For the first training session, panelists were presented with a taste or mouthfeel standard, the definition of the taste or mouthfeel attribute, and the corresponding location that the standard would be placed on the line scale. In the second training session, panelists were presented with training standards blind and were asked to score the samples on the line scales for
each attribute to ensure panelists understood the scaling. Panelists wore nose clips while evaluating samples for both training sessions. Between each sample set, panelists had a 60-second break during which they were instructed to rinse their palate with the pectin rinse.

Descriptive analysis took place over two, 1-hour sessions. Panelists evaluated nine model wine samples at each session. The wines were presented in a random order and all treatments were presented at least once per session. All samples were evaluated in triplicate by each panelist over the 2-day panel. Characteristics evaluated were sweetness, bitterness, viscosity, astringency, and acidity. Panelists were instructed to put on nose clips, taste each sample and rate the sample on the five attributes using their corresponding line scales. Line scales ranges from “not ___” to “very ___” for each given attribute. The attribute definition from training was provided above the corresponding line scale to help increase memory recall. Panelists were also provided a scale labeled “Other” to evaluate the sample on an attribute not previously presented. Panelists were instructed to write in the attribute they used for the “Other” scale. Panelists had 60-second breaks between each sample and were instructed to rinse their palate with the provided pectin rinse during the break.

2.7 Phenolic Triangle Tests

The third experiment followed the same procedure as the initial triangle tests, except those panelists received only four different triangle tests, each comparing the treatment groups to the control. The control contained 800 mg/L of polyphenols in the base model wine. This concentration of polyphenols was selected from the results following internal testing of 126 commercially available Pinot Noir wines, via MCP Assay (Sarneckis et al., 2006) (data not shown). The polyphenols were added by grinding grape seed extract (Masquelier’s Tru-Ope, Nature’s Way Brands, Green Bay, WI) into a fine powder, weighing out the necessary amount, and dissolving it in the base model wine solution. These polyphenols were chosen to represent wine astringency based on preliminary tastings (data not shown).

Table 1. Descriptive analysis training standards and definitions

<table>
<thead>
<tr>
<th>Attribute</th>
<th>Compound</th>
<th>Concentration</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sweetness</td>
<td>Sucrose</td>
<td>Low: 25 g/L</td>
<td>1 – Being one of the five basic taste sensations that is usually pleasing to the taste and typically induced by sugars (Merriam-Webster, n.d.)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>High: 75 g/L</td>
<td></td>
</tr>
<tr>
<td>Bitterness</td>
<td>Caffeine</td>
<td>Low: 0.5 g/L</td>
<td>Intensity of bitter taste perceived in the mouth (Sparrow et al., 2016)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>High: 1 g/L</td>
<td></td>
</tr>
<tr>
<td>Astringency</td>
<td>Aluminum Sulfate</td>
<td>Low: 0.4 g/L</td>
<td>Intensity of the drying and mouth puckering sensation in the mouth (Sparrow et al., 2016)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>High: 1 g/L</td>
<td></td>
</tr>
<tr>
<td>Viscosity</td>
<td>Carboxymethyl Cellulose</td>
<td>Low: 2 g/L</td>
<td>Perception of body, weight, or thickness of the wine in the mouth (Sparrow et al., 2016)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>High: 4 g/L</td>
<td></td>
</tr>
<tr>
<td>Acidity</td>
<td>Tartaric Acid</td>
<td>Low: 0.25 g/L</td>
<td>Intensity of the acid taste perceived in the mouth (Williamson, 2013)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>High: 1 g/L</td>
<td></td>
</tr>
</tbody>
</table>

2.8 Statistical Analysis

Statistical analysis was performed on triangle tests by bimodal distribution using ISO4120:2004(E) method Table A.1, then confirmed via Z-test on proportion. Z-scores and p-values are presented in Table 2 and Table 3. Analysis of Variance (ANOVA) was performed on descriptive analysis results using XLSTAT (XLStat 2020.3.1 Sensory Package, Addinsoft, Paris, France) and are displayed in Figure 1.

3. Results

3.1 Triangle Tests

Triangle test results shown in Table 2 and Table 3 indicate that none of the four treatments were significantly different from the control sample with or without phenolics present (α=0.05). It is worth noting that the proportion of correct responses increased with increasing oligosaccharide concentration when phenolics were present. Additionally, no significant difference was noticed between the GOS450 and the GOS900 samples. However, the results did show a significant difference (α=0.05) between the FOS450 and FOS900 samples in the initial triangle test. Additionally, the GOS900 sample was incredibly close to significance when added with phenolics, p-value=0.056.
Table 2. Initial Triangle Test Results

<table>
<thead>
<tr>
<th>Comparison</th>
<th>Number of Participants</th>
<th>Number of Correct Responses</th>
<th>Z-Score</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control v. FOS 450</td>
<td>90</td>
<td>36</td>
<td>1.23</td>
<td>0.109</td>
</tr>
<tr>
<td>Control v. FOS 900</td>
<td>90</td>
<td>29</td>
<td>-0.34</td>
<td>0.633</td>
</tr>
<tr>
<td>FOS 450 v. FOS 900</td>
<td>90</td>
<td>39</td>
<td>1.90</td>
<td>0.029*</td>
</tr>
<tr>
<td>Control v. GOS 450</td>
<td>90</td>
<td>28</td>
<td>-0.56</td>
<td>0.712</td>
</tr>
<tr>
<td>Control v. GOS 900</td>
<td>90</td>
<td>27</td>
<td>-0.78</td>
<td>0.782</td>
</tr>
<tr>
<td>GOS 450 v. GOS 900</td>
<td>90</td>
<td>35</td>
<td>1.01</td>
<td>0.156</td>
</tr>
</tbody>
</table>

*Significance at α = 0.05

Table 3. Secondary Triangle Test Results

<table>
<thead>
<tr>
<th>Comparison</th>
<th>Number of Participants</th>
<th>Number of Correct Responses</th>
<th>Z-Score</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control + Phenols v. FOS 450</td>
<td>75</td>
<td>19</td>
<td>-1.59</td>
<td>0.944</td>
</tr>
<tr>
<td>Control + Phenols v. FOS 900</td>
<td>75</td>
<td>25</td>
<td>-0.12</td>
<td>0.548</td>
</tr>
<tr>
<td>Control + Phenols v. GOS 450</td>
<td>75</td>
<td>22</td>
<td>-0.86</td>
<td>0.805</td>
</tr>
<tr>
<td>Control + Phenols v. GOS 900</td>
<td>75</td>
<td>32</td>
<td>1.59</td>
<td>0.056</td>
</tr>
</tbody>
</table>

*Significance at α = 0.05

3.2 Descriptive Analysis

ANOVA and Tukey Cramer HSD pairwise comparisons revealed no significant differences in any of the treatment groups for any of the attributes evaluated. However, a few notable trends were noticed and can be seen in Figure 1. Interestingly, sweetness appeared to increase slightly in all the treatments when compared to the control. This is interesting as preliminary tastings (data not shown) did not indicate any difference in sweetness intensity. Bitterness was reduced for each of the GOS treatments (particularly GOS900) and reduced bitterness perception more than FOS. Perceived astringency increased with each treatment except for GOS900 which had an average lower astringency than the treatments or control. Viscosity increased with both FOS treatments, and the GOS900 treatment. Finally, acidity increased with all four treatments, with the FOS treatments increasing acidity perception more than the GOS treatments.

![Figure 1. Descriptive analysis results by attribute for model wine with different concentrations (450mg/L or 900mg/L) of FOS or GOS](image-url)
4. Discussion

4.1 Discussion

For both FOS and GOS, the tests between the higher and lower concentrations had a higher proportion of correct responses than any of the concentrations versus the control. Except for FOS450, and while not statistically significant, FOS450 sample versus the control was in fact close to significance at a lower alpha level of \( \alpha = 0.10 \). It is worth noting that the same is true for the GOS450 versus GOS900 at an alpha level of \( \alpha = 0.15 \). These results indicate that a change in perception is occurring, but it is not large enough to be obvious. The change in taste and mouthfeel perception may be due to interactions between oligosaccharides and other compounds in saliva. Previous studies have demonstrated oral enzymatic activity, such as \( \alpha \)-amylase activity, can incite oral digestion of starches into malto-oligosaccharides and alter taste perception. (Robyt, 2009; Roberts & Whelan, 1960; Whelan & Roberts, 1952). Such enzymatic activity could increase the presence of carbohydrate monomers within the mouth due to the breakdown of oligosaccharides. This phenomenon might also explain the trend of increased sweetness perception noticed during the descriptive analysis.

While descriptive analysis did not yield statistically significant results some trends can be seen. As mentioned previously there appears to be a slight increase in sweetness perception for all treatments containing FOS or GOS. There is evidence to suggest that more complex carbohydrates are perceived as sweet (Lapis et al., 2014, 2016; Low et al., 2017; Pullicin et al., 2017, 2019; Ruiz-Aceituno et al., 2018). This sweetness perception varies depending on mDP and compound structure (Low et al., 2017; Pullicin et al., 2017). Additionally, sweetness of complex carbohydrates has been shown to be independent of enzymatic activity or hT1R2/hT1R3 receptor (Lapis et al., 2016; Yoon & Robyt, 2003). While these studies used different oligosaccharide types in a simpler system (water), they clearly show that oligosaccharides can elicit a sweet response. The trend of increased sweetness in all FOS and GOS samples within the descriptive analysis in our experiment is consistent with these findings.

The trend noted in viscosity perception could be explained by oligosaccharide size. Chong et al., (2019) found that soluble cell wall carbohydrates may play a role in mouthfeel perception, specifically viscosity, of Cabernet Sauvignon. Further, Gawel et al., (2016) noted polysaccharides had a small effect on mouthfeel specifically medium molecular mass polysaccharides, on wine viscosity at a higher pH. mDP and inulin concentration can impact rheological measurements in low-fat yogurt, leading to perceived changes in “thick”, “airy”, and “sticky” attributes. These terms are often associated with increased viscosity (Kip et al., 2006). Our findings provide further support, as the presence of oligosaccharides increased viscosity perception compared to the control in all treatments except GOS450.

The decreased bitterness in all oligosaccharide treatments is likely due to the increased sweetness perception, given sweet tastes have been found to mask bitterness in certain food systems (Hutchings et al., 2016; Mastaneh et al., 2013). Although this has not been explored specifically within a wine system, it is likely that similar perception changes would occur in wine. The lack of significant findings from the descriptive analysis tests might be due to differences in delivery mechanism. Previous oligosaccharide sweetness studies utilized water-based oligosaccharide solutions, while the present study employed oligosaccharides solubilized in a more complex wine matrix (Lapis et al., 2014, 2016; Low et al., 2017; Pullicin et al., 2017, 2019; Ruiz-Aceituno et al., 2018). This complex matrix could make discrimination test more difficult, particularly if the differences between treatments were more nuanced.

Interestingly, initial triangle test results revealed statistically significant differences between the FOS450 and FOS900 treatments only but not between either sample or the control. This lack of difference from the control rules out that the detection threshold of the oligosaccharide played a role in the difference noted between the two concentrations. Thus, another phenomenon must be at work within the system resulting in the statistically significant difference. Descriptive analysis results indicated FOS450 and FOS900 samples differ most in their acidity perception. This may be due to what many wine experts and consumers refer to as wine balance. While the definition of wine balance varies, it is generally referred to a holistic attribute with multiple components (Green et al., 2011; Parr et al., 2011). The increased carbohydrate concentration between FOS450 and FOS900 could alter the wine balance, explaining both the trends noticed in acidity perception and the decreased bitterness perception.

In the triangle tests where polyphenols were added, there were no significant differences noticed between any of the treatments and the control. While the proportion of correct responses increased for both FOS and GOS, these trends were not of statistical significance. This indicates that the concentrations tested are below a detection threshold for most consumers. Utilizing higher concentrations of oligosaccharides could result in more
perceivable differences. However, this study included oligosaccharide concentrations above levels previously reported in wine in order to tease out potential differences and higher concentrations would not be possible without significant changes to the extraction process which are outside the scope of typical winemaking.

Currently our understanding of oligosaccharide effects on organoleptic perception has mostly focused on astringency perceptions. Quijada-Morín et al. (2021) concluded that while astringency was positively related to mannose and galactose in the oligosaccharide faction, this was most likely due to the reduction in mannoproteins and polysaccharides rich in arabinose and galactose rather than the glycoside residues While not statistically significant, the positive correlation between astringency perception and oligosaccharide concentration is in agreement with Boulet et al., (2016). However, when polyphenols known to influence astringency (Ferrer-Gallego et al., 2014) were added to the base model wine solution, no difference was noticed. Trends were noticed during the descriptive analysis, and differences found via the initial triangle testing. The present study suggests that oligosaccharides, specifically fructo-oligosaccharides and galacto-oligosaccharides, do not have a significant influence on the taste and mouthfeel of wine and are an irrelevant factor in the winemaking process.

4.2 Limitations

Given the limited available literature, there was little previous knowledge in terms, established attributes, and training standards. Thus, a broad approach to the descriptive analysis training was utilized to encompass all possible responses. Due to this approach, the training standards included high concentration standards that panelists may have felt did not accurately represent the scale that the samples fell under.

Additionally, due to increased COVID-19 restrictions imposed by the state of Oregon (Executive Order 20-65) on November 17th, 2020, the training and testing schedule was shortened. This shortened training time resulted in less overall attribute training and could account for greater variability in the descriptive analysis.

4.3 Future Work

Future work should look at additional oligosaccharide types and interactions with other compounds, as different types could be perceived differently. Longer training using lower concentration standards should be conducted to help reduce variance in descriptive analysis.

4.4 Conclusion

Oligosaccharides appear to have very minor effects on taste and mouthfeel perception within a wine system. Statistically significant differences were found between the high and low concentrations of Fructo-oligosaccharides. None of the concentrations were found to be significantly different from the control samples in either of the triangle tests. Given that there was a significant difference found between the two Fructo-oligosaccharide samples in the first test, and not the controls in either test, it would appear there are more complex systems at work. This is illustrated by the trends noticed in descriptive analysis. These trends show an increase in sweetness and acidity perception, a decrease in bitterness, and differences in viscosity and astringency dependent upon concentration. While the trends are apparent in descriptive analysis, given the lack of differences noticed in both triangle tests, oligosaccharides do not play a large role in wine perception, and we do not recommend focusing on them during the winemaking process.

Acknowledgments

We would like to acknowledge Aubrey DuBois for her help in editing this manuscript.

References


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