

Soybean Industrial Seed Treatment: Effect on Physiological Quality During Storage

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Abstract

The aim of this study was to analyze soybean seed physiological quality after being subjected to various mixtures of pesticides via industrial seed treatment. The experiment was performed at the seed laboratory of the company BioGrow, located at São Paulo-SP, using soybean seeds cultivar NS 6700 IPRO which were subjected to 11 different treatments. Seed treatment was carried out using a treater Momesso, model L5-K, calibrated to apply a spray volume of 0.5 L 100 kg⁻¹ of seeds in which the volume of each treatment was adjusted using distilled water. After treatment, seeds were spread over plastic strays for drying for a period of 24 hours under environmental conditions. Once dry, seeds were packed in paper bags and stored for 0 (control), 45, 90, 135 and 180 days, under uncontrolled conditions of temperature and relative humidity, when seed physiological quality was evaluated using the following tests: germination, accelerated aging, seedling emergence, speed of emergence index and speed of emergence. Soybean industrial seed treatment before storage for up to 180 days is practicable using the mixtures of pesticides tested for storing seeds under environmental conditions. All treatments tested contribute to the maintenance of seed quality throughout storage.

Keywords: *Glycine max*, germination, vigor

1. Introduction

Soybean (*Glycine max* L. Merrill) has wide expression in the commodity market due to a major socioeconomic value which is determined by the numerous uses of products and byproducts on human and animal feed. World production, at the 2016/2017 crop season, was of 315.1 million tons, occupying 118.1 million of hectares (USDA, 2015). Brazil is the second greater soybean world producer, with a production of 94.8 million tons and a cultivated area of 31.4 million hectares (IBGE, 2016).

Various advances in soybean production technology have allowed the increase in area and crop yield in the recent years, among them is the quality of seeds used. According to Baudet and Peske (2007), and Silva (1998), uniform seed germination and vigorous growth of seedlings at the onset of crop development are essential to guarantee the maximum yield potential genetically contained. Likewise, Baudet and Peres (2004) observed that seeds with high germination, field emergence and growth potential can produce a uniform plant stand which results in high crop yields. However, seeds are sown at the field being exposed to numerous biotic (pests and diseases) and abiotic factors which impair physiological performance, affecting germination and reducing seedling emergence uniformity. Thus, to protect seeds and seedlings against a set of adversities, phytosanitary products as fungicides and insecticides are applied to the seeds (Ludwig et al., 2011; Pereira et al., 2011).

According to Avelar et al. (2011) soybean seed treatment is economically recommended, provided that the products or mixture of products used are adequate, using the recommended dosage and are uniformly distributed throughout the seed lot. Additionally, Lucca Filho (2006) determined that an efficient chemical treatment must select a product which is able to eradicate the pathogens present in the seeds, non-toxic to the crop, humans and the environment, non-corrosive, presents high stability, adherence and coverage, low cost and easy access, besides being compatible with other products.

Concomitantly with the increasing perception of the value of seeds and of the importance of protecting/increasing performance, the availability of products for seed treatment for different purposes has grown, such as protection (fungicides or insecticides) or nutrition (micronutrients), aiming to improve seed performance, both physiologically and economically (Avelar et al., 2011). However, besides increasing seed protection and assisting seedling initial development, the products and mixtures of products used in seed treatment shall not interfere negatively the physiological quality of seed lots, both immediately after treatment or during the storage period.

Accordingly, the present study aimed to analyze seed physiological quality after being subjected to various mixtures of products via seed treatment.

2. Method

The experiment was performed at the seed laboratory of BioGrow, located in São Paulo-SP, using soybean seeds cultivar NS 7600 IPR subjected to 11 different treatments described in Table 1.

Table 1. Treatment Identification (TR); commercial product used; active ingredient; concentration of the active ingredient contained in each product; dosage of the commercial product used and class of the product

TR	Name of the commercial product	Active ingredient (i.)	Concentration (g a. i. L ⁻¹)	Dosage of the commercial product (mL 100 kg ⁻¹ of seeds)	Class of the product	
TR-1	Control	Without treatment				
TR-2	Maxim XL	Fludioxonil	25	100	Fungicide	
		Metalaxyl-M	10			
TR-3	NUF 323F1 535 FS	Azoxystrobin	130	100	Fungicide	
		Fluazinam	105			
		Carbendazim	300			
TR-4	NUF 323F1 535 FS	Azoxystrobin	130	150	Fungicide	
		Fluazinam	105			
		Carbendazim	300			
TR-5	Maxim XL	Fludioxonil	25	100	Fungicide	
		Metalaxyl-M	10			
TR-6	Standak Top	Fipronil	250	200	Insecticide	
	Maxim XL	Fludioxonil	25	100	Fungicide	
TR-7	NUF 323F1 535 FS	Metalaxyl-M	10	200	Insecticide	
		Maestro FS	Fipronil			250
		Azoxystrobin	130			75
TR-8	NUF 323F1 535 FS	Fluazinam	105	200	Insecticide	
		Carbendazim	300			
		Maestro FS	Fipronil			250
TR-9	NUF 323F1 535 FS	Azoxystrobin	130	75	Fungicide	
		Fluazinam	105			
		Carbendazim	300			
TR-10	Biocoat GR Red	Film coater	-	100		
	NUF 323F1 535 FS	Azoxystrobin	130	75	Fungicide	
		Fluazinam	105			
TR-11	NUF 323F1 535 FS	Carbendazim	300	200	Insecticide	
		Maestro FS	Fipronil			250
		Biocoat GR Red	Film coater			-
TR-11	NUF 323F1 535 FS	Azoxystrobin	130	75	Fungicide	
		Fluazinam	105			
		Carbendazim	300			
TR-11	Maestro FS	Fipronil	250	200	Insecticide	
		Biocoat GR Red	Film coater			-

Seed treatment was performed using a treater Momesso, Model L5-K, calibrated to apply a spray volume of 0.5 L 100 kg of seeds⁻¹, in which the volume of each treatment was adjusted with distilled water. After treatment, seeds were distributed over plastic trays for drying for a period of 24 hours at room temperature. Once dried, seeds were placed into paper bags and stored for 0 (control), 45, 90, 135 and 180 days, under uncontrolled conditions of temperature and relative humidity, when seed physiological quality was evaluated using the following tests:

Germination: performed using germitest paper rolls wetted with distilled water, in a volume of 2.5 times de weight of the dry paper. Seeds were distributed in four paper rolls of 50 seeds each, totaling 200 seeds per repetition, which were transferred for a germination chamber and held at a constant temperature of 25 °C. Normal seedlings were counted at five and nine days after sowing accordingly to the Rules for Seed Testing (Brasil, 2009) and results were expressed in percentage.

Accelerated aging: carried out using the gerbox method; seeds were disposed in a single layer over a wire mesh which was held suspended above 40 mL of distilled water. The gerbox were covered and transferred to a Biological Oxygen Demand (B.O.D.) chamber at 41 °C for 48 hours (Marcos Filho, 2005). Then, seeds were removed from the gerbox and placed to germinate in the conditions described for the germination test. The evaluation was performed at the fifth day after sowing, counting the number of normal seedlings and expressing the results in percentage.

Seedling emergence: performed in trays filled with sand as substrate, sowing four repetitions of 50 seeds per treatment. Sowing depth was of approximately three centimeters and irrigation was carried out daily, at early morning and late afternoon; the percentage of emerged seedlings was counted at 21 days after sowing.

Speed of emergence index (SEI): carried out alongside the seedling emergence test. The index was calculated for each repetition accordingly to the procedure proposed by Maguire (1962), using the following equation: $SEI = (G1/N1) + (G2/N2) + \dots + (Gn/Nn)$, where G is the number of normal seedlings emerged each day and N is the number of days after sowing in which the counting was performed.

Speed of emergence (SE): evaluated herewith the seedling emergence and calculated using the following equation $SE = [(G1N1) + (G2N2) + \dots + (GnNn)] / (G1 + G2 + \dots + Gn)$. Where, G is the number of normal seedlings emerged at each day and N is the number of days after sowing in which the counting was carried out.

The experiment was performed as a 5 × 11 (storage periods × seed treatment) factorial under completely randomized design using four repetitions. Data processing was performed using R Software (R Core Team, 2014). Data were subjected to analysis of variance using the F-Test ($p < 0.05$) and, when significant differences were observed, a polynomial regression was calculated ($p < 0.05$) for the factor storage period and means were compared using the least significative difference ($p < 0.05$) for the factor seed treatment.

3. Results

According to the results of the analysis of variance, the storage period and the treatments applied presented a simple effect on the percentage of germination, however, there was no interaction between factors. Figure 1A presents the percentage of germinated seeds for each treatment applied where the control presented the greater percentage of germination (92.5%) which did not differ statistically from treatments TR-2, TR-3, TR-6, TR-7 and TR-10. Likewise, treatments TR-4, TR-9 and TR-11 presented a statistically lower percentage of germination compared to the other treatments. Regarding the percentage of germination throughout storage (Figure 1B), the variable was adjusted to a negative quadratic model where the maximum germination of 93% was observed immediately after seed treatment, which correspond to time zero of storage. Then, seed germination decreased accordingly with the increase of the storage time.

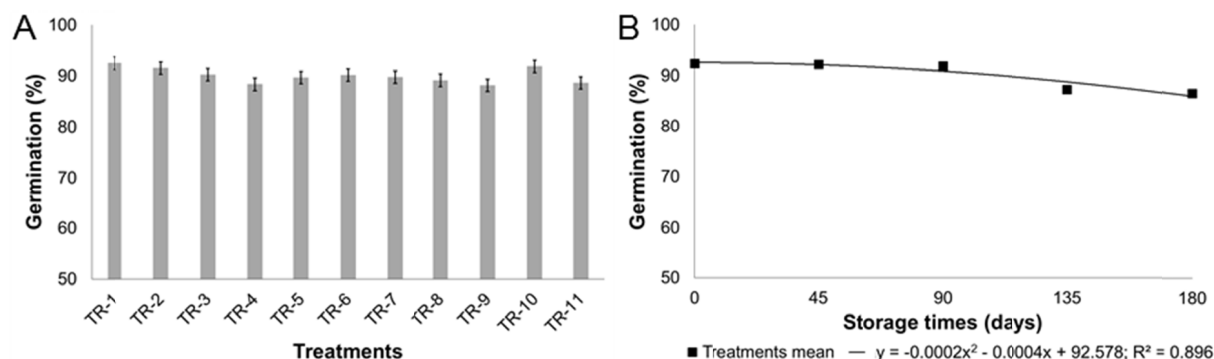


Figure 1. Germination percentage of soybean seeds cv. NS 6700 IPRO, in response to 11 seed chemical treatments (A) and to storage under uncontrolled conditions of temperature and relative humidity (B)

Note. TR-1: Control (without treatment); TR-2: Maxim XL (100 mL 100 kg⁻¹ of seeds); TR-3: NUF 323F1 535 (100 mL 100 kg⁻¹ of seeds); TR-4: NUF 323F1 535 FS (150 mL 100 kg⁻¹ of seeds); TR-5: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Standak Top (200 mL 100 kg⁻¹ of seeds); TR-6: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-7: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-8: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds); TR-9: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-10: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-11: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds). CV = 4.31%. In Figure 1A, bars correspond to the LSD value (Least significant difference) between different seed treatments ($p < 0.05$), therefore bars which overlap are statistically equal.

The analysis of variance demonstrated that the variable accelerated aging was affected by the interaction between the factors studied. Figure 2A shows that right after seed treatment, TR-8 was statistically superior and TR-9 was statistically inferior while the remaining treatments did not differ between each other. After 45 days of storage, TR-6 was statistically inferior to the other treatments. In the other periods of storage (90, 135 and 180 days), treatments TR-2, TR-9 and TR-10 were statistically superior to the remaining treatments, on the other hand, TR-5 presented the lower percentage of germinated seeds.

Treated seeds, when evaluated by the accelerated aging test, presented a linear decrease herewith the increase of storage time for all treatments studied, with exception of TR-9 which presented a quadratic behavior (Figures 2B and 2C). The rate of vigor reduction varied from 0.02 to 0.1% day⁻¹ where treatments TR-4, TR-5, TR-6, TR-7, TR-8 and TR-11 presented the greater rates of vigor reduction (Figures 2B and 2C). In contrast, TR-9, due to the quadratic behavior, showed a maximum percentage of germinated seeds of 89% at the 81 days of storage (Figure 2C).

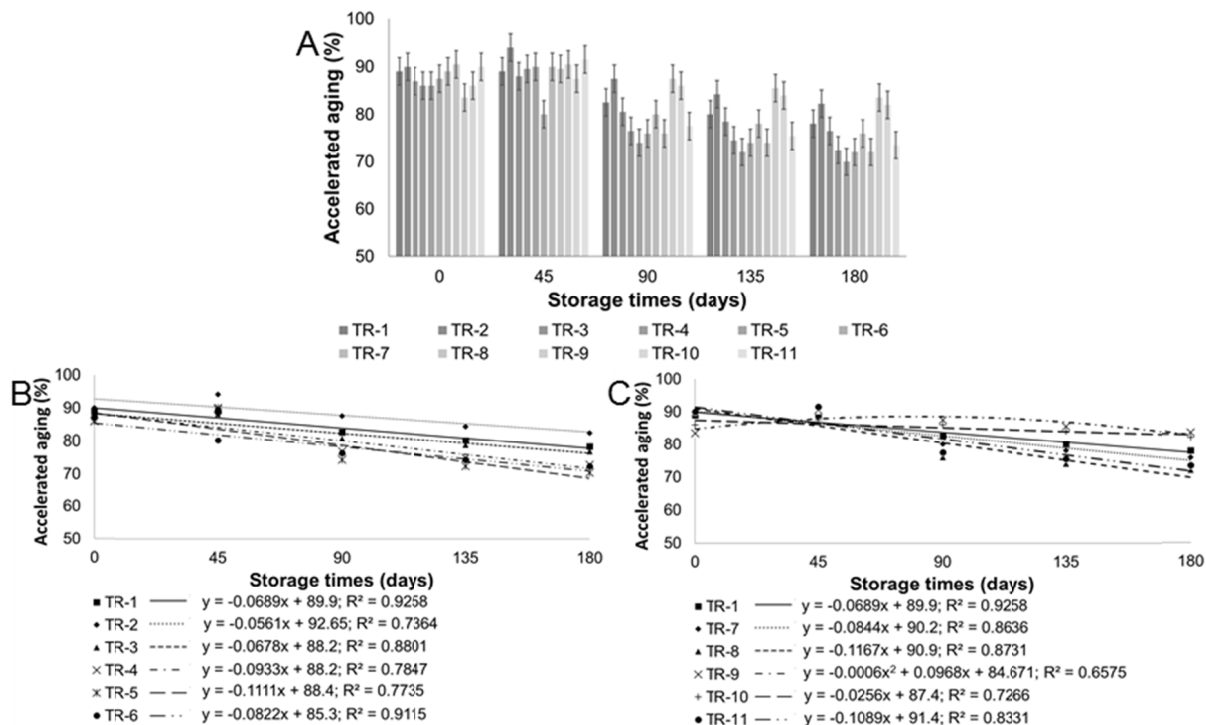


Figure 2. Vigor of soybean seeds cv. NS 6700 IPRO, evaluated by accelerated aging, in response to the interaction of 11 seed treatments with different chemicals (A) and the storage under uncontrolled conditions of temperature and relative humidity (B and C)

Note. TR-1: Control (without treatment); TR-2: Maxim XL (100 mL 100 kg⁻¹ of seeds); TR-3: NUF 323F1 535 (100 mL 100 kg⁻¹ of seeds); TR-4: NUF 323F1 535 FS (150 mL 100 kg⁻¹ of seeds); TR-5: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Standak Top (200 mL 100 kg⁻¹ of seeds); TR-6: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-7: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-8: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds); TR-9: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-10: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-11: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds). CV = 4.99%. In Figure 2A, bars correspond to the LSD value (least significant difference) between different seed treatments ($p < 0.05$), therefore bars which overlap are statistically equal.

Seedling emergence in sand showed interaction between the factors tested, according to the analysis of variance. In Figure 3A the response of the treatments at each storage time is presented. At time zero, the control (TR-1) presented the lower number of emerged seeds. On the other hand, at 45 and 90 days there were no significant differences between treatments studied. At the 135 days of storage, TR-7 was statistically inferior to the remaining treatments while at 180 days of storage, treatments TR-1 and TR-6 were significantly inferior to the other treatments, where treatments TR-8 and TR-10 presented the greater percentage of emerged seeds.

The behavior of seedling emergence in sand throughout storage demonstrated a quadratic negative model, reaching a maximum emergence of 96% at the 63 days of storage. Thereafter, emergence reduced overtime (Figure 3B). Treatments TR-2, TR-5 and TR-6 adjusted to a negative linear model, demonstrating rates of reduction of emergence in sand of 0.06; 0.04 and 0.08% day⁻¹, respectively. The remaining treatments did not adjust to any mathematic models (Figures 3B and 3C).

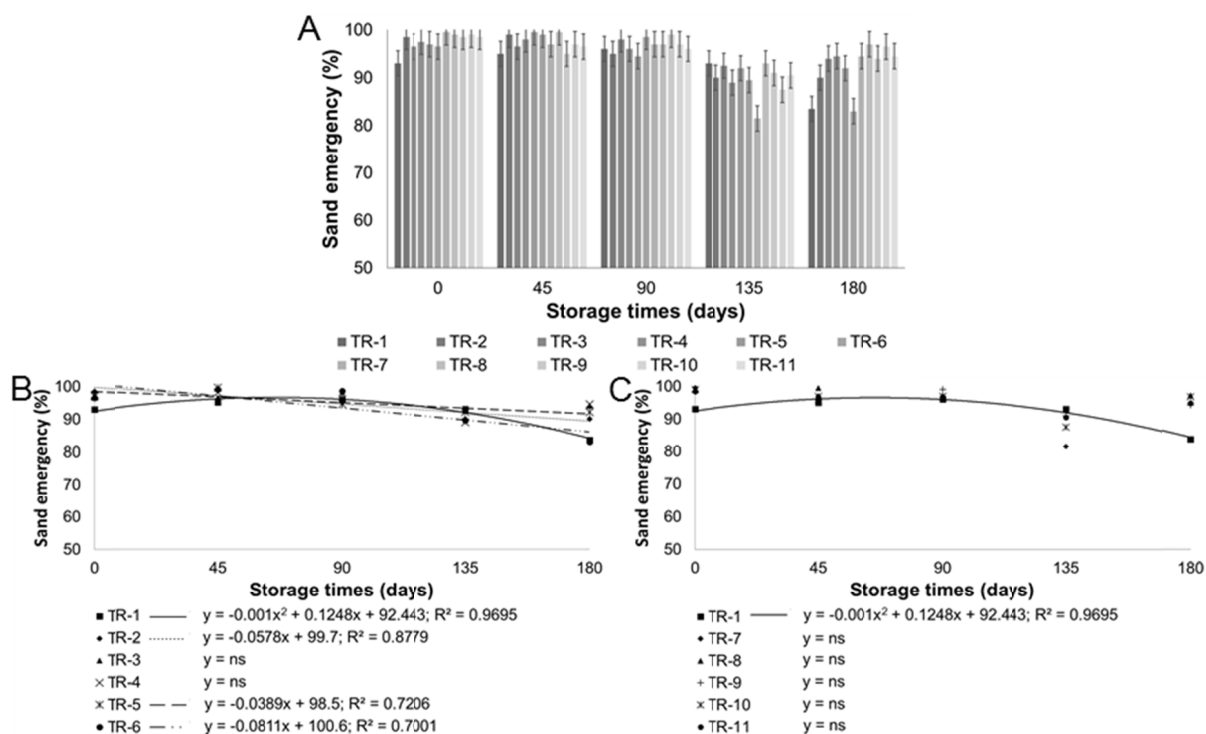


Figure 3. Vigor of soybean seeds cv. NS 6700 IPRO, evaluated by seedling emergence in sand, in response to the interaction of 11 seed treatments with different chemicals (A) and the storage under uncontrolled conditions of temperature and relative humidity (B and C)

Note. TR-1: Control (without treatment); TR-2: Maxim XL (100 mL 100 kg⁻¹ of seeds); TR-3: NUF 323F1 535 (100 mL 100 kg⁻¹ of seeds); TR-4: NUF 323F1 535 FS (150 mL 100 kg⁻¹ of seeds); TR-5: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Standak Top (200 mL 100 kg⁻¹ of seeds); TR-6: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-7: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-8: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds); TR-9: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-10: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-11: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds). CV = 3.98%. In Figure 3A, bars correspond to the LSD value (least significant difference) between different seed treatments (p < 0.05), therefore bars which overlap are statistically equal.

The speed of emergence index (SEI) also showed response to the interaction between the factors studied, according to the analysis of variance. In storage time zero TR-2 presented the greater SEI, followed by TR-1 and TR-4. TR-7, in contrast, demonstrated the lowest SEI of all treatments. At 45 days, TR-2 presented the higher SEI, followed by TR-9 and TR-1 while TR-11 and TR-3 presented the lower SEI. At 90 days, TR-3, TR-6, TR-2 and TR-1 presented the higher SEI while TR-11 and TR-8 presented the lower performance. At 135 days, TR-1, TR-3 and TR-2 presented the higher SEI and TR-4, TR-8 and TR-7 the lower. Lastly, at 180 days TR-2, TR-10, TR-9 and TR-3 outperformed the remaining treatments wherein TR-11, TR-1 and TR-6 presented the lower performances (Figure 4A).

The values of SEI presented a gradual decrease throughout storage (Figures 4B and 4C) which can be explained by a quadratic model in treatments TR-1, TR-2 and TR-9, reaching maximum values of 27.61; 28.19 and 26.74 at the 42, 17 and 53 days of storage, respectively.

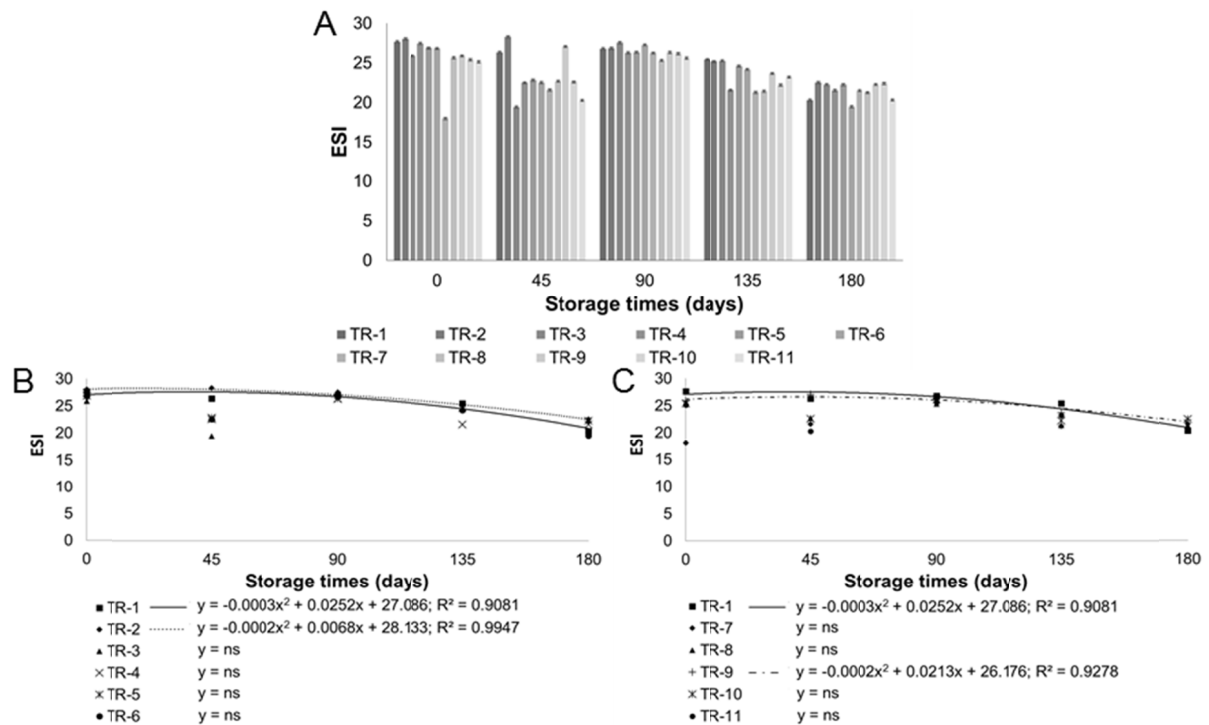


Figure 4. Vigor of soybean seeds cv. NS 6700 IPRO, evaluated by the speed of emergence index, in response to the interaction of 11 seed treatments with different chemicals (A) and storage under uncontrolled conditions of temperature and relative humidity (B and C)

Note. TR-1: Control (without treatment); TR-2: Maxim XL (100 mL 100 kg⁻¹ of seeds); TR-3: NUF 323F1 535 (100 mL 100 kg⁻¹ of seeds); TR-4: NUF 323F1 535 FS (150 mL 100 kg⁻¹ of seeds); TR-5: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Standak Top (200 mL 100 kg⁻¹ of seeds); TR-6: Maxim XL (100 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-7: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds); TR-8: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds); TR-9: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-10: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds); TR-11: NUF 323F1 535 FS (75 mL 100 kg⁻¹ of seeds) + Maestro FS (200 mL 100 kg⁻¹ of seeds) + Film coater (100 mL 100 kg⁻¹ of seeds). CV = 0.68%. In Figure 4A, bars correspond to the LSD value (least significant difference) between different seed treatments ($p < 0.05$), therefore bars which overlap are statistically equal.

The results of the analysis of variance for speed of emergence demonstrated the simple effect of the factor time of storage. Figure 5 shows the behavior of this variable throughout storage which was adjusted to a positive linear model, with the increase in the days required for germination at a rate of 0.0127 days for each day of storage. However, this result must be interpreted cautiously as the coefficient of determination (R^2) obtained indicates that only 33% of the response variable variation observed is explained by the model proposed.

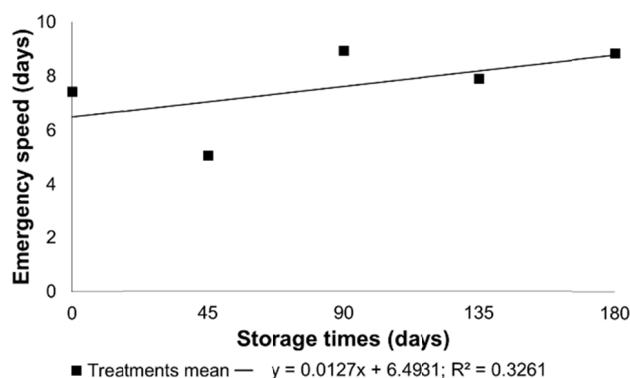


Figure 5. Speed of emergence of soybean seeds cv. NS 6700 IPRO, treated with different chemicals in response to storage under uncontrolled conditions of temperature and relative humidity

4. Discussion

Generally, the results obtained demonstrate that seed physiological quality was differentially affected by allseed treatments applied, wherein each vigor test performed entailed different behaviors for each seed treatment tested. However, the effect of storage time in seed physiological quality was remarkable, causing vigor decrease on all variables studied independently of the seed treatment tested. The results reported in this study corroborate with the findings of Dan et al. (2010), which evaluated the effect seed treatment with insecticides in the physiological quality of soybean seeds during storage and verified the decrease. Almeida et al. (2014) observed that vigor, which was measured by the accelerated aging test, of soybean seeds treated with various fungicides and insecticides before storage decreased with the increase of storage time. Likewise, Piccinin et al. (2013) detected the decrease of seed physiological quality of soybean seeds treated with fipronil and thiamethoxam and stored for 180 days. According to Horii and Shetty (2007), seed treatment with chemical products, in general, causes the reduction of the speed of emergence index (SEI) as the storage time increases. On the other hand, Pires et al. (2004) reported that seed treatment with polymers and fungicides did not affect bean seeds germination during storage for 120 days. Furthermore, Krohn and Malavassi (2004), which tested the effect of soybean seed treatment with fungicides before storage, did not observe the impairment of germination, however, emergence reduced when seeds were stored for more than 4 months after treatment.

Despite the statistical differences between seed treatments in each storage time evaluated and throughout storage, in this experiment there were no germination percentages inferior to 80%, which is the minimum value required by legislation for soybean seed commercialization in Brazil. Therefore, seed industrial treatment may be performed before storage using any of the mixture tested and without restrictions if seeds are stored for up to 180 days. Furthermore, the differences observed between each treatment tested may be attributed to the different active ingredients present in each pesticide. According to Ludwig et al. (2011) the decrease of germination of seeds treated with pesticides may be caused by phytotoxic effects of the active ingredient.

5. Conclusions

The industrial seed treatment in soybean before is storage for up to 180 days is viable using the mixtures of pesticides and storage conditions tested in this study.

All treatments tested contributed to the maintenance of seed quality throughout storage.

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