

Effect of Gibberellic Acid and Harvesting Time on the Seed Quality of Four Okra Cultivars

Ghadir Mohammadi¹, Ebrahim M. Khah¹, Spyridon Alexandros Petropoulos¹, Dimosthenis B. Chachalis², Fariba Akbari³ & Garip Yarsi⁴

¹ Department of Agriculture, Crop Production and Rural Environment, School of Agricultural Sciences, University of Thessaly, VOLOS, Greece

² Benaki Phytopathological Institute, ATHENS, Greece

³ Jihad-Agriculture Education Center of Sarpol-e-Zahab, Kermanshah, Iran

⁴ Plant and Animal Production Department, Vocational School of Silifke, University of Mersin, MERSIN, Turkey

Correspondence: Spyridon Alexandros Petropoulos, Department of Agriculture, Crop Production and Rural Environment, School of Agricultural Sciences, University of Thessaly, VOLOS, Greece. E-mail: fangio57gr@gmail.com

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Abstract

In an experiment at the University of Thessaly during the summer season of 2011, the effect of foliar application of gibberellic acid (GA₃) to okra at an early stage of plant growth (3-4 leaves) on plant growth, pod and seed characteristics was studied in relation to harvest time. GA₃ was applied at concentrations of 0 (Control), 50, and 100 mg L⁻¹ to four okra cultivars ('Boyatiou', 'Veloudo', 'Clemson' and 'Pylaias') and pods were harvested 30, 35, 40 and 50 days after anthesis (DAA) from the lower part of the plant. From the results it was found that GA₃ application increased plant height irrespective of cultivar and GA₃ concentration (50 and 100 mg L⁻¹), but without increasing flower induction or pod set. Similarly, GA₃ had no effect on pod dimensions (which were determined by genotype) or mean 100 seed weight, except in Boyatiou. Similarly, GA₃ application did not consistently affect seed moisture content, but it did however, increase the number of seeds per pod. Germination was either promoted ('Veloudo'), inhibited ('Boyatiou') or not affected ('Pylaias', 'Clemson') by GA₃. Differences in germination were apparently related to the incidence of hard seeds. Storage of seeds for 18 months improved germination. Overall, pod and seed characteristics were affected more by genotype and harvest time than by GA₃ application.

Keywords: gibberellic acid; okra; seed germination; seed moisture; seed yield; seed hardness

Abbreviations: GA₃ - gibberellic acid, DAA - days after anthesis, DAT - days after transplantation

1. Introduction

Gibberellic acid (GA₃) or gibberellins comprise a group of naturally occurring plant hormones which play a central role in the early germination processes of seeds by activating enzyme production and mobilizing storage reserves (Bewley & Black, 1983). Additionally, foliar application of gibberellins stimulates and synchronizes flowering and fruit set (Briant, 1974), as well as enhancing photosynthesis and growth (Yuan & Xu, 2001), or stimulating growth but not the rate of photosynthesis (Dijkstra & Kuiper, 1989).

Okra seeds frequently exhibit seed hardness and physical dormancy, which complicates crop management because of non-uniform germination after sowing resulting in a lack of crop uniformity and causing difficulties in weed control, harvest time etc. (Mohammadi, Khah, & Bannayan, 2011). Several publications have reported that soaking okra seeds in growth regulators, particularly gibberellin, increases germination (Nandpuri, Sooch, & Randhawa, 1969) possibly due to a reduction in seed hardness (Passam & Polyzou, 1997). On the other hand, Pal and Hossain (2001) found no effect of GA₃ on okra seed germination and seed weight, but a significant effect of this hormone on plant height and the number of pods per plant.

The positive effect of GA₃ on plant height has been widely reported for many crops, including okra (Ilias, Ouzounidou, Giannakoula, & Papadopoulou, 2007). Moreover, foliar application of GA₃ (20-40 ppm) and NAA

(50-100 ppm) on okra plants was found to increase the number of pods per plant and the number and weight of seeds (Asghar, Hussain & Ali, 1997; Abduljabbar, Abduljabbar, & Shukri, 2007). Similarly, Dhankhar and Singh (2009) reported that GA₃ application increased okra pod length and diameter. These results could be attributed mainly to the stimulatory effect of GA₃ on plant growth, which resulted in higher rates of biosynthesis and therefore higher amounts of assimilates available for distribution to the pods.

The objective of this study was to determine the effect of foliar application of gibberellic acid on the pod and seed quality of four okra genotypes in relation to the time of harvest and the position of the pods on the plant.

2. Material and Methods

2.1 Plant Material and Experimental Conditions

The experiment was carried out at the experimental farm of the University of Thessaly, Greece, throughout the growing season of 2011. The sand, silt and clay content of the soil at the experimental site was 48, 29 and 23%, respectively. The pH of the soil was 7.7-8.1 and the organic matter content was 1.3%. The soil was a clay loam Inceptisol, typic Xerochrept. Fertilization and crop management were carried out according to standard procedures. Fertilizers were applied as base and side dressing (a total of 30, 35 and 60 Kg ha⁻¹ of N, P and K respectively).

Gibberellic acid (GA₃) was applied as a foliar spray at three concentrations: GA₃(1) = 0.5 g GA₃ in 1 litre water; GA₃(2) = 1 g GA₃ in 1 litre water and GA₃(0) = 0 g GA₃ in 1 litre water to four cultivars of okra, namely cv. 'Clemson' (very productive with medium sized pods and seeds with high germination percentage), 'Boyatiou' (early and very productive cultivar with small, pentagonal fruits), 'Veloudo' (a very productive cultivar with small, pentagonal fruits and seeds with high germination percentage) and 'Pylaias' (very productive cultivar when irrigated, with small, pentagonal fruits suitable for processing).

The seeds of the four okra cultivars were sown in seed trays containing a peat-based substrate on 20 May and held under controlled temperature (20°C) until transplantation. Plants were transplanted to the field at the stage of 3-4 true leaves on 10 June. GA₃ was dissolved in water (GA₃(1) = 0.5 g GA₃ in 1 litre water; GA₃(2) = 1 g GA₃ in 1 litre water and GA₃(0) = 0 g GA₃) and applied to the plants twice: (1) at a plant height of approximately 50 cm, (2) 10 days after the first application. All plants were sprayed at the same time of application. The crop was irrigated regularly on a weekly basis with a drip irrigation system, whereas for weed control, Fusillade herbicide (fluazifop-P-butyl, 800 ml/1000m²) was applied when needed. Anthesis was first recorded 29 days after transplantation (DAT), and from the on each flower was tagged on emergence and its position and the date of flowering were recorded. Harvesting was carried out at four different time periods after anthesis (30, 35, 40 and 45 DAA) for seed quality tests.

2.2 Measurements

Seed germination tests were conducted using 100 seeds per treatment, with four replications. The germination tests were carried out on seeds after extraction from the pods either immediately after harvest. Just prior to the germination tests, seeds were surface sterilized first with Chlorine Dioxide, 1 to 20 parts deionised water for 10 sec. and then Mancozeb fungicide (86 %) and placed in sterilized Petri dishes on a double layer of Whatman No. 1 filter paper moistened with distilled water. The Petri dishes were placed in the dark in an incubator at 25 °C for 21 days and every 2 days germinated seeds were counted and removed and, in the case of moisture deficiency, distilled water was added. Seeds were considered germinated when the tip of the radical had grown free of the seed coat (Wiese & Binning, 1987; Auld et al., 1988). Those seeds which did not germinate and remained rigid after the time period of 21 days in the incubator were considered to be hard seeds, but without testing by scarification (Passam and Polyzou, 1997) or another means, e.g. tetrazolium (Moore, 1985). Germination percentage was calculated by the following formula (ISTA, 2009).

The rate of germination, or germination speed, of the seed lots was derived from the formula:

$$\text{Mean Germination Time} = \Sigma n / \Sigma (n \times D_n) \times 100$$

$$\text{Rate of Germination} = 1/\text{mean germination Time}$$

Where, n is the number of seeds which germinated on day n. D is the number of days counted from the beginning of the germination test, Σ is the total days of germination period, R is speed or rate of germination. During cultivation, at each harvest the following parameters were recorded: total flower induction per plant, the number of pods set per plant, pod length and diameter (cm), the average number of seeds per pod, the 100 seeds weight (g) and plant height. Seed germination was tested before and after storage for 18 months at room temperature (25 °C).

2.3 Experimental Design and Statistical Analysis

The experiment was laid out according to a Randomized Complete Block design with factorial arrangement, with each block being replicated three times and consisting of 12 plots of 9 m² (3×3 m) each, i.e. a total of 36 plots with 30 plants per plot.

Statistical analysis was carried out with the aid of the S.A.S. statistical package (SAS Institute Inc., USA) and mean comparison was made according to Duncan's Multiple Range Test (DMRT) and the Least Significant Difference (LSD) test at $p \leq 0.05$. Graphs were generated by using Microsoft Excel software (Microsoft Corporation, USA).

3. Results

3.1 Climatic Conditions

The climatic conditions of the experimental site are given in Figure 1. The temperature gradually increased from May to July and then decreased till October. There was a difference of about 12-16 °C between minimum and maximum temperatures (in mean overall differences) throughout the experimental period. There was no rainfall during July, but during the other months varied from 23 to 43 mm; relative humidity thus decreased during July, but afterwards increased progressively. This means that as the number of days from anthesis increased (from 30-50 DAA) pods were increasingly exposed to lower temperatures and higher humidity.

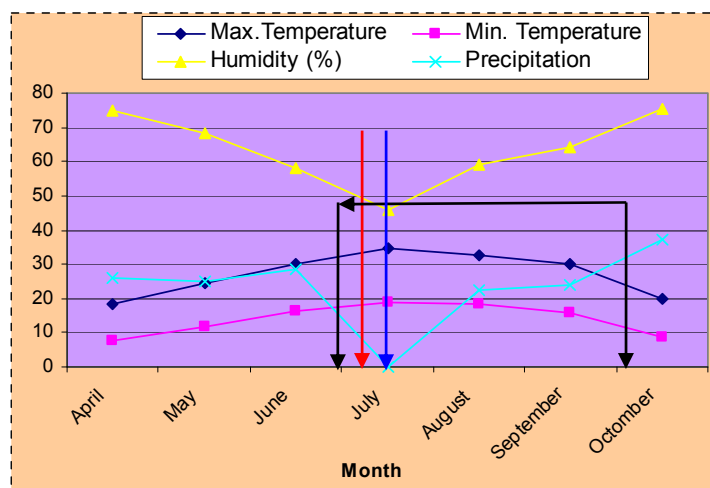


Figure 1. Climatic conditions during the experiment 2011 (black arrows show the duration of pod harvest, the red arrow shows the time of first GA₃ application and the blue arrow shows the time of second GA₃ application)

3.2 Effect of Gibberellic Acid on Flower Induction and Pod Set

The application of GA₃ had no significant effect on flower induction or the number of pods per plant in any of the cultivars (Table 1). Irrespective of GA₃ application, both flower induction and pod set were significantly higher in cv. 'Clemson' than in the other three cultivars.

Table 1. The effect of gibberellic acid (GA₃) on flower induction and pod set

Cultivar (C)	Gibberellic acid rate (mg L ⁻¹)(GA ₃)	Total flower induction (flowers/plant)	Pod set (pods/plant)
'Boyatiou'	GA ₃ (1)	13.0 ^a	11.2 ^a
	GA ₃ (2)	13.2 ^a	11.4 ^a
	GA ₃ (0)	11.8 ^a	9.9 ^a
<i>Mean</i>		12.6	10.8
LSD		2.7	2.8
'Veloudo'	GA ₃ (1)	11.5 ^a	10.2 ^a
	GA ₃ (2)	15.5 ^a	13.8 ^a
	GA ₃ (0)	14.8 ^a	13.1 ^a
<i>Mean</i>		13.9	12.3
LSD		4.2	4.3
'Pylaias'	GA ₃ (1)	12.0 ^a	10.9 ^a
	GA ₃ (2)	13.5 ^a	12.1 ^a
	GA ₃ (0)	12.1 ^a	10.6 ^a
<i>Mean</i>		12.5	11.2
LSD		3.9	3.8
'Clemson'	GA ₃ (1)	20.1 ^a	19.1 ^a
	GA ₃ (2)	20.0 ^a	18.5 ^a
	GA ₃ (0)	18.9 ^a	17.6 ^a
<i>Mean</i>		19.6	18.4
LSD		3.7	3.8
C x G			
	(C x GA ₃ (1))	*	*
	(C x GA ₃ (2))	*	*
	(C x GA ₃ (0))	*	*

Mean values for each cultivar separately in the same column followed by different letters differ significantly at P = 0.05.

*: statistically significant (p<0.05); ns: not significant.

GA₃(1) = 50 mg L⁻¹ Gibberellic acid; GA₃(2) = 100 mg L⁻¹ Gibberellic acid; GA₃(0) = 0 mg L⁻¹ Gibberellic acid.

3.2 Effect of Gibberellic Acid on Pod Length and Diameter

Despite occasional differences, GA₃ had no significant effect on either pod length or pod diameter, irrespective of the time of harvest (Table 2). Overall, pod length did not differ with harvest time, indicating that growth was maximal by 30 DAA, whereas in all cultivars there was a decrease in pod diameter at 40-50 DAA due to progressive drying of the pods. 'Clemson', which had the highest number of pods per plant (Table 1), also had the smallest pods, in terms of length and diameter (Table 2), perhaps as a result of higher inter-pod competition.

Table 2. The effect of gibberellic acid (GA₃) on dry pod dimensions (length and diameter) in relation to harvest time

Cultivar (C)	Gibberellic acid rate (mg L ⁻¹) (GA ₃)	Mean pod length (cm)				Mean pod diameter (cm)			
		30 DAA	35 DAA	40 DAA	50 DAA	30 DAA	35 DAA	40 DAA	50 DAA
'Boyiatiou'	GA ₃ (1)	17.1 ^a	16.4 ^a	16.8 ^a	17.1 ^a	2.2 ^a	1.7 ^a	1.9 ^a	1.8 ^a
	GA ₃ (2)	15.3 ^b	15.8 ^a	18.1 ^a	15.9 ^a	2.1 ^a	2.0 ^a	1.9 ^a	1.6 ^a
	GA ₃ (0)	15.5 ^b	15.1 ^a	16.9 ^a	16.8 ^a	2.0 ^a	2.0 ^a	1.8 ^a	1.9 ^a
	<i>Mean</i>	15.9	15.7	17.2	16.6	2.1	1.9	1.8	1.7
LSD		1.5	1.8	2.1	1.7	0.2	0.3	0.3	0.2
'Veloudo'	GA ₃ (1)	18.1 ^b	18.2 ^a	20.3 ^a	16.6 ^a	2.0 ^a	1.7 ^a	2.1 ^a	1.6 ^a
	GA ₃ (2)	19.4 ^a	17.3 ^a	22.1 ^a	17.3 ^a	1.9 ^a	1.6 ^a	2.1 ^a	1.6 ^a
	GA ₃ (0)	18.5 ^b	17.3 ^a	19.6 ^a	19.2 ^a	2.0 ^a	1.8 ^a	2.0 ^a	1.7 ^a
	<i>Mean</i>	18.6	17.6	20.6	17.7	1.9	1.7	2.0	1.6
LSD		0.8	2.7	3.7	2.2	0.1	0.3	0.3	0.2
'Pylaias'	GA ₃ (1)	18.0 ^a	19.6 ^b	19.5 ^a	18.1 ^{ab}	2.0 ^a	2.0 ^a	1.8 ^b	1.7 ^a
	GA ₃ (2)	18.4 ^a	24.9 ^a	18.4 ^a	16.0 ^b	2.0 ^a	2.0 ^a	1.9 ^{ab}	1.6 ^a
	GA ₃ (0)	16.7 ^a	18.8 ^b	18.6 ^a	20.4 ^a	1.9 ^a	2.0 ^a	2.0 ^a	1.8 ^a
	<i>Mean</i>	17.7	21.1	18.8	18.1	1.9	2.0	1.9	1.7
LSD		2.3	1.9	3.6	3.5	0.2	0.3	0.1	0.2
'Clemson'	GA ₃ (1)	13.9 ^a	15.0 ^a	14.2 ^c	15.9 ^a	1.4 ^a	1.5 ^a	1.2 ^a	1.3 ^a
	GA ₃ (2)	14.1 ^a	14.9 ^a	15.6 ^b	15.7 ^a	1.5 ^a	1.5 ^a	1.3 ^a	1.2 ^a
	GA ₃ (0)	14.9 ^a	14.5 ^a	18.5 ^a	16.0 ^a	1.5 ^a	1.5 ^a	1.3 ^a	1.1 ^a
	<i>Mean</i>	14.3	14.8	16.1	15.8	1.4	1.5	1.2	1.2
LSD		1.5	2.5	0.9	1.4	0.1	0.4	0.1	0.1
C x G									
	(C x GA ₃ (1))	*	*	*	*	*	*	*	*
	(C x GA ₃ (2))	*	*	*	*	*	*	*	*
	(C x GA ₃ (0))	*	*	*	*	*	*	*	*

Mean values for each cultivar separately in the same column followed by different letters differ significantly at P = 0.05.

*: statistically significant (p<0.05); ns: not significant.

GA₃(1) = 50 mg L⁻¹ Gibberellic acid; GA₃(2) = 100 mg L⁻¹ Gibberellic acid; GA₃(0) = 0 mg L⁻¹ Gibberellic acid.

3.3 Effect of Gibberellic Acid on Plant Height

GA₃ application increased the mean plant height of all cultivars tested in the present study, although in cv. 'Pylaias' the increase observed was not statistically significant (Table 3). Plant height was significantly lower in 'Clemson' than in the other three cultivars. In both 'Clemson' and 'Boyiatiou' plant height was significantly increased by GA₃ at both concentrations, whereas in 'Veloudo' the increase was only significant at the higher GA₃ level (GA₃(2)).

Table 3. The effect of gibberellic acid (GA₃) on mean plant height

Cultivar (C)	Gibberellic acid rate (mg L ⁻¹) (GA ₃)	Plant height (cm)
'Boyiatiou'	GA ₃ (1)	230.0 ^a
	GA ₃ (2)	240.0 ^a
	GA ₃ (0)	153.0 ^b
<i>Mean</i>		207.0
LSD		65.5
'Veloudo'	GA ₃ (1)	219.0 ^{ab}
	GA ₃ (2)	242.0 ^a
	GA ₃ (0)	206.0 ^b
<i>Mean</i>		222.0
LSD		32.0
'Pylaias'	GA ₃ (1)	188.0 ^a
	GA ₃ (2)	200.0 ^a
	GA ₃ (0)	178.0 ^a
<i>Mean</i>		188.0
LSD		36.0
'Clemson'	GA ₃ (1)	162.0 ^a
	GA ₃ (2)	180.0 ^a
	GA ₃ (0)	74.0 ^b
<i>Mean</i>		138.0
LSD		42.8
C x G		
	(C x GA ₃ (1))	*
	(C x GA ₃ (2))	*
	(C x GA ₃ (0))	*

Mean values for each cultivar separately in the same column followed by different letters differ significantly at P = 0.05.

*: statistically significant (p<0.05); ns: not significant.

GA₃(1) = 50 mg L⁻¹ Gibberellic acid; GA₃(2) = 100 mg L⁻¹ Gibberellic acid; GA₃(0) = 0 mg L⁻¹ Gibberellic acid.

3.4 Effect of Gibberellic Acid on the Number of Seeds Per Pod the 100 Seed Weight

In all cultivars, the application of GA₃ significantly increased the number of seeds per pod, irrespective of the time of harvest (Table 4). GA₃ was equally effective at both concentrations used in this study. In 'Boyiatiou' and 'Veloudo' the number of seeds per pod also tended to increase with increasing pod age at harvest.

The 100 seed weight of 'Pylaias', 'Clemson' and 'Veloudo' was not affected by GA₃ application, irrespective of the time of harvest (Table 4). In 'Boyiatiou', there was no effect of GA₃ on the mean 100 seed weight of seeds harvested 30 DAA, but thereafter the seeds from the control GA₃(0) had a higher 100 seed weight than those of seeds from the GA₃ treatments, especially that with the high GA₃ level (GA₃(2)). It is likely that the decrease in seed size (100 seed weight) correlated with the mean seed number, i.e. as seed number increased, mean 100 seed weight decreased.

Table 4. The effect of gibberellic acid on the number of seeds per pod and the 100 seed weight in relation to harvesting time

Cultivar (C)	Gibberellic acid rate (mg L ⁻¹)(GA ₃)	Mean number of seeds/pod				Mean 100 seed weight (g)			
		30 DAA	35 DAA	40 DAA	50 DAA	30 DAA	35 DAA	40 DAA	50 DAA
'Boyiatiou'	GA ₃ (1)	42.9 ^a	58.0 ^a	55.7 ^a	56.1 ^a	21.3 ^a	26.3 ^a	32.0 ^c	19.2 ^b
	GA ₃ (2)	51.8 ^a	52.8 ^a	58.8 ^a	60.7 ^a	19.5 ^a	17.7 ^b	34.1 ^b	19.6 ^b
	GA ₃ (0)	25.1 ^b	24.7 ^b	31.8 ^b	20.5 ^b	21.3 ^a	26.8 ^a	37.5 ^a	23.3 ^a
<i>Mean</i>		39.9	45.1	48.7	45.7	34.5	23.6	20.7	20.7
LSD		15.6	17.7	19.4	15.4	4.0	4.4	1.0	3.1
'Veludo'	GA ₃ (1)	34.2 ^a	43.2 ^a	40.5 ^a	58.8 ^a	25.6 ^a	29.0 ^a	39.7 ^a	22.4 ^a
	GA ₃ (2)	46.2 ^a	45.3 ^a	54.4 ^a	51.8 ^a	24.9 ^a	22.7 ^b	41.8 ^a	23.4 ^a
	GA ₃ (0)	15.7 ^b	19.3 ^b	22.8 ^b	29.2 ^b	26.8 ^a	33.5 ^a	39.4 ^a	23.7 ^a
<i>Mean</i>		32.0	35.9	39.2	46.6	40.3	28.4	25.7	23.1
LSD		12.7	14.1	16.5	11.7	3.9	4.6	3.2	1.4
'Pylaias'	GA ₃ (1)	58.9 ^a	54.5 ^a	52.6 ^a	48.1 ^a	22.7 ^a	32.5 ^a	39.4 ^a	25.4 ^a
	GA ₃ (2)	57.9 ^a	57.1 ^a	53.8 ^a	54.7 ^a	23.4 ^a	34.4 ^a	40.0 ^a	24.2 ^a
	GA ₃ (0)	31.0 ^b	27.9 ^b	29.6 ^b	25.5 ^b	21.5 ^a	33.2 ^a	38.5 ^a	25.9 ^a
<i>Mean</i>		49.2	46.5	45.3	42.7	39.3	33.3	22.5	25.1
LSD		13.0	12.6	11.5	10.5	3.4	9.2	4.6	2.1
'Clemson'	GA ₃ (1)	72.4 ^a	54.1 ^a	51.7 ^a	64.0 ^a	21.9 ^a	26.2 ^a	38.6 ^a	23.5 ^a
	GA ₃ (2)	62.8 ^a	62.5 ^a	62.5 ^a	60.1 ^a	21.4 ^a	26.5 ^a	36.4 ^{ab}	23.1 ^a
	GA ₃ (0)	28.6 ^b	28.8 ^b	29.6 ^b	34.0 ^b	20.3 ^a	20.8 ^b	34.8 ^b	23.6 ^a
<i>Mean</i>		54.6	23.5	47.9	52.7	36.6	24.5	21.2	23.4
LSD		13.7	11.5	12.3	14.2	3.1	3.2	3.4	5.9
C x G									
	(C x GA ₃ (1))	*	*	*	*	*	*	*	*
	(C x GA ₃ (2))	ns	ns	*	*	*	*	*	*
	(C x GA ₃ (0))	*	ns	ns	*	*	*	*	ns

Mean values for each cultivar separately in the same column followed by different letters differ significantly at P = 0.05.

*: statistically significant (p<0.05); ns: not significant.

GA₃(1) = 50 mg L⁻¹ Gibberellic acid; GA₃(2) = 100 mg L⁻¹ Gibberellic acid; GA₃(0) = 0 mg L⁻¹ Gibberellic acid.

3.5 Effect of Gibberellic Acid on Seed Germination, Seed Moisture Content and the Incidence of Hard Seeds

The moisture content of seeds harvested 30 DAA was very high in all cultivars (49-55%), but progressively decreased as harvesting was delayed, so that at 50DDA moisture content was just 12-13% (Table 5). There was no consistent effect of GA₃ application on seed moisture content, although at 50 DAA the moisture content of seeds from the (GA₃(2))treatment of 'Veludo' and (GA₃(1)) and (GA₃(2))treatments of 'Clemson' was significantly lower than that of the control (GA₃(0)).

In 'Boyiatiou' seed germination was highest for pods harvested 35 DAA and decreased significantly for pods harvested 50 DAA. In the other cultivars, however, seed germination was low for pods harvested at 30 DAA, but higher for pods harvested at later dates (35-50 DAA) (Table 5). In 'Boyiatiou' germination was significantly lower in seeds harvested from plants treated with the lower GA₃ concentration (GA₃(1)). In contrast, in 'Veludo' germination was significantly higher in both GA₃ treatments compared with the control, irrespective of harvest

time, whereas in ‘Pylaias’ and ‘Clemson,’ no effect of GA₃ on seed germination was detected. The incidence of hard seeds was highest in ‘Boyiatiou’ and lowest in ‘Clemson’. In ‘Veloudo’ there appeared to be a reduction in the percentage of hard seeds within the GA₃ treatments, which may account for the positive effect of GA₃ on germination in this cultivar (Table 5).

Table 5. The effect of gibberellic acid on seed moisture content at harvest, seed germination and seed hardness in relation to harvesting time

Cultivar (C)	Gibberellic acid rate (mg L ⁻¹)(GA ₃)	Mean moisture content at harvest (%)				Mean germination (%)				Seed hardness (%)
		30 DAA	35 DAA	40 DAA	50 DAA	30 DAA	35 DAA	40 DAA	50 DAA	30-35-40-50 DDA
‘Boyiatiou’	GA ₃ (1)	48.1 ^b	34.7 ^a	14.5 ^a	13.5 ^a	26.4 ^b	48.9 ^b	40.5 ^b	23.0 ^b	54.8
	GA ₃ (2)	52.8 ^a	20.3 ^b	12.4 ^a	13.9 ^a	38.8 ^a	69.0 ^a	64.2 ^a	31.3 ^a	48.7
	GA ₃ (0)	48.7 ^b	30.0 ^a	15.2 ^a	12.9 ^a	40.5 ^a	70.5 ^a	61.6 ^a	33.1 ^a	45.0
	<i>Mean</i>	<i>49.8</i>	<i>28.3</i>	<i>14.0</i>	<i>13.4</i>	<i>35.2</i>	<i>62.8</i>	<i>55.4</i>	<i>29.1</i>	<i>49.5</i>
LSD		4.2	8.7	4.2	2.1	5.9	6.2	6.9	5.2	
‘Veloudo’	GA ₃ (1)	51.8 ^c	35.6 ^a	19.4 ^a	12.6 ^a	34.8 ^a	72.6 ^a	72.1 ^a	72.1 ^a	20.0
	GA ₃ (2)	55.8 ^a	25.3 ^b	18.4 ^a	10.6 ^b	35.7 ^a	70.6 ^a	72.2 ^a	74.3 ^a	17.7
	GA ₃ (0)	53.8 ^b	41.6 ^a	22.2 ^a	12.9 ^a	23.5 ^b	46.6 ^b	52.2 ^b	44.9 ^b	32.8
	<i>Mean</i>	<i>53.8</i>	<i>34.1</i>	<i>20.0</i>	<i>12.0</i>	<i>31.3</i>	<i>63.2</i>	<i>65.5</i>	<i>63.7</i>	<i>23.5</i>
LSD		1.6	10.0	8.7	1.8	4.9	5.4	5.9	6.3	
‘Pylaias’	GA ₃ (1)	53.5 ^a	38.4 ^a	18.4 ^a	11.7 ^a	38.7 ^a	84.1 ^a	86.1 ^a	85.4 ^a	10.6
	GA ₃ (2)	53.8 ^a	41.8 ^a	17.2 ^a	12.2 ^a	39.6 ^a	84.0 ^a	87.7 ^a	83.6 ^a	12.0
	GA ₃ (0)	54.3 ^a	44.1 ^a	17.8 ^a	12.9 ^a	43.5 ^a	88.3 ^a	86.1 ^a	80.6 ^a	14.0
	<i>Mean</i>	<i>53.8</i>	<i>41.4</i>	<i>17.8</i>	<i>12.2</i>	<i>40.6</i>	<i>85.4</i>	<i>86.6</i>	<i>83.2</i>	<i>12.2</i>
LSD		2.6	6.3	8.5	1.3	7.0	5.7	3.7	5.2	
‘Clemson’	GA ₃ (1)	55.1 ^a	28.6 ^a	12.9 ^b	11.9 ^b	37.5 ^b	77.3 ^a	91.0 ^b	88.7 ^a	5.3
	GA ₃ (2)	55.1 ^a	28.5 ^a	16.9 ^a	11.6 ^b	38.8 ^b	75.9 ^a	93.8 ^a	91.3 ^a	4.3
	GA ₃ (0)	54.0 ^a	33.0 ^a	12.8 ^b	13.1 ^a	60.9 ^a	77.9 ^a	94.9 ^a	91.3 ^a	4.8
	<i>Mean</i>	<i>54.7</i>	<i>30.0</i>	<i>14.2</i>	<i>12.2</i>	<i>45.7</i>	<i>77.0</i>	<i>93.2</i>	<i>90.4</i>	<i>4.8</i>
LSD		2.7	13.7	3.6	0.9	6.4	2.7	2.7	3.8	
C x G										
	(C x GA ₃ (1))	*	*	*	*	*	*	*	*	
	(C x GA ₃ (2))	ns	*	*	*	*	*	*	*	
	(C x GA ₃ (0))	*	*	*	*	*	*	*	*	

Mean values for each cultivar separately in the same column followed by different letters differ significantly at P = 0.05.

*: statistically significant (p<0.05); ns: not significant.

GA₃(1) = 50 mg L⁻¹ Gibberellic acid; GA₃(2) = 100 mg L⁻¹ Gibberellic acid; GA₃(0) = 0 mg L⁻¹ Gibberellic acid.

3.6 Effect of Storage On Seed Germination

The storage of seeds for 18 months remarkably improved the germination of all cultivars, but especially ‘Veloudo’ and ‘Boyiatiou’ where the increase in germination after storage was statistically significant (increased by 41% and 33 % respectively) (Figure 2).

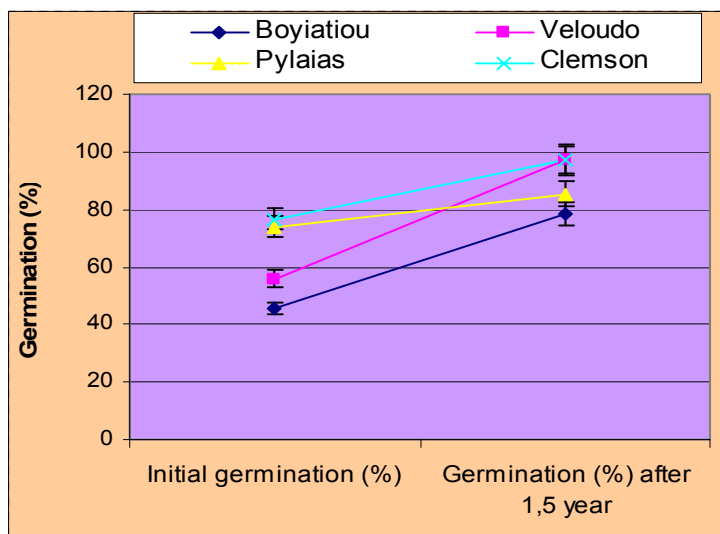


Figure 2. Seed germination percentage at harvest time and after 18 months storage at room temperature of 25 °C (pooled data for all treatments)

4. Discussion

Although GA₃ application at an early stage in okra growth resulted in a significant increase in plant height (Table 3), it did not cause a parallel increase in flower induction or pod set (Table 1). Similarly, GA₃ did not significantly affect pod dimensions (length and diameter) (Table 2), which were primarily determined by the genotype (cultivar), but it did significantly increase the number of seeds per pod, all at the expense of seed size in ‘Boyatiou’ (Table 4).

The positive effect of gibberellin on plant height is well documented for okra and for other species. In okra, GA₃ increases plant height by increasing intermodal length, but has no effect on the number of nodes per plant. Since flowers are induced singly at each node (Dhingra, 2009), there is therefore no effect of GA₃ on flower or pod induction. Similarly, the application of Gibberellin inhibitors, e.g. chlormequat chloride, reduce internode length and therefore plant height, but with no positive effect on node or flower induction (Thanopoulos, Petropoulos, Alexopoulos, Karapanos, Khah, Akoumianakis & Passam, 2013). The effect of GA₃ may be exerted from an early stage in plant growth, such as the seedling (Pal and Hossain, 2001) or even the seed (Das & Pattanaik, 1971). In other studies, GA₃ has been found to have no effect on pod induction (Syed, Hussain, Nawab, Asghar & Ali, 1997), although an increase in flowering, but not the percentage of fruit set, was observed when GA₃ was applied as a seed soak (Nandpuri et al., 1969). In contrast, Abduljabar et al. (2007) reported an increase in okra seed production when the plant apex was removed and GA₃ (100 ppm) applied one day later. The increase in seed yield was apparently a result of GA₃-promoted side shoot development and is therefore not comparable with the present experiment where the main shoot was allowed to grow continually.

The lack of effect of GA₃ on pod growth (length and diameter) is not surprising since this characteristic is influenced mainly by genotype (Raji, 1994). A decrease in pod size when harvesting is delayed (50 DAA) may be attributed to pod desiccation under the conditions of high temperatures and low humidity experienced in the field (Figure 1). Kokare, Bhalerao, Prabu, Chavan, Bansode and Kachare (2006) and Pal and Hossain (2001) also noted an absence of effect of plant growth regulators on okra pod growth, although Asghar et al. (1997) and Dhankhar and Singh (2009) suggested an increase in pod length and diameter following GA₃ application. In these studies, however, pods were harvested at a smaller size, i.e. before full maturation, as here. Moreover, the effect of GA₃ may depend on the timing and number of applications made, as shown by Ayyub, Ahmad and Akhtar (2013) who reported an increase in fresh pod yield when okra was repeatedly sprayed with GA throughout the growing season.

The principal effect of GA₃ on okra in the present experiment was the significant increase in the number of seeds per pod (Table 5). However, Bhatt and Srinivasa (1998) found no effect of GA₃ on the seed number per pod in okra, possibly due to the fact that they used a lower concentration of GA₃ (50 and 100 mg L⁻¹) compared with that used here. Although an increase in the number of seeds per pod is of immediate concern for seed producers, it may nevertheless result in lower seed quality, as indicated by the significantly lower 100 seed weights of seeds of ‘Boyatiou’ from the GA₃ treated plants (Table 5). Mean 100 seed weight at harvest depends mainly on the cultivar

and on the extent of seed drying. In the present experiment seeds harvested 50 DAA had a significantly lower mean 100 seed weight than those harvested earlier (especially at 30 DAA). Although Singh et al. (1999) indicated an increase in okra seed weight per pod following foliar application of GA₃ (20-40 ppm), this result was obtained when seeds were produced on plants that had already been harvested for fresh pods.

In general, there was little or no effect of GA₃ on seed moisture content at harvest. Seeds rapidly dried as pod harvesting was delayed, especially at 40-50 DAA. A negative effect of rapid drying under high field temperatures is an increase in the percentage of hard seeds (Demir, 1997). This is particularly the case in cv. 'Boyiatiou' and to some extent 'Veloudo'. In 'Veloudo' there was some indication that GA₃ may reduce the incidence of hard seeds, thereby improving seed quality, as shown by the higher germination of seeds from GA₃ treated plants in this cultivar (Table 5). A similar result has been reported for cv. other cultivars (Patil, Kadam, & Kolase, 2007) and cv. 'Balady' grown under winter conditions in Egypt. There was, however, no effect of GA₃ on seed germination (i.e. seed quality) in either 'Clemson' or 'Pylaias', whereas in 'Boyiatiou' germination was significantly lower in seeds harvested from plants treated with the lower GA₃ concentration (GA₃(1)). The reason for this effect of GA₃ is not clear since it occurred only at the lower GA₃ level (50 mg L⁻¹) and not the higher level (100 mg L⁻¹); it cannot therefore be a toxicity effect. It did, however, occurs at all stages of harvest and therefore requires further investigation.

It should be noted that because GA₃ was applied only once to plants at an early stage of growth, the results described here refer to pods formed on the lower part of the plant, where GA₃ would be expected to have maximum effect. It is known that the sensitivity of plants to GA₃ varies with the stage of growth at the time of application (Moore, 1979). Consequently, GA₃ applied to okra at the 3-4 leaf stage might be expected to affect pods and seeds formed early on in the life cycle of the plant (i.e. in the lower part of the mature plant) but not necessarily later on. When seed germination was recorded in pods from different positions on the plant (data not presented), it was observed that in all cultivars except 'Clemson' germination was higher in seeds from pods harvested from the lower part of the plant than those from pods in the upper part of the plant, irrespective of GA₃ treatment. Additionally, the percentage of hard seeds in all cultivars except 'Clemson' was higher in seeds from the upper part of the plant, clearly indicating that the reduction of germination observed here was due to an increase in seed hardness. We cannot, however, attribute these differences to GA₃ application because the pods from the lower and upper parts of the plant were harvested at different times and therefore subjected to different climatic conditions. Rather, it appears that pods and seeds from the upper plant part matured under higher temperatures and lower humidity; in consequence seeds dried more rapidly than those from the lower part of the plant, leading to the formation of a hard seed testa (Demir, 1997). These considerations were not necessarily taken into account in other experiments where GA₃ was applied to the seed or seedling and measurements taken at the end of the biological cycle of the crop (Das & Pattanaik, 1971; Singh et al., 1999).

5. Conclusion

Application of GA₃ at a concentration of 50-100 mg L⁻¹ to okra at the 3-4 leaf stage increased plant height irrespective of cultivar, but without increasing flower induction or pod set. Similarly, GA₃ had no effect on pod dimensions (length, diameter) or mean 100 seed weight, except in Boyiatiou. Similarly, GA₃ application did not consistently affect seed moisture content, but it did however, increase the number of seeds per pod. Germination was either promoted ('Veloudo'), inhibited ('Boyiatiou') or not affected ('Pylaias', 'Clemson') by GA₃. Differences in germination were apparently related to the incidence of hard seeds. Storage of seeds for 18 months improved germination. It therefore seems that foliar application of GA₃ may be beneficial for seed production of 'Veloudo' by increasing seed quality (germination), but is not of value for the other cultivars.

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