Anal Cancer Patients in a Metropolitan Hospital Present Infection with Multiple Oncogenic Viruses

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Abstract

Background: Oncoviruses such as HPV, KSHV, and EBV have been reported in patients with HIV/AIDS. Exactly how oncovirus-associated cancers rise in AIDS patients is not fully established. We highlight results from anal cancer samples that were screened for certain known oncoviruses in HIV/AIDS patients.

Methods: Tissues were obtained from 14 anal cancer patients. DNA was extracted from the tissues and used in PCR assays to identify the oncoviruses. Immunohistochemical assays were performed on slides using specific antibodies.

Results: Using PCR we detected HPV-16 (P<0.0001), HPV-45 (P<0.0001), HPV-31 (P<0.0001), HPV-33 (P<0.0001), and EBV (P<0.0001) which were statistically significantly different, and KSHV (P=0.090). Utilizing immunohistochemistry, the samples were statistically significantly different in HPV-16 E6 (P<0.0001), HPV-16 E7 (P<0.0001), EBV (P<0.0001), and KSHV (P<0.0001).

Conclusion: The findings indicate that oncogenic human papilloma and herpesviruses are present in anal cancer tissues. Their causal role to the cancer may have implications for long-term pathogenesis.

Keywords: HIV-1, EBV, HPV, KSHV, and Anal cancer, oncogenic virus

1. Introduction

It has been reported that greater than 15% of human cancers are currently associated with viruses that are major contributors to disease (zur Hausen, 1991). The oncogenic viruses possess different genomes, life cycles, and represent a number of virus families. The course from viral infection to tumor initiation is slow and may take years after primary infection. Virus infection is generally not sufficient for cancer initiation, with additional epigenetic events and host factors such as immunosuppression, somatic mutations, and genetic predisposition possibly contributing (Liao, 2006). We have shown association between herpesviruses and human papilloma viruses (HPV) with some of the cancers in our study (Simbiri et al., 2010). Sub-Saharan Africa and most of the developing countries have seen a spike in viral associated cancers in Human immunodeficiency virus (HIV) infected patients (Simbiri et al., 2010).

In this study we looked at the association of human papilloma and herpesviruses in anal cancer among

HIV-infected patients. Anal cancer is considered to be a non-AIDS defining cancer (NADC). It has been shown to be similar to cervical cancer, which is strongly associated with HPV infection and the presence of pre-cancerous lesions (Bernal et al., 2003; Cooley et al., 2003). The incidence in the HIV positive individuals is as high as 60 fold compared to the HIV negative individuals (Feigal, 2001; Frisch et al., 2001; Rabkin, 2001). An increased incidence of HPV-associated cancers has been observed among AIDS patients with increased elevated immunosuppression, likely a factor of prolonged survival that leads to enhanced risk of some HPV-associated cancers (Chaturvedi et al., 2009). Meanwhile, HPV-associated anal cancers in HIV-positive men increased in incidence despite the introduction of highly active antiretroviral therapy (HAART) (Kreuter & Wieland, 2009). Additionally, in a study of HIV, Epstein Barr virus (EBV), and Kaposi sarcoma herpes virus (KSHV) cooperation in the pathogenesis of human malignancies, a decrease of Kaposi's sarcoma (KS), non-Hodgkin's lymphoma (NHL), and primary central nervous system lymphoma (PCNSL) was noted, while Burkitt's lymphoma and Hodgkin's lymphoma decreased slightly under HAART (da Silva et al., 2011). Some studies have suggested that these NADCs have not become more prevalent (Romassi & Nagle, 2008; Silverberg & Abrams, 2007). However, it is evident that these cancers are increasing and are expected to rise in the coming years in the HIV population. The AIDS defining cancers (ADC) are mostly referred to as Kaposi's sarcoma, non-Hodgkin's Lymphoma and cervical cancer. The impact of HAART on the incidence of ADC and NADC among HIV/AIDS patients showed that the overall risk increased in NADCs after introduction of HAART, while the incidence of ADC decreased, and that the increased incidence of NADCs after early use of HAART was likely due to better control of viral replication, increased immunity, and prolonged survival (Cobucci et al., 2014; Pinzone et al., 2012; Kan et al., 2014).

The aim of this study was to identify the oncogenic human papilloma and herpesviruses in anal cancer tissues from patients in Pennsylvania Hospital, Philadelphia to show that oncogenic viruses may be involved in the pathogenesis of the cancer.

2. Materials and Methods

We identified 14 anal cancer tissue samples from Pennsylvania Hospital in Philadelphia, Pennsylvania, USA. University of Pennsylvania IRB approved the study (protocol # 808695). The study was conducted in compliance with the ethical principles of the Declaration of Helsinki. Patients included in the study were adults (age >18 years) with histologically proven anal carcinoma malignancy. The patients or their family members signed informed consent document (Table 1).

Case#	Diagnosis	Age	HIV	HCB/C	
1	Anal carcinoma	-	No		
2	Anal carcinoma-invasive squamous carcinoma	47	Yes		
3	Infiltrating moderately differentiated squamous carcinoma		No		
4	Invasive squamous cell carcinoma -rectal abscess		No		
5	Anal carcinoma	72	Yes		
6	Squamous cell carcinoma-anal	-	No		
7	Anal intraepithelial neoplasm	45	Yes	Hep B	
8	Anal cancer	54	Yes		
9	Anal cancer	47	No		
10	Anal carcinoma	53	Yes		
11	Kaposi sarcoma-anal carcinoma	29	Yes		
12	Kaposi sarcoma-anal carcinoma	44	Yes		
13	Kaposi sarcoma-anal carcinoma	-	No		
14	Hodgkin's lymphoma-anal carcinoma	-	No		

 Table 1. Anal Cancer patient characteristics

HIV status- (+) vs. (-), p=0.19793, odds ratio- 1.3059, 95% CI=0.0254 - 67.1672.

Table 1 shows diagnosis, age, HIV status, and hepatitis status of each patient in the study.

2.1 DNA Extraction

Total cellular DNA was extracted from anal tissues by digestion with proteinase K and phenol-chloroform extraction as previously described (Simbiri et al, 2010). Briefly, tissue samples were prepared by cutting 5 μ m thick sections using clean blades, followed by de-paraffinization and extraction of the DNA. PCR analysis was performed as previously described (Simbiri et al, 2010) with each virus specific primer including HPV types using consensus primer GP5+/GP6+ that detects more than 20 HPV high-risk (HR) and low- risk (LR) types, as well as EBV, KSHV, JC, and BK viruses.

2.2 Polymerase Chain Reaction

The PCR assay was performed with primers as indicated in Table 2. The PCR conditions were as follows: HPV-GP5+/GP6+ 94 \C -5 minutes, 94 \C -30 seconds, 48 \C -30 seconds, and 72 \C -30 seconds, for 40 cycles; elongation at 72 \C -5 minutes and incubation at 4 \C . EBV-94 \C -5 minutes and incubation at 4 \C . KSHV-94 \C -5 minutes, 94 \C -30 seconds, 47 \C -30 seconds, and 72 \C -30 seconds, for 40 cycles; elongation at 72 \C -5 minutes and incubation at 4 \C . KSHV-94 \C -5 minutes, 94 \C -30 seconds, 52 \C -30 seconds, and 72 \C -30 seconds, for 40 cycles; elongation at 72 \C -5 minutes and incubation at 4 \C . KSHV-94 \C -5 minutes and incubation at 4 \C . KSHV-94 \C -5 minutes and incubation at 4 \C . JC-94 $^{\circ}$ -5 minutes, 94 $^{\circ}$ -10 seconds, for 40 cycles; elongation at 72 \C -5 minutes and incubation at 4 \C . JC-94 $^{\circ}$ -5 minutes, 94 $^{\circ}$ -10 seconds, 58 $^{\circ}$ C-5 seconds, 72 $^{\circ}$ C-10 seconds, for 35 cycles; elongation at 72 \C -5 minutes and incubation at 4 \C . Annealing temperatures for specific HPV primers were adjusted accordingly. The PCR products were analyzed on 2.5% agarose gel at 100 V for 1 hour.

Primer	Sequence	Size (bp)
HPV-L1	5'-TTTGTTACTGTGGTAGATACTAC-3'	150
	3'-CTTATACTAAATGTCAAATAAAAAG-5'	
EBV-BamH1W	5'-CCAGACAGCAGCCAATTGTC-3'	129
	3'-GGTAGAAGACCCCCTCTTAC-5'	
KSHV-ORF-73	5'-CCATCTCTTGCATTGCCAC-3',	293
	5'-AACTACGGTTGGCGAAGTCA-3'	
JC VP2/VP3	5'-GAAGAACCCAAAACTATTTGTTGAAA-3',	133
	5'-GCCTAACTGGAGACAATCTAGAATAATAGTC-3'	
BK- Large T	5'-AAATCCATTTTATCTAATATATG-3,	370
	5'-CATATATTAGATAAAATGGATTT-3'	

Table 2. Primers used in PCR analysis.

Table 2 shows the primers, primer sequences, and product sizes in the PCR analysis.

2.3 Immunohistochemistry

Immunohistochemistry was performed as previously reported (Simbiri et al., 2010). Briefly, we used commercial antibodies HPV-16E6, HPV-16E7, and HPV-16E6/E7 (DAKO Inc., Carpentaria, CA), and monoclonal antibody S12 for EBV-LMP1, and monoclonal antibody derived from KSHV encoded LANA on 5 μ m slides as previously reported (Simbiri et al., 2010).

2.4 Data Analysis

The data for this study was collected in Excel and imported into a statistical software program. Analysis encompassed determination of proportions (MedCalc Software bvba MedCalc Software, Ostend, Belgium).

3. Results

We identified the oncogenic human papilloma and herpesviruses in the tissues derived from anal cancer. In this study using PCR and immunohistochemistry we identified human papilloma and herpesviruses in tissue samples from anal cancer patients (Table 3).

	EBV	HPV-G5+/6+	HPV-18	HPV-16	HPV-45	HPV-11	HPV-31	HPV-33	KSHV
1	+	-	-	+	-	+	-	-	+/-
2	+	+	+	+	-	+	+	+	+
3	+	+	+	+	+	-	+	-	+
4	+	-	-	+	-	+	-	-	-
5	+	-	-	+	-	-	-	-	-
6	+	+	+	+	-	+	+	+	+
7	+	+	-	+	-	-	+	-	-
8	+	+	+	+	+	-	+	-	+
9	+	-	-	+	-	+	+	-	-
10	+	+	+	+	-	+	+	-	-
11	+	-	-	+	+	-	+	+	-
12	+	+	+	+	-	+	+	-	+
13	+	+	-	+	-	+	-	-	-
14	+	-	-	+	-	-	-	-	-
Total	14	8	6	14	3	8	9	3	6
Percent	100%	56%	44%	100%	22%	56%	67%	22%	44%
OR	40401	1.6198	0.6173	40401	0.0796	1.6198	4.1221	0.0796	0.6173
CI	79-20651	0.92-2.83	0.35-1.07	793-20651	0.04-0.15	0.92-2.82	2.28-7.46	0.04-0.15	0.35-1.07
Р	< 0.0001	0.0905	0.0905	< 0.0001	< 0.0001	0.0905	< 0.0001	< 0.0001	0.0905

Table 3. Oncogenic viruses identified by PCR in anal cancer tissues

Table 3 shows oncogenic viruses identified in each anal cancer patient by PCR analysis. Most cases tested positive for HPV types, EBV, and KSHV.

Utilizing PCR we detected HPV using GP5+/GP6+ in 56% of the samples (P=0.091, OR 1.62, 95% CI 0.92-2.83); HPV-18 in 44% (P=0.091, OR 0.62, 95% CI 0.35-1.07); HPV-16 in 100% (p<0.0001, OR 40401, 95% CI 793-20651); HPV-45 in 22% (P<0.0001, OR 0.08, 95% CI 0.04-0.15); HPV-11 in 56% (P=0.09, OR 1.62, 95% CI 0.92-2.82); HPV-31 in 67% (P<0.0001, OR 4.12, 95% CI 2.28-7.46); HPV-33 in 22% (P<0.0001, OR 0.80, 95% CI 0.04-0.15); EBV in 100% (P<0.0001, OR 40401, 95% CI 793-20651); and KSHV in 44% (P=0.09, OR 0.62, 95% CI 0.35-1.07) (Table 3 and Figure 1).



Figure 1. Representative HPV-16 PCR for Anal cancer patients

Figure 1 shows a representative PCR result of anal samples processed using HPV-L1 primer. We observed that all the samples contained HPV genome. M-marker; $N - H_2O$ negative control; Samples, 1-14.

Further, when we used immunohistochemistry, we detected HPV-16 E6 in 88.9% (P<0.0001, OR 65.46, 95% CI 26.9-158.7); HPV-16 E7 was similar to HPV-16 E6 88.9% (P<0.0001, OR 65.46, 95% CI 26.9-158.7), and HPV-16 E6/E7 in100% (P<0.0001, OR 40401, 95% CI 793-20651). For EBV we detected 100% (P<0.0001, OR 40401, 95% CI 793-20651), and for KSHV we detected 77.8% (P<0.0001, OR 12.57, 95% CI 6.4-12.57) (Table 4 and Figure 2).

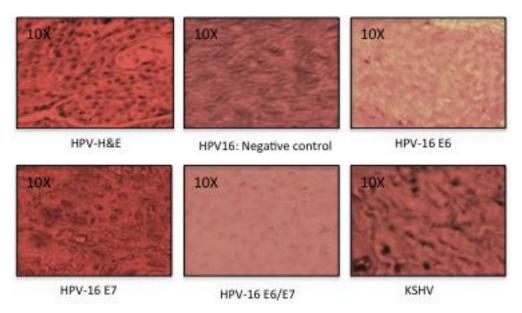


Figure 2. Representative Immunohistochemistry of oncoviral proteins detected in anal cancer samples

Figure 2 shows the immunohistochemistry results using specific antibodies. Positive cells stained dark brown. Some of the tissues did not stain robustly possibly due to status of the tissues.

Number of cases	EBV-S12	LANA	HPV-E6	HPV-E7	HPV-E6/E7	
1	++	-	+	-	++	
2	+	++	+	+	+++	
3	+	+	-	++	++	
4	+++	+	+	+	++	
5	++	+	++	+	++	
6	+++	++	++	++	++	
7	+	-	+	+++	+++	
8	++	+	+	+++	+	
9	+++	+	++	++	++	
10	++	+	++	++	++	
11	++	++	+++	++	+++	
12	+	++	++	++	+	
13	+++	+	++	++	++	
14	++	-	-	-	++	
Total	14	11	12	12	14	
Percent	100%	77.8%	88.9%	88.9%	100%	
OR	40401	12.57	65.46	65.46	40401	
CI	793-20561	6.4-12.57	26.9-158.7 26.9-158.7 79		793-20651	
Р	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	

Table 4. Oncogenic viruses identified by Immunohistochemistry in anal cancer tissues

Table 4 shows oncogenic viruses identified in each anal cancer patient as determined by IHC analysis. Most cases tested positive for EBV, KSHV, HPV-E6, and HPV-E6/E7.

High risk, low risk human papilloma viruses and herpesviruses were detected in anal cancer samples retrieved from a local hospital. We were interested in determining if similar oncogenic viruses that we had previously identified in samples from Botswana could be present in anal cancer samples. Thus, we screened samples from HIV patients from Pennsylvania hospital in Philadelphia, a local hospital, to determine if similarity in co-infection persisted. It was interesting to observe that the oncogenic viruses that we had previously identified in Botswana-based samples were also present in the few samples from the region, an indication that these viruses are prevalent, may be associated with the cancer, and may contribute to its pathogenesis through similar pathways. We observed significant differences for EBV, all HPV types except HPV-18, and no significant difference for KSHV using PCR (Table 3). However, on IHC there were significant differences for EBV, KSHV, HPV-16 E6 and E6/E7 (Table 4).

High-risk HPV types were detected in both HIV positive and negative samples. In all anal cancer samples we identified high risk HPV. Other HPV types were also identified (Table 3). These results suggest a high incidence of HPV in patients. The HPV may become opportunistic and oncogenic with long latency period once the patients are immunocompromised due to HIV.

EBV was consistently detected in anal cancer. As has been reported in previous studies (Simbiri et al, 2010), the ubiquitous EBV was again detected to a high degree in all the anal cancer samples (Table 3). In future studies, it will be interesting to identify the EBV type (EBV1 or EBV2) associated with the cancer, or if both EBV types co-infect and may contribute at different levels to pathogenesis.

The cancer was identified in young HIV infected patients. One recurring observation was the detection of the oncogenic viruses and anal cancer in younger HIV infected patients. This may be due to immunosuppression by HIV infection and other opportunistic infections at an early age, which renders their immune system to mount surveillance at low levels similar to that of older individuals who are more susceptible to the infections and associated diseases. The impact of age on the cancer patients with HIV warrants further investigation (Table 4).

4. Discussion

In this study we have shown the presence of oncogenic viruses in anal cancer tissues. We show the increase in incidence of the cancer in young HIV-infected patients, and the prevalence of the oncogenic human papilloma and herpesviruses in the cancer.

Our past study detected multiple tumor viruses in OSSN tissues. For the first time we showed the presence of HPV, EBV, KSHV, and CMV in ocular tissues in HIV/AIDS patients (Simbiri et al., 2010), vulvar cancer (Simbiri et al., 2014). Associations of viruses with human cancers have already been reported, such as KSHV with KS and PEL (Kiire & Dhillon, 2006); EBV with primary CNS lymphoma (Kestelyn et al., 1990), nasopharyngeal carcinoma (Cook-Mozaffari et al, 1998) and Burkitt's lymphoma (Cohen, 2000); HPV with cervical cancer (Mirzamani et al., 2006), anal cancer (Salit et al., 2009), head and neck (Goldenberg et al., 2001), and testicular cancer (Munoz et al., 2003). However, the contribution by HIV to the development of the malignancies remains unclear and may be through a number of different mechanisms that target variable cellular processes.

Viral proteins expressed in infected cells mediate infection and establishment of latency. Importantly, the immunomodulatory mechanisms used by one virus may actually enhance or benefit the activity or replication of other viruses. Many of these viruses (for example, HPV, CMV, and EBV) are quite common in the human population (Waddell et al., 1996) but do not always lead to development of malignancies in healthy individuals. The localization of these oncogenic proteins in ocular surface squamous neoplasia (OSSN) (Simbiri et al., 2010), vulvar cancer (Simbiri et al., 2014), and anal cancer tissues in our present study show association, however establishment of a causal effect by any of the viruses, or a multiple of these viruses in an HIV environment has yet to be fully confirmed. Determining the pathways involved in initiation and establishment of the cancers in HIV patients will give us a better understanding of the interplay of the different oncogenic viral proteins in the normal immune state and immunosuppressed state.

The localization of these tumor viruses in the different cancer tissue sections suggest that these viruses may functionally interact to contribute to OSSN, vulvar, and anal cancer phenotypes. Similar to our findings of HPV and EBV in ocular, vulvar, and anal tissues, Prayitno reported that in 19 cervical carcinoma samples 89% were positive for HPV and 68% were positive for EBV (Prayitno, 2006). It has been suggested that HPV probably does not act alone in initiating ocular neoplasia, but that other factors including the ultraviolet light are involved (Nakamura et al., 1997). Previous reports have suggested that infection with more than one HPV type enhances cervical cancer and oral squamous carcinoma (Premoli-De-Percoco et al., 1998). Mirzamani et al. reported co-infections with EBV and HPV-16/18 in nasopharyngeal carcinoma (NPC) patients and concluded that both

viruses were important in contributing to the pathogenesis of NPC (Mirzamani et al., 2006). The selection pressure that may exist as a consequence of competition between opportunistic infections may further lead to mutations occurring in the HPV types as well as in the other infectious agents that may contribute to oncogenesis.

Our findings in this study showed the presence of similar human papilloma and herpesviruses in the anal samples examined as was observed on OSSN and vulvar samples. HAART has been shown not to alter HPV persistence or rates of progression or regression of premalignant anogenital lesion (Salit et al., 2009; Palefsky et al., 2005; Heard et al., 2004). Palesfky et al. suggest that the increase in anal cancer in the HAART era may reflect the likelihood that in the pre-HAART era patients at highest risk of anal cancer would have died from other causes (Palefsky et al., 2005; Heard et al., 2004). Prior to the introduction of large scale HAART therapy in Botswana in 2002 (WHO, 2006) and other regions, most of the patients who had these oncogenic neoplasias pre-HAART era died without diagnosis or therapy. In future a longitudinal study that addresses the impact of HAART on anal cancer in relation to associated oncogenic viruses need to be initiated. Equally significant would be to investigate the effect of HIV on selection of other infections such as bacteria microbiome that may affect pathogenesis as was shown in our previous findings (Simbiri & Robertson, 2012).

Though this study had a small sample size it highlights the significance of human papilloma and herpesviruses in the pathogenesis of this cancer and other cancers in HIV/AIDS patients that will lead to new therapies and prevention strategies.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

KOS designed the study, conducted experiments, interpreted data, drafted the manuscript and supervised the study, HCJ interpreted data and drafted the manuscript, DHH identified anal cancer tissues and drafted the manuscript, and ESR designed the study, interpreted data, edited the manuscript, and supervised the study. All authors have read and approved the final manuscript.

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