

## Fluorescence and Reflectance Spectroscopy for Early Detection of Different Mycorrhized Plantain Plants

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### Abstract

Sustainable agriculture with use of Arbuscular Mycorrhizal Fungi (AMF) is an emerging farm management that improves crops nutrient and water use efficiency. Decision making on the effect of AMF is still dependent on agronomic diagnosis which is long, tedious, expensive and destructive. This study demonstrates the applicability of proximal fluorescence and reflectance spectroscopy for evaluating and detecting at early stage distinct types of mycorrhized plantain from two cultivars (*Musa paradisiaca*).

Visible-near infrared (400-1000 nm) reflectance and fluorescence data were collected from control and three levels mycorrhized plants designed in randomized and complete block under greenhouse conditions. Two spectral measurements at a week interval were performed on plant leaves by using an USB spectrometer mounted with an Arduino-based LED driver clip.

A new normalized reflectance water NWI5 index shows with Datt5 alone highly significant differences at  $P < 0.001$  respectively for Orishele and thia21 cultivars. dNIRmin920\_980, NDVI3 and GI reflectance index are significant at  $P < 0.01$ . Seven other reflectance and 3 fluorescence indices ANTH, FRF\_R and NBI\_R are significant at  $P < 0.05$ . The two first principal components for each cultivar spectral features explaining 94.1 % of variance were used to build predictive classification models. LogitBoost algorithm indicates accuracy of 90.27% on stratified cross-validation and 87.5% on test split. Our results confirm that fluorescence and reflectance spectroscopy is a valuable tool for early assessment of mycorrhization success rate evaluation and pattern recognition. They also show promise for the development of non-destructive and cost-effective detectors in monitoring crops under biofertilizers with arbuscular mycorrhizae.

**Keywords:** classification, fluorescence and reflectance spectroscopy, LogitBoost algorithm, mycorrhized plantain plants.

### 1. Introduction

Agricultural remote sensing has gained great importance in sustainable and intensification farming management. It generates digital data from sensors that help in reducing the risk and minimize damage of inappropriate fertilization. Indeed, low or excessive uses of chemical fertilizers can have high adverse impact on the environment and grain production system. Unused nitrogen released to the environment can have detrimental effects (Cameron, Di, & Moir, 2013). Large application of phosphate fertilizers and by-products has been practiced on arable lands, to improve crop production, induced soil nutrients deficiency, and increasing the levels of available S and P (Kassir, 2014). In the increasing demand of environmental sustainability, more attention of agricultural scientific community is being turned on biofertilizers particularly Arbuscular Mycorrhizal Fungi

(AMF) (Weber, 2014; Sahraoui, 2013). Thus investigating the potential output of such biological microorganisms on crops with use of remote sensing tools is highly relevant.

Controlled endomycorrhization is a current problem that stayed long time ago in agriculture, horticulture and arboriculture (Gianinazzi, 1981). It was recently reported that diversity of plant response to mycorrhizal symbiosis essentially depends on isolates spores of species, soil fertility and environmental culture conditions (Garbaye, 2013). Optical sensing systems are potential and suitable tools for assessing mycorrhization success rate since they gather information on plant response, rather than costly, time-consuming, and destructive laboratory analyses. In this field, many efforts are done with various acquisitions sensing system and several spectral features developed (Mulla, 2013; Pinheiro & Gusmo Dos Anjos., 2014). However, there is an apparent lack of study in remote sensing crop investigation with arbuscular mycorrhizae. New spectral indices that simultaneously allow assessment of multiple crops characteristic and reveal better understanding of plant molecular mechanism could be retrieved.

An incident photosynthetically active radiation (PAR) interaction with plant leaves induced various spectral response in absorbance, reflectance and transmittance. Reflectance expresses light use efficiency of plant (Barton & North, 2001). Reflectance properties of leaves are determined by the concentration of chlorophyll, other pigments (carotenoids, xanthophylls and anthocyanins) and absorbing biochemicals in the visible (400–700 nm) wavelength region, by mesophyll structure in the near infrared (700–1200 nm) region and by amount of water in the near infrared (950–970 nm) and in the middle infrared (1200–2400 nm) region (Peñuelas, 1998). When a fluorescent molecule absorbs the energy of a given wavelength, a part of it is dissipated by light emission at longer wavelengths within a very short time. This small but variable amount of energy loss is known as the fluorescence emission which is fingerprint of leaf photosynthetic activity. Coupling the measurements of reflectance and fluorescence with high spectral resolution has been suggested for improving crop productivity evaluation (Baret, Guyot, & Major, 1988), stress diagnosis (Lichtenthaler, Wenzel, & Buschmann, 2006) and nutrient diagnosis (Belanger, Viau, Samson, & Chamberland, 2005). Decades of research in vegetation reflectance and fluorescence-based methods has gone into finding multiple indices precisely related to several leaf pigments, stresses, and mineral content. Combination of fluorescence and reflectance could reveal some functional properties of AMF in crop production particularly in plantain and banana case where their effects have been studied in recent Africa's agricultural researches (Nwaga et al., 2011; Gaidashova et al., 2013; Jefwa et al., 2013).

Multi-class classification in crops remote sensing is performed by means of data mining techniques. In this approach, several methods of machine learning have been utilized for plant characteristics detection. Support Vector Machine are important advanced techniques intensively used (Mountrakis & Ogole, 2011; Moshou, Pantazi, Kateris, & Gravalos, 2014). Others efficient tools are concerned with fuzzy logic, neural network and extreme learning machine (Moreno, Corona, Lendass, Graña, & Galvão 2014; Murmu & Biswas, 2015; Cvetković, Stojanović, & Nolić, 2015). The boosting meta-algorithm is relatively new, efficient, simple, and easy to manipulate additive modeling technique that can use potentially any weak learner available. LogitBoost is another variant of the boosting algorithm that performs additive logistic regression (Friedman, Hastie, & Tibshirani, 2000) and has shown better performance than many other machine learning algorithm especially in protein data structure classification (Krishnaraj & Reddy, 2009). At present, new improvements of this algorithm are proposed (Kanamori & Takenouchi, 2013; Sun, Reid & Zhou, 2014).

In this study, we aimed at evaluating early detection of mycorrhized plantain using active fluorescence and reflectance spectrometry. The specific objectives were to (1) identify the most significant fluorescence and reflectance vegetation indices explaining mycorrhizae effects on plantains and to (2) build predictive classification model for each mycorrhizal treatment using spectral features discovered.

## 2. Material and methods

### 2.1 Plant Material and Experimental Design

The experiment was conducted under greenhouse conditions at the Laboratoire des Biotechnologies Végétale et Microbienne in National Polytechnic Houphouët-Boigny Institute of Yamoussoukro (Côte d'Ivoire). Two plantain varieties namely Orishele (sensitive to water stress) and Fhia 21 (tolerant to water stress) were cultivated in pots containing sterilized substrate composed of loam and sand. The plants were inoculated with a mycorrhizal morphotype of *Glomus* (T1), a complex of spores from the rhizosphere of plantain in Bouaflé (T2) and another complex of spores from the rhizosphere of plantain in Azaguié (T3). All treatments including control plants without any mycorrhizae (T0) were split into a randomized complete block design consisting of three replicates per treatment in each of the three blocks.

## 2.2 Formulation and Preparation of Inoculum

Spores are extracted at the Laboratoire des Biotechnologies Végétale et Microbienne in National Polytechnic Houphouët-Boigny Institute of Yamoussoukro (Côte d'Ivoire) according to the method described by Gerdemann & Nicholson (1963). They were disinfected using bleach 2% and 0.2% streptomycin. The banana suckers were firstly cleaned, washed and acclimated in coco peat and then inoculated with these spores at 150 spores per pot.

## 2.3 Optical Instrumentation and Spectral Measurements

A hand-held clip LEDs was built in our laboratory and mounted with USB4000 spectrometer Ocean Optics for acquiring fluorescence and reflectance spectra on the upper leaf surface. The spectrometer can collect data in the 350–1000 nm spectral region, with a sampling interval of 0.22 nm. Three Light-emitting-diodes excited the fluorescence at 375 nm (UV), 520 nm (green) and 630 nm (red) while the emitted fluorescence light was detected in the red (RF: 680–690 nm) and far-red (FRF: 720–755 nm) spectral regions using a low-pass filter at 650 nm. A white LED emitting (400-700 nm) was utilized for reflectance measurement. All four LEDs were controlled using the computer and Arduino, a cost effective, open source and easy programmable microcontroller board (Arduino - Home). Reflectance spectra were carried out after comparing with a 99% white reflectance standard (Labsphere®, Edmund Optics Inc. — 101 East Gloucester Pike, Barrington, NJ 080071380 USA). After three scans, an average spectrum is recorded using SpectraSuite® Software. Spectra acquisitions occurred between 29 November and 06 December 2014 with experimental setup showed in Figure 1.

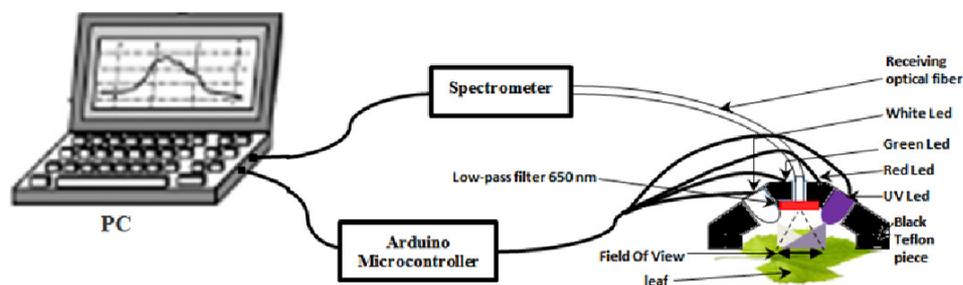


Figure 1. Schematic drawing of the experimental setup used for spectra acquisition

## 2.4 Spectra Preprocessing and Normalization

The spectra dataset recorded are firstly filtered using Gaussian normal probability density function in matlab. Then, each spectrum is standardized by its own average and standard deviation for baseline and quantity correction (Barnes, Dhanoa & Lister., 1993). Finally, in the hypothesis that the number of photon at any wavelength should be constant across shots, we proceeded to min/max normalization. Each spectrum is normalized to have a minimum value of 0 and a maximum value of 1. This normalization was accomplished according to the formula:

$$x_{norm} = \frac{(x - \min_i x_i)}{(\max_i x_i - \min_i x_i)} \quad (1)$$

## 2.5 Fluorescence and Reflectance Indices Calculation

Vegetation indices are widespread in remote sensing scientific literature. A database composed of 18 fluorescence and 90 reflectance indices were implemented in matlab for spectra features calculation. We tried to extend to almost reviewed vegetation index at leaf and canopy level for enabling maximum crop characteristic detection. These indexes are also supplemented with our proposed one.

## 2.6 Statistical Analysis

### 2.6.1 One-Way Variance Analysis with Step-Down Dunnett Post Hoc Test

ANalysis Of VAriance (often referred to as ANOVA) is a technique for analyzing the way in which the mean of a variable is affected by different types and combinations of factors. One-way analysis of variance is the simplest form in which we are interested in comparing the dependent variable means of two or more groups defined by a categorical grouping factor (Bewick, Cheek, & Ball, 2004). Analysis of variance is initially performed on two groups (control and all mycorrhized plants together) then on four groups (control and each of three mycorrhizal treatments). In order to understand subgroup differences among the different experimental and control group, we

chose to perform step-down Dunnett test which is more suitable and powerful (Bretz, Hothorn & Westfall, 2010). R software version 3.1.2 (R Core Team, 2014) was used for ANOVA analysis and for multiple comparisons.

### 2.6.2 Principal Components and Vegetation Index Correlation Analysis

Principal components analysis (PCA) is a data reduction technique that transforms a larger number of correlated variables into a much smaller set of uncorrelated variables called principal components. The weights used to form the linear composites are chosen to maximize the variance each principal component accounts for, while keeping the components uncorrelated. We generated principal components for each cultivar with their respective significant vegetation indices.

Correlation analysis was also used for establishing relationship between new indice developed and existing ones.

### 2.6.3 LogitBoost Classifier

Boosting implements forward stagewise additive modeling. This class of algorithms starts with an empty ensemble and incorporates new members sequentially. At each stage the model that maximizes the predictive performance of the ensemble as a whole is added, without altering those already in the ensemble. Additive logistic regression use similar adaptation by modifying the forward stagewise modeling method (Witten, Frank, & Hall, 2011).

For  $K$ -class classification ( $K \geq 2$ ), consider an  $N$ example training set  $\{x_i, y_i\}_{i=1}^N$  where  $x_i$  denotes a feature value and  $y_i \in \{1, \dots, K\}$  denotes a class label. Class probabilities conditioned on  $x$  denoted by  $p = (p_1, \dots, p_K)^T$  are learned from the training set. For a test example with known  $x$  and unknown  $y$ , we predict a class label by using the Bayes rule:  $y = \operatorname{argmax}_k p_k$ ,  $k = 1, \dots, K$ .

Instead of learning the class probability directly, one learns its “proxy”  $F = (F_1, \dots, F_K)^T$  given by the so-called Logit link function:

$$p_k = \frac{\exp(F_k)}{\sum_{j=1}^K \exp(F_j)} \quad (2)$$

with the constraint  $\sum_{k=1}^K F_k$  (Friedman et al., 2000). For simplicity and without confusion, we hereafter omit the dependence on  $x$  for  $F$  and for other related variables.

The  $F$  is obtained by minimizing a target function on training data:

$$\text{Loss} = \sum_{i=1}^N L(y_i, x_i), \quad (3)$$

where  $F_i$  is shorthand for  $F(x_i)$  and  $L(y_i, F_i)$  is the Logit loss for a single training example:

$$L(y_i, F_i) = -\sum_{k=1}^K r_{ik} \log p_{ik} \quad (4)$$

Where  $r_{ik} = 1$  if  $y_i = k$  and 0 otherwise. The probability  $p_{ik}$  is connected to  $F_{ik}$  via (2).

To make the optimization of (3) feasible, a model is needed to describe how  $F$  depends on  $x$ . For example, linear model  $F = W^T x$  is used in traditional Logit regression, while Generalized Additive Model is adopted in LogitBoost:

$$F(x) = \sum_{m=1}^M f_m(x), \quad (5)$$

where each  $f_m(x)$ , a  $K$  dimensional sum-to-zero vector, is learned by greedy stage-wise optimization. That is, at each iteration  $f_m(x)$  is added only based on  $F = \sum_{j=1}^{m-1} f_j$ . Formally,

$$f_m(x) = \operatorname{argmin}_f \sum_{i=1}^N L(y_i, F_i + f(x_i)), \quad (6)$$

$$s. t. \sum_k f_k(x_i) = 0, i = 1, \dots, N,$$

This procedure repeats  $M$  times with initial condition  $F = 0$ . Owing to its iterative nature, we only need to know how to solve (6) in order to implement the optimization (Sun et al., 2014).

Models were performed in WEKA 3.6.13 software Explorer interface (Hall et al., 2009). They were built separately for each cultivar using their own significant fluorescence and reflectance indices according to ANOVA results. The four output classes used in the analysis were T0, T1, T2 and T3 distributed on 72 observations in each model. In holdout evaluation, the data were randomly separated into training and test datasets such that 66% of the data was utilized for training the classifier model; while 33% of the data was used for testing the developed classifier model. In order to mitigate any bias by the particular sample chosen for holdout, we performed a tenfold stratified cross-validation. The outcomes metrics are automatically generated in WEKA. Two-class outcomes are presented in figure 2 for interpretation needs.

		Predicted Class	
		yes	no
Actual Class	yes	true positive (TP)	false negative (FN)
	no	false positive (FP)	true negative (TN)

$$\text{TP rate(\%)} = \frac{\text{TP}}{\text{TP} + \text{FN}} \times 100$$

$$\text{FP rate(\%)} = \frac{\text{FP}}{\text{FP} + \text{TN}} \times 100$$

$$\text{Precision(\%)} = \frac{\text{TP}}{\text{TP} + \text{FP}} \times 100$$

$$\text{Recall(\%)} = \frac{\text{TP}}{\text{TP} + \text{FN}} \times 100$$

Figure 2. Confusion matrix and associated classification measures in two-class case

### 3. Results and Discussion

#### 3.1 Data Representation after Preprocessing and Normalization

Fluorescence spectra using UV (375 nm), Green (520 nm) and Red (630 nm) essentially show two peaks in red band (685- 690nm) and far –red band (735-740 nm) known as chlorophyll a fluorescence (Buschmann Langsdorf & Lichtenhaler, 2008; Misra & Singh, 2012). Intensity at these two bands vary when using an UV (375 nm) or Green (520 nm) light because of respective absorption at these wavelengths of flavonols (Cerovic *et al.*, 2002) and anthocyanins pigments (Agati *et al.*, 2005). Analysis of reflectance curve presents a deep through in 850-1000 nm band and two others at 778-788 nm and 834-840 nm bands. These observations may be attributed to water absorption.

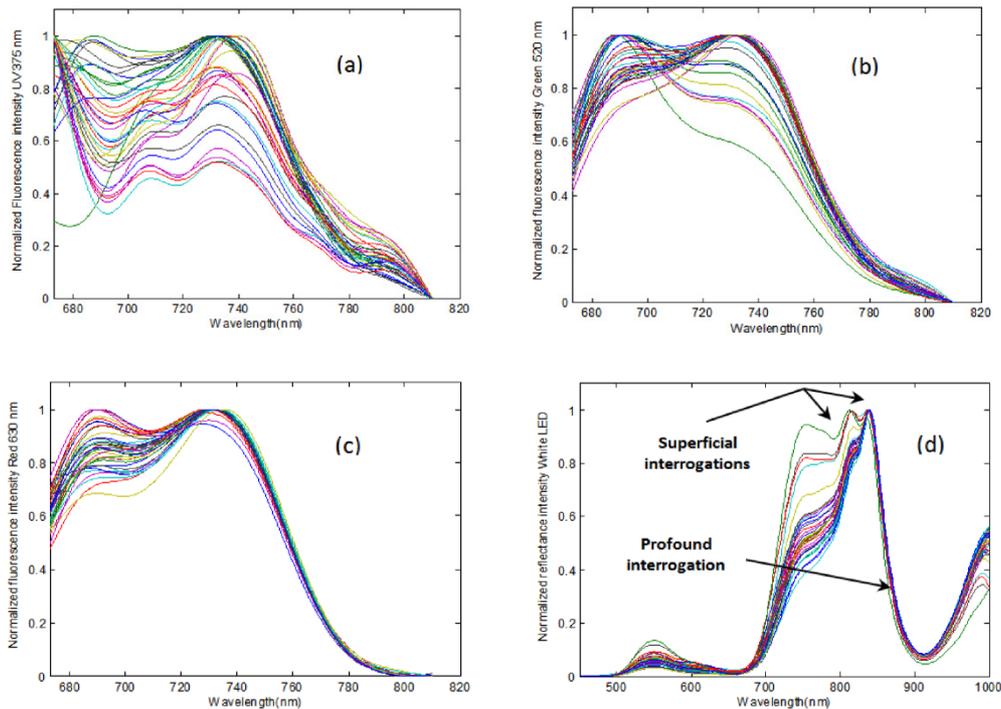


Figure 3. Normalized fluorescence spectra using excitation Led source at 375 nm (a), excitation Led source at 520 nm (b), excitation Led source at 630 nm (c) and normalized reflectance spectra using white light Led source 400-700 nm (d) in experimental setup

### 3.2 Spectral Responses to Plantain Mycorrhizal Treatment

Analysis of variance of 108 vegetation indexes reveals 3 significant fluorescence indices and 12 significant indices for reflectance spectra. ANOVA results in Table 1 and 2 show that differences between control and each of three mycorrhizal treatments are revealed by reflectance indexes. Two reflectance indexes are highly significant ( $P < 0.001$ ): Datt5 for Fhia 21 cultivar and the new index NWI5 for Orishele cultivar.

Table 1. Effect of AM fungal inoculation on calculated vegetation indexes of plantain cultivar Fhia 21 under greenhouse conditions

ANOVA	Vegetation Index											
	Fluorescence		Reflectance									
	ANTH	FRF_R	Carter5	Datt5	SR5	GI	NDVI3	NWI	NWI4	NWI5	dNIRMin 920_980	min950_970
All mycorrhized treatment	*	*	NS	**	NS	**	**	NS	NS	*	*	*
Three level mycorrhized treatment	NS	NS	*	***	*	**	**	*	*	**	NS	*

Significance code: \* Significant at 0.05, \*\* significant at 0.01, \*\*\* significant at 0.001, NS Not significant

Table 2. Effect of AM fungal inoculation on calculated vegetation indexes of plantain cultivar Orishele under greenhouse conditions

ANOVA	Vegetation Indexes							
	Fluorescence	Reflectance						
	NBI_R	DPI	RatiodRE_703	NWI	NWI5	dNIRmin920_980	min950_970	
All mycorrhized treatment	*	*	NS	*	**	**	*	
Three level mycorrhized treatment	NS	*	*	*	***	*	*	

Significance code: \* Significant at 0.05, \*\* significant at 0.01, \*\*\* significant at 0.001, NS Not significant.

Table 3 gives us an overview on their definition and relationship with bioparameters. From most to least importance order, these spectral features are related to water content (NWI, NWI5, NWI4, dNIRMin920\_980, min950\_970), chlorophyll content (Datt5, NDVI3, SR5, and RatiodRE\_703), anthocyanin content (ANTH), foliar density (FRF\_R), nitrogen content (NBI\_R), photosynthetic activity (DPI) and leaf area index (GI).

As expected, most of these spectral features appear to be relevant in plantain mycorrhization studies. On the one hand, we can see that indices linked to water content appear with great occurrence in both varieties: NWI, NWI4, NWI5, dNIRmin920\_980, min950\_970. These water-based indexes demonstrate that mycorrhized plantains have better water content in comparison to control. This result agrees with Nwaga et al. (2011) study which has shown good correlation between water use efficiency and drought tolerance of mycorrhized plantains. As an example, Orishele water content indices given in Figure 4 show that water-based indexes means for all mycorrhized classes are higher than control ones.

Table 3. Overview of significant fluorescence and reflectance indexes in Fhia 21 and Orishele plantain cultivars

Vegetation index	Related to	Equation	Reference
<i>Fluorescence indexes</i>			
Anthocyanin (ANTH)	Anthocyanin content	$\log(FRF\_R/FRF\_G)$	Agati et al. (2005)
Far-Red Fluorescence with Red light (FRF_R)	Foliar density, leaf strength	$FRF\_R$	Bramley et al. (2011)
Red Nitrogen Balance Index (NBI_R)	Nitrogen content	$\log(FRF\_UV/FRF\_R)$	Cartelat et al. (2005)
<i>Reflectance indexes</i>			
SR5	Chlorophyll a	$R_{675}/R_{700}$	Chappelle, Kim & McMurtrey
Datt5	Chlorophyll b	$R_{672}/R_{550}$	Datt (1998)
Normalized Difference Vegetation Index (NDVI3)	Chlorophyll total	$(R_{682} - R_{553}) / (R_{682} + R_{553})$	Gandia, Fernandez, Garcia & Moreno (2004)
Ratio of first derivative maximum value in red-edge with reflectance one at 703 nm (RatiodRE_703)	Chlorophyll, Water content	$maxD_{680-780}/D_{703}$	Filella & Peñuelas (1994)
Greenness Index (GI)	Chlorophyll, Leaf Area Index	$R_{554}/R_{677}$	Smith, Adams, Stephen & Hick (1995)
Carter5	Stress	$R_{695}/R_{670}$	Carter (1994)
Double Peak Index (DPI)	Photosynthetic activity	$(D_{688} \times D_{710}) / D_{697}^2$	Zarco-Tejada, Pushnik, Dobrowski & Ustin (2003)
Normalized Water Index (NWI)	Water content	$(R_{970} - R_{900}) / (R_{970} + R_{900})$	Barbar et al. (2006)
Normalized Water Index 4 (NWI4)	Water content	$(R_{970} - R_{920}) / (R_{970} + R_{920})$	Prasad et al. (2007)
Normalized Water Index 5 (NWI5)	Water content	$\frac{(minR_{778-788} - minR_{834-840})}{(minR_{778-788} + minR_{834-840})}$	In this study
Minimum value of the first derivative in 920-980 range (dNIRMin920_980)	Water content	$minD_{920-980}$	Peñuelas, Fillela, Biel, Serrano & Salvé (1993)
Minimum value in 950-970 range (min950_970)	Water content	$minR_{950-970}$	Peñuelas, Fillela, Biel, Serrano & Salvé (1993)

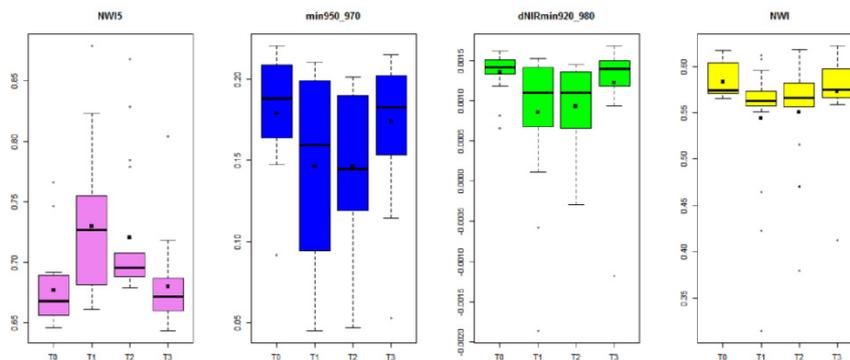


Figure 4. Boxplots of water-based vegetation indices (NW15, min950\_970, dNIRmin920\_980 and NWI) for each treatment group for Orishele variety. Means are shown by fat dots, boxes indicate median and interquartile range, with vertical lines depicting the range. Dots outside vertical range are outliers' data

As suggested from the reflectance curve analysis, we computed a new normalized water index NW15. This index shows significant differences between treatments for both varieties and is strongly correlated with leaf water content as it appears in correlation matrices (Table 4 and 5) particularly with water index content dNIRmin920\_980 and min950\_970 ( $-0.84 < r < -0.88$ ). Therefore this parameter may stand as an indicator of water status. The negative correlation with the two literature indexes mentioned above suggests that water content in leaf is high when NW5 is high. Indeed dNIRmin920\_980 and min950\_970 are negatively correlated with relative water content. In addition, NW15 presents more significance in Orishele variety than in Fhia 21 because Orishele is more sensitive to water stress. As a result, NW15 could better track plant water stress sensitivity.

Table 4. Pearson correlation coefficients (all with p value < 0.01) for water content reflectance indexes of Fhia 21 variety

Reflectance indexes	dNIRmin920_980	min950_970	NWI4	NWI5	NWI
dNIRmin920_980	-				
min950_970	0.842	-			
NWI4	0.572	0.508	-		
NWI5	-0.843	-0.879	-0.511	-	
NWI	0.445	0.358	-0.860	-0.452	-

Table 5. Pearson correlation coefficients (all with p value < 0.01) for water content reflectance indexes of Orishele variety

Reflectance indexes	dNIRmin920_980	min950_970	NWI5	NWI
dNIRmin920_980	-			
min950_970	0.826	-		
NWI5	-0.859	-0.838	-	
NWI	0.780	0.559	-0.754	-

On the other hand, related works (Tropical Soil Biology and Fertility [TSBF], 2007; Declerck, Cevos, Devos, & Plenchette, 1994) have reported that AMF improves nutrient uptake and growth of bananas particularly an enhancement of P uptake. In this context, our study provides additive information especially about mycorrhization effects on plantain nutrition, leaf pigment content and photosynthetic activity. Indeed, in variety Fhia 21, ANTH index which is negatively correlated with anthocyanin content as demonstrated by J. Baluja (2012), indicates that all mycorrhizal treatment induces an increase of anthocyanins. FRF\_R index is related to

foliar density. Bramley et al. (2011) showed that NDVI provided by GreenSeeker has a positive relationship with index FRF\_R obtained using the Multiplex long distance to flowering when the vegetation is less developed. So we deduce low rate of chlorophyll content in Fhia 21 variety for mycorrhizal plants. This is also illustrated in figure 5 by chlorophyll index (Datt5, SR5, NDVI3 and GI) and is due to higher anthocyanin content.

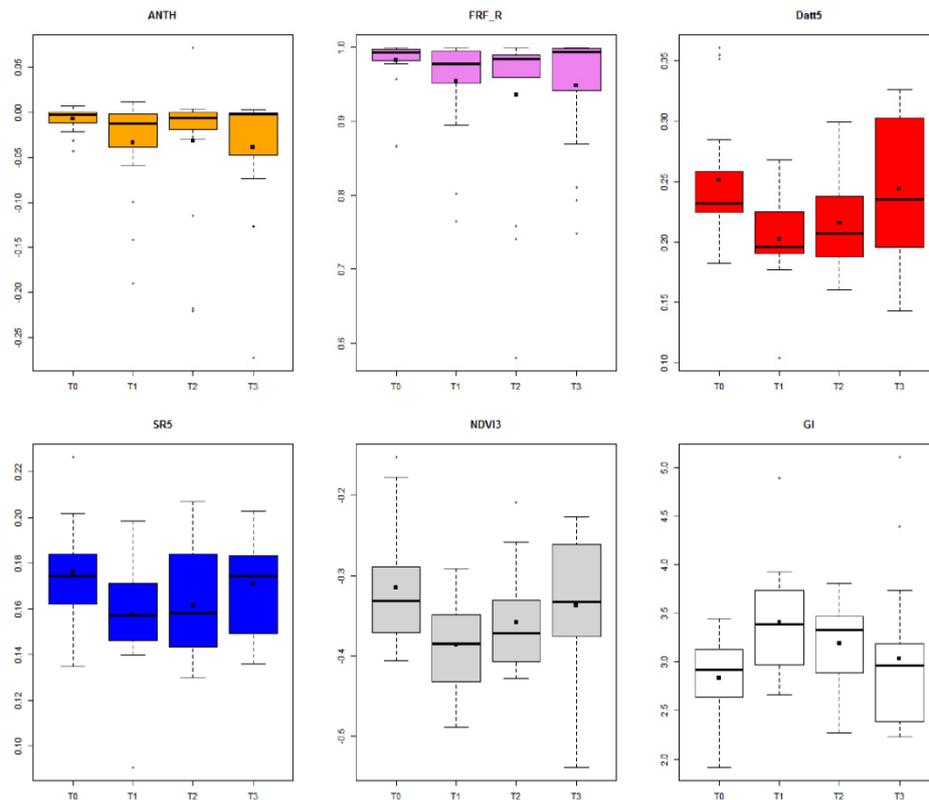


Figure 5. Boxplots of vegetation indices (ANTH, FRF\_R, Datt5, SR5, NDVI3 and GI) for each treatment group for Orishele variety. Means are shown by fat dots; boxes indicate median and interquartile range, with vertical lines depicting the range. Dots outside vertical range are outliers' data

In Orishele variety, improved effects of mycorrhizae on chlorophyll content and photosynthetic activity appear and are respectively shown by higher value of red-edge spectral feature (RatiodRE\_703) and double peak indice (DPI) in T1 and T2 mycorrhizal treatments (Figure 6). Experience conducted by Zarco-Tejada et al (2003) have demonstrated that natural fluorescence emission is observable on the first derivative reflectance spectra as a double-peak feature in the 690–710 nm spectral region. DPI is capable for tracking natural steady-state fluorescence and is closely related to photosynthetic activity so that DPI is low when photosynthetic rates are high. Nethertheless, NBI\_R boxplot means show that plants with inoculated mycorrhizae have low nitrogen content even if we can observe highest value of this index (1.3-1.51) for T1 treatment. NBI\_R which is expressed by the ratio between epidermal phenolic compounds and chlorophyll is positively well correlated to the leaf N content (Agati, Foshi, Grossi & Volterrani, 2015). Figure 6 illustrates distribution of vegetation indices (NBI\_R, RatiodRE\_703 and DPI) according to mycorrhizal treatments for Orishele variety.

Step-down Dunnett test helps us identifying the most effective mycorrhizal treatments in comparison to control. From multiple comparison analysis, only significant vegetation index with four groups in ANOVA are concerned. Results in Table 6 show clearly that for Fhia 21 cultivar treatment T1 is the best and then treatment T2 follows concerning with improvement of water content performance. Reflectance indexes in Table 7 indicate that treatment T1 and T2 are efficient with few precision about the first one and the second one according to step-down Dunnett statistical test. In both varieties, following indexes namely Carter5, GI, RatiodRE\_703 and NW15 were not able to discriminate mycorrhizal treatments with this test. In addition, treatment T3 is not at all significant.

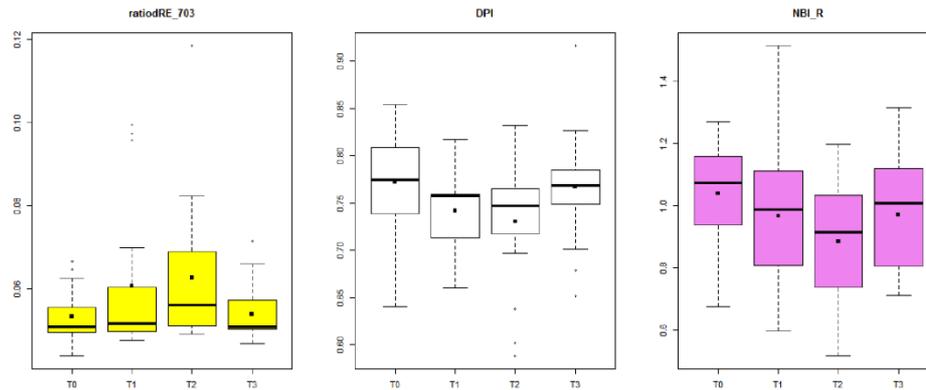


Figure 6. Boxplots of vegetation indices for each treatment group for Orishele variety (RatiodRE\_703, NBI\_R, and DPI). Means are shown by fat dots; boxes indicate median and interquartile range, with vertical lines depicting the range. Dots outside vertical range are outliers' data

Table 6. Multiple comparisons of means to a control of plantain cultivar Fhia 21 under greenhouse condition

Mycorrhizal Treatment vs control	Step-down Dunnett statistical significance								
	Carter5	Datt5	SR5	GI	NDVI3	NWI	NWI4	NWI5	min950_970
T1-T0	NS	***	**	NS	***	*	**	NS	**
T2-T0	NS	**	*	NS	*	NS	NS	NS	*
T3-T0	NS	NS	NS	NS	NS	NS	NS	NS	NS

Significance code: \* Significant at 0.05, \*\* significant at 0.01, \*\*\* significant at 0.001, NS Not significant.

Table 7 Multiple comparisons of means to a control of plantain cultivar Orishele under greenhouse conditions

Mycorrhizal Treatment vs control	Step-down Dunnett statistical significance					
	DPI	RatiodRE_703	NWI	NWI5	dNIRmin920_980	min950_970
T1-T0	*	NS	**	NS	**	**
T2-T0	**	NS	*	NS	**	**
T3-T0	NS	NS	NS	NS	NS	NS

Significance code: \* Significant at 0.05, \*\* significant at 0.01, NS Not significant.

Analysis of mycorrhizal effects on Fhia 21 and Orishele plantain cultivars leads us discovering a new water index as well as others relevant and varied fluorescence and reflectance indices. Spectral responses analysis of plants show essentially improved effects of mycorrhizae in water content for both varieties. Good effects are also revealed in anthocyanin, chlorophyll, photosynthetic activity and nitrogen content whose treatments T1 and T2 are mostly responsible for.

### 3.3 LogitBoost Classification Evaluation

In addition to finding most significant vegetation indexes and efficient mycorrhization treatment, additive logistic regression models are constructed to automatically predict specific class of a of mycorrhized plantain. Principal components used as variables input of models explains 99.47% of spectral features variability for Fhia 21 variety and 94.163 % in Orishele one. The PC1 and PC2 extracted allow this as shown in Table 8.

Cross-validation gives us a first insight in classification performance. On total instances of 72 samples, 90.2778 % are correctly classified while 9.7222 % are misclassified for both varieties (Table 10). Detailed performance metrics are given in Table 9. Precisely, classification accuracy is higher for treatment without any mycorrhizae T0 (94.4 %) than those with (88.9 %) in overall variety. In the mycorrhized classes treatment T3 has the best precision parameter (94.1 %) because of its low FP Rate. In addition, we can explain higher value of recall in

control class in such way that the FN samples are low. It is also notable to precise that mycorrhized treatment T1 and T2 present the same performance metrics.

Table 8. Principal components results

Variety	Principal component	Variance explained (%)
Fhia 21	PC1	89.44
	PC1, PC2	99.47
Orishele	PC1	80.04
	PC1, PC2	94.163

Table 9. Classification performance metrics of the LogitBoost classifier by class

Variety	Evaluation method	Class	TP Rate	FP Rate	Precision	Recall
Fhia 21	Stratified cross-validation	T0	94.4	3.70	89.5	94.4
		T1	88.9	3.70	88.9	88.9
		T2	88.9	3.70	88.9	88.9
		T3	88.9	1.90	94.1	88.9
	Holdout	T0	100	11.1	75.0	100
		T1	60.0	0.00	100	60.0
		T2	100	5.30	83.3	100
		T3	87.5	0.00	100	87.5
Orishele	Stratified cross-validation	T0	94.4	3.70	89.5	94.4
		T1	88.9	3.70	88.9	88.9
		T2	88.9	3.70	88.9	88.9
		T3	88.9	1.90	94.1	88.9
	Holdout	T0	100	11.1	75.0	100
		T1	60.0	0.00	100	60.0
		T2	100	5.30	83.3	100
		T3	87.5	0.00	100	87.5

Table 10. General performance of the LogitBoost classifier

Variety	Evaluation method	Correctly classified (%)	Incorrectly classified (%)
Fhia 21	Stratified cross-validation	90.2778	9.7222
	Holdout	87.5	12.5
Orishele	Stratified cross-validation	90.2778	9.7222
	Holdout	87.5	12.5

Predictions by treatment with holdout method display two tendencies for both varieties (Table 10). Treatment T0 and T2 are highly accurate with no FN samples (100 %) while T1 and T3 treatments accuracy are low respectively 60 % and 87.5 % with no FP rate.

From models analysis, we can assess that general predictive classification in cross-validation results are good and reflect prediction on each treatment but in testing the models, treatment T1 is overestimated. Anyway, models developed for each cultivar appear acceptable and greatly encouraging even if 10 times tenfold cross-validation should be performed for reliable results and accurate error estimation.

#### 4. Conclusion

In this article, we focus on feasibility for remotely detecting at early stage two plantain varieties inoculated with different Arbuscular Mycorrhizal Fungi. Our results firstly emphasize a new water content indice NWI5 highly significant with Datt5 index at  $P < 0.001$ . New findings also concern 3 fluorescence features (ANTH, FRF\_R, and NBI\_R) related to anthocyanin, foliar density and nitrogen content, and 10 others in reflectance spectra (SR5, NDVI3, radiodRE\_703, NWI, NWI4, dNIRmin920\_980, min950\_970, Carter5, DPI, GI) linked to chlorophyll content, water content, photosynthetic activity and leaf area index. Secondly, we derive predictive multiclass classification models for each cultivar with around 90.3 % and 87.5 % accuracy respectively in stratified cross-validation and holdout evaluation. This demonstrates capability of our fluorescence and reflectance-based instrumentation for quantitatively investigating crop growth under mycorrhizae fertilization. Effective vegetation indices identification in crops specialty as well as rapid pattern recognition of mycorrhizal treatment represent advanced key for developing non-destructive and cost-effective crops monitoring detectors which will more contribute to sustainability in agriculture. Furthermore, we might expect better results in this study using improved version of the LogitBoost classifier. Elaborating a unique model for both plantain varieties could also represent another challenge.

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